



**MONASH** University

**NEUROPROTECTIVE EFFECT OF EMBELIN IN EXPERIMENTAL  
MODELS OF ALZHEIMER'S DISEASE**

By

**B SAATHEEYAVAANE BHUVANENDRAN PILLAI**

**Bachelor of Science (Hons)**

A thesis submitted for the degree of Doctor of Philosophy at  
Monash University in 2018

Jeffrey Cheah School of Medicine and Health Sciences,  
Monash University Malaysia

## **Supervisors**

### **Principal Supervisor:**

Dr. Mohd. Farooq Shaikh  
Senior Lecturer in Pharmacology  
Neuropharmacology Research Laboratory,  
Jeffrey Cheah School of Medicine and Health Sciences,  
Monash University Malaysia,  
Jalan Lagoon Selatan, Bandar Sunway 47500, Selangor, Malaysia

### **Associate Supervisors:**

Dr. Yatinesh Kumari  
Senior Lecturer  
Neuropharmacology Research Laboratory,  
Jeffrey Cheah School of Medicine and Health Sciences,  
Monash University Malaysia,  
Jalan Lagoon Selatan, Bandar Sunway 47500, Selangor, Malaysia

Professor Iekhsan Othman  
Head of Biomedical Sciences  
Jeffrey Cheah School of Medicine and Health Sciences,  
Monash University Malaysia,  
Jalan Lagoon Selatan, Bandar Sunway 47500, Selangor, Malaysia

## **Copyright notice**

### *Notice 1*

© B SAATHEEYAVAANE BHUVANENDRAN PILLAI (2018).

I certify that I have made all reasonable efforts to secure copyright permissions for third-party content included in this thesis and have not knowingly added copyright content to my work without the owner's permission.

## TABLE OF CONTENTS

Copyright notice.....	III
Table of Contents .....	IV
Abstract.....	VI
Declaration.....	VIII
Thesis including published works declaration.....	IX
Other PhD related publications, presentations and award during the PhD period.....	XII
Acknowledgments.....	XIV
Chapter 1: Introduction.....	1
Chapter 2: Research Questions .....	5
Chapter 3: Literature Review .....	7
3.1 Introduction	
3.2 Pathology of AD	
3.2.1 Amyloid hypothesis	
3.2.2 Tau hypothesis	
3.2.3 Cholinergic hypothesis	
3.2.4 Vascular hypothesis	
3.3 Limitations of current therapy	
3.4 Role of embelin in CNS related disorders	
3.5 Publication: Plant Derived Phytochemical, Embelin in CNS Disorder: A Systematic Review	
Chapter 4.....	25
4.1 Introduction	
4.2 Publication: Amelioration of cognitive deficit by Embelin in Scopolamine-Induced Alzheimer’s Disease-Like Condition in a Rat Model	
Chapter 5.....	39
5.1 Introduction	
5.2 Publication: Embelin Improves the Spatial Memory and Hippocampal Long-Term Potentiation in a Rat Model of Chronic Cerebral Hypoperfusion	
Chapter 6.....	62

6.1 Introduction	
6.2 Publication: Embelin prevents amyloid beta accumulation via GSK-3 pathway in an in vitro model of streptozotocin-induced AD like condition	
Chapter 7.....	<b>90</b>
7.1 Introduction	
7.2 Publication: Embelin, a potent molecule for Alzheimer’s disease: A proof of concept from BBB permeability, AChE inhibition and molecular docking studies	
Chapter 8.....	<b>108</b>
8.1 Strengths and limitations	
8.2 Future work	
8.3 Conclusion	
References.....	<b>112</b>

## Abstract

Alzheimer's disease (AD) is a severe progressive neurodegenerative brain disorder which displays global cognitive decline involving memory, orientation, judgment, and reasoning. It affects millions of people and it is estimated that by 2050, 1 in 85 individuals worldwide will be living with the disease. Embelin (2, 5dihydroxy-3-undecyl-1,4 benzoquinone) is the major active constituent in the fruits of *Embelia ribes* Burm. The first part of our thesis results revealed that embelin is a potent acetylcholinesterase (AChE) inhibitor when compared with donepezil. Besides that, both nootropic and anti-amnesic effects of embelin were evaluated using the scopolamine model of amnesia in rats. These research findings suggested that embelin is a potent nootropic and anti-amnesic phytochemical. When tested in experimental models of amnesia, embelin significantly improved recognition index and memory retention in both novel object recognition (NOR) and elevated plus maze (EPM) tests. The hippocampal tissues from scopolamine-induced amnesia model were extracted for gene expression, neurotransmitter, and immunocytochemistry studies. The results revealed that embelin elevated the mRNA expression of brain-derived neurotrophic factor (BDNF), cAMP response element-binding protein (CREB1), and scavenger enzymes superoxide dismutase 1 (SOD1) and catalase (CAT). Rats pre-treated with embelin could mitigate scopolamine-induced neurochemical and histological changes in a manner comparable to donepezil. The second part of this research investigated the neuroprotective effect of embelin in permanent bilateral common carotid artery occlusion (PBOCCA) model of AD. PBOCCA and embelin (0.3, 0.6, and 1.2mg/kg) treated rats were subjected to behavioural analysis to assess learning and memory functions. The hippocampal tissues from PBOCCA model were extracted for gene expression and neurotransmitter studies. Overall, embelin improved cognitive dysfunction in an animal model of chronic cerebral hypoperfusion. We reported downregulation of amyloid precursor protein (APP) and microtubule-associated protein tau (MAPT) mRNA expression, upregulation of synaptic plasticity-related genes, reduced oxidative stress and the inflammatory response which might have contributed to the neuroprotective effects of embelin. In the third part of our thesis, we investigated whether embelin would confer protection against streptozotocin (STZ) mediated neuronal damage in rat primary hippocampal neuronal cells. In this study, embelin demonstrated protective effect against STZ induced neurotoxicity. Pre-treatment with embelin reduced the mRNA expression levels of APP, MAPT, glycogen synthase kinase 3 alpha (GSK-3 $\alpha$ ) and GSK-

$3\beta$  mRNA indicating that this compound could revert the STZ-induced insulin signalling (IR) dysfunction. Notably, embelin pre-treatment increases SOD1 and reduces nuclear factor kappa B (NF- $\kappa$ B) mRNA levels, which is responsible for oxidative stress and neuroinflammation respectively. Finally, we found that embelin inhibited STZ-induced amyloid beta protein expression. In addition, the results from porcine brain endothelial cells permeability assay indicated that embelin could cross the blood-brain barrier. Using molecular docking, we could predict that embelin has favourable binding mode within the AChE and A $\beta$  peptide active sites. Based on the studies so far, it is postulated that embelin could be a promising molecule as a memory enhancer and neuroprotective in Alzheimer's disease condition. Embelin exerts its effect via cholinergic pathway, amyloid cascade and vascular pathway by protecting the neurons against AD like condition.

## **Declaration**

This thesis contains no material which has been accepted for the award of any other degree or diploma at any university or equivalent institution and that, to the best of my knowledge and belief, this thesis contains no material previously published or written by another person except where due reference is made in the text of the thesis.

Signature: 

Print Name: B SAATHEEYAVAANE BHUVANENDRAN PILLAI

Date: 18<sup>th</sup> November 2018

## Thesis including published works declaration

I hereby declare that this thesis contains no material which has been accepted for the award of any other degree or diploma at any university or equivalent institution and that, to the best of my knowledge and belief, this thesis contains no material previously published or written by another person, except where due reference is made in the text of the thesis.

This thesis includes (1) original paper published in peer-reviewed journal and (3) currently submitted for publications. The core theme of the thesis is neuroprotective effect of embelin in experimental models of Alzheimer's disease. The ideas, development and writing up of all the papers in the thesis were the principal responsibility of myself, the student, working within the Jeffrey Cheah School of Medicine and health Sciences under the supervision of Dr. Mohd Farooq Shaikh.

The inclusion of co-authors reflects the fact that the work came from active collaboration between researchers and acknowledges input into team-based research.

In the case of chapters 3-7, my contribution to the work involved the following:

Thesis Chapter	Publication Title	Status (published, in press, accepted or returned for revision)	Nature and % of student contribution	Co-author name(s) Nature and % of Co-author's contribution*	Co-author(s), Monash student Y/N*
4	Amelioration of Cognitive Deficit by Embelin in a Scopolamine-Induced Alzheimer's Disease-Like Condition in a Rat Model	Published	Performed all the experiments, data collection, analysis and writing the manuscript in its entirety. 60%	1) Yatinesh Kumari, input into manuscript 10% 2) Iekhsan Othman, input into manuscript. 10% 3) Mohd Farooq Shaikh, critical revision of the manuscript for important intellectual content. 20%	No
5	Embelin Improves the Spatial Memory and Hippocampal Long-Term 1	Submitted	Performed experiments, data collection, analysis and drafting the manuscript. 50%	1) Siti Najmi Syuhadaa Bakar, performed Morris water maze. 5% 2) Yatinesh Kumari, input into manuscript 5% 3) Iekhsan Othman, input	No

	Potential in a Rat Model of Chronic Cerebral Hypoperfusion			into manuscript. 5% 3) Mohd Farooq Shaikh, critical revision of the manuscript for important intellectual content. 15% 4) Zurina Hasaan, performed LTP and critical revision of the manuscript for important intellectual content. 20%	
6	Embelin prevents amyloid beta accumulation via GSK-3 pathway in an in vitro model of streptozotocin-induced AD like condition	Submitted	Performed all the experiments, data collection, analysis and writing the manuscript in its entirety. 60%	1) Yatinesh Kumari, input into manuscript 10% 2) Iekhsan Othman, input into manuscript. 10% 3) Mohd Farooq Shaikh, critical revision of the manuscript for important intellectual content. 20%	No
7	Embelin, a potent molecule for Alzheimer's disease: A proof of concept from BBB permeability, Ache inhibition and molecular docking studies	Submitted	Performed experiments, data collection, analysis and drafting the manuscript. 50%	1) Nur Aziah Hanapi, performed BBB permeability assay. 10% 2) Nafees Ahmed, docking studies and input into manuscript 10% 3) Iekhsan Othman, input into manuscript. 5% 3) Mohd Farooq Shaikh, critical revision of the manuscript for important intellectual content. 10% 4) Siti Rafidah Yusof, critical revision of the manuscript for important intellectual content. 15%	No

I have renumbered sections of submitted or published papers in order to generate a consistent presentation within the thesis.

**Student signature:** 

**Date:** 18<sup>th</sup> December 2018

The undersigned hereby certify that the above declaration correctly reflects the nature and extent of the student's and co-authors' contributions to this work. In instances where I am not the

responsible author I have consulted with the responsible author to agree on the respective contributions of the authors.

**Main Supervisor signature:**

A handwritten signature in red ink, appearing to be 'S. J. ...', written over a horizontal line.

**Date:** 18<sup>th</sup> December 2018

### **Other PhD-related publications during the PhD period**

1. Uday Kundap, **Saatheeyavaane Bhuvanendran**, Yatinesh Kumari, Iekshan Othman and Mohd Farooq Shaikh. Plant derived phyto compound, Embelin in CNS disorders: A systematic review. *Front. Pharmacol.* 2017; 8(76). doi: 10.3389/fphar.2017.00076
2. Alina Arulsamy, Yogini S Jaiswal, Bey Hing Goh, **Saatheeyavaane Bhuvanendran**, Thaarvena Retinasamy, Yatinesh Kumari, Iekhsan Othman, Leonard L Williams, Mohd Farooq Shaikh. Neuroactive drugs—A perspective on drugs of synthetic and medicinal plants origin. *Pharm Pharmacol Int J.* 2018; 6(6):422–430.

### **PhD-related presentations during PhD period**

1. **Poster:** Pharmacological evaluation of phyto compound Embelin, derived from *Embelia ribes* in Alzheimer's disease using chronic cerebral hyperfusion rats. **Saatheeyavaane Bhuvanendran**, Tharveena Retinasamy, Mohd Farooq Shaikh, Yatinesh Kumari, Iekshan Othman, Zurina Hassan. CDR Mini Symposium on Drug Addiction and Neurodegeneration, USM, Penang November 2015.
2. **Poster:** Memory enhancing effect of embelin against scopolamine-induced amnesia in Sprague Dawley rats. **Saatheeyavaane Bhuvanendran**, Yatinesh Kumari, Iekshan Othman, Mohd Farooq Shaikh. 14th Meeting of Asian-Pacific Society for Neurochemistry (APSN 2016), Kuala Lumpur, Malaysia.
3. **Poster:** Neuroprotective effect of Embelin in primary rat hippocampal culture model of neurotoxicity. **Saatheeyavaane Bhuvanendran**, Yatinesh Kumari, Iekshan Othman, Mohd Farooq Shaikh. EMBO Workshop on Neural Development, 02-06 March 2018, Taipei, Taiwan

### **Award during the PhD period**

1. Received **travel grant award** and selected as APSN-ISN Neuroscience School 2016 delegate for APSN-ISN School 2016, UPM, Malaysia (Basic Techniques in in Vitro Neural Differentiation from Stem Cells).

2. Received **travel grant award** (NTD 17 500) for EMBO Workshop on Neural Development, 02-06 March 2018, Taipei, Taiwan

3. **Bronze winner:** NP-A as a memory enhancer and potential molecule for neurodegenerative diseases. Mohd Farooq Shaikh, Saatheeyavaane Bhuvanendran, Yatinesh Kumari, Iekshan Othman, Nafees Ahemad Mahammed Yunus. International Conference and Exhibition on Inventions by Institutions of Higher Learning (PECIPTA 2017), 7-9 October 2017, Kuala Terengganu, Malaysia

## **Acknowledgements**

Upon the production of this dissertation I would like to express my gratitude and sincere appreciation to my supervisor, Dr. Mohd. Farooq Shaikh, who gives me support, invaluable guidance, advice and patience throughout the entire progress of this project. His dedication and critical reading are highly appreciated. My thanks also go to my co-supervisors, Prof Iekhsan Othman and Dr Yatinesh Kumari for their constructive suggestions and unsparing assistance. I also would like to express my deepest appreciation to my collaborators Dr. Zurina Hassan and Dr. Siti Rafidah Yusof from Universiti Sains Malaysia (USM) for the kindness of providing lab facilities and experimental support for this study.

I also would like to take this opportunity to acknowledge the intellectual and conducive learning environment created at the Monash University Malaysia. In particular, I appreciate the cooperation and spirit of sharing expertise, equipments as well as facilities that exist among the staffs or students from Jeffery Cheah School of Medicine and Health Sciences. Besides that, the financial supports and fellowship received from Monash University Malaysia are indispensable in conducting this study.

I am also delighted to offer my warmest appreciation to my Neuropharmacology Research Laboratory labmates; Uday Kundap, Vanessa Lee, Brandon Choo, Atiqah Abdullah, Thaarvena, Alexis, Yamnath and all the staffs in the research laboratory for their unfailing help, immeasurable support, assistance and encouragement they had given me throughout the research. My sincere thanks also go to administrative staff, especially Kong Li San, Linda Patricia and Lee Ching Teng for their valuable supports.

It is with great honor and appreciation that I would like to give accolade to my husband Aravind Raja who never failed to amaze me with his support, encouragement and help during my hard and easy times. Last but not least, I thank my family and dearest friends for the love, concern and support they had showered on me whenever I needed them. I would say this splendid course of research could not have been completed without all these people. May they be well and happy all the time. Thank you.

# **Chapter 1**

## 1.0 Introduction

Alzheimer's disease (AD) is a permanent and progressive neurodegenerative brain disorder of elderly humans with loss of cognitive functioning (1). The term AD was first coined by German psychiatrist, Alois Alzheimer's following the admission of a 50 year old woman with progressive memory loss, sleep disturbance, aggression, and confusion (2). In AD patients, these symptoms occur due to damage of neurons in the brain part involved in cognitive function and motor coordination (3). The patient died after 5 years and from her brain autopsies, Dr. Alzheimer detected the presence of distinctive deposits, which are now known as amyloid plaques and neurofibrillary tangles (4, 5).

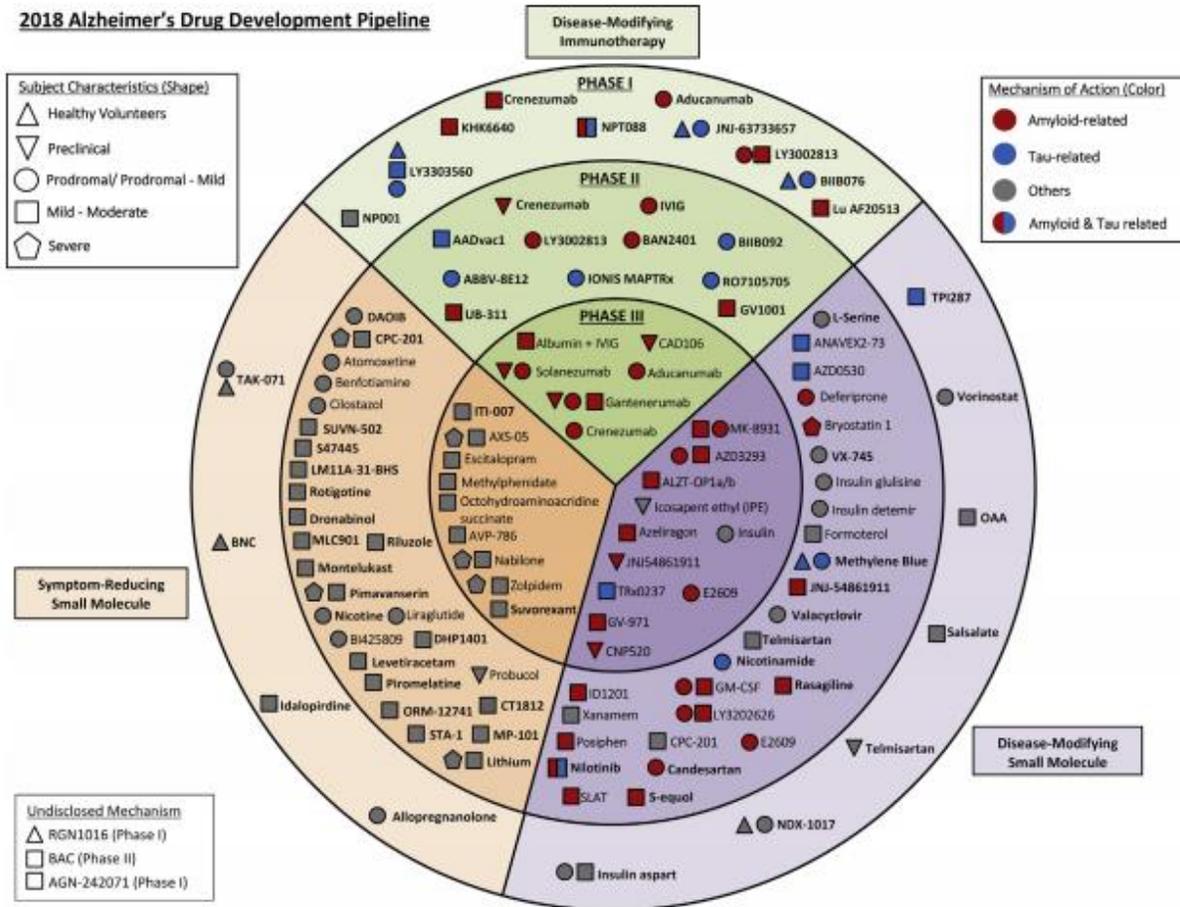
Currently about 50 million people are living with dementia globally and this figure will be more than triple by 2050 (6). Furthermore, 60 percent to 80 percent of the cases involve solely Alzheimer's pathology (3). World health organization (WHO) reported that dementia is the seventh leading cause of death. On the other hand, 4.9 million cases of dementia were recorded in Asian continents (7). This shows that Asia comprised of low-income and middle-income countries will face financial burden (8) as their economic sources are not enough for long-term care of AD patients by family or at nursing homes. In short, it can be said that AD is one of the major public health problems in the world. In Malaysia, the prevalence of dementia is estimated at 0.126% in 2020 and will increase to 0.454% by 2050 (9-11). The mentioned low prevalence rate is due to the fact that family members perceive AD and other related dementia as a normal part of aging and therefore the patients are missed to be diagnosed (9).

AD is typically categorized into three general stages known as mild (early stage), moderate (middle stage) and severe (late stage) (12). However, AD begins with mild cognitive impairment (MCI) with amnesic symptoms without affecting other cognitive area (13, 14). The patients may fail to recall the latest activity or become confused with input they recently received (15). At this mild Alzheimer's stage, patients may function independently as language, motor, and sensory functions are not affected. Interestingly, not all patients with MCI develop into AD (12) and according to Ward, Tardiff (16) an average of 32% of MCI patients developed into AD within 5 years of follow up. Then followed by the mild AD in which the symptoms include poor object recognition, poor direction sense and reading problem. Even though the patients may function independently, but family and friends may be able to notice difficulties faced by these patients. In the moderate stage,

which for some is the longest and can last for many years, the patients will have personality changes and having difficulty in performing routine tasks (3). Thus, as the disease progress, these individuals require greater level of care. In the severe stage, the brain part involve in cognition and movement become damaged and therefore they become bed-bound (3). Thus, at this stage patients require help in basic activity of daily living.

Current drug therapy for AD is ineffective as the treatment does not prevent, slow or stop the progression of the disease. Treatment of patients diagnosed with AD is restricted to treat the cognitive symptoms of AD by helping to lessen or stabilize the symptoms for a limited time (17). Therefore, there is a pressing need for new and better treatments for AD as this disease condition is so disabling, and social and economic burden continue to grow on. Currently, there are 26 agents in phase III clinical trial of the 2018 AD pipeline (Figure 1). Among those treatments, 14 addressed amyloid targets, one involved a tau-related target, eight drugs for behavioral symptoms, one cognitive-enhancing agent, one involved neuroprotection, and one had a metabolic mechanism of action (18). Thus, search for a potential drug that might reverse the disease progression or even stop it from developing is at top priority.

**2018 Alzheimer's Drug Development Pipeline**



**Figure 1: Agents in clinical trials for treatment of Alzheimer's disease in 2018 ; adapted from Cummings, Lee (18)**

# Chapter 2

## 2.0 Research Questions

Alzheimer's disease (AD) is a progressive neurodegenerative brain disorder. The current AD drugs that are available in the market are purely symptomatic, with little or no beneficial effect on the disease progression. AD is not a result of a single factor but instead is a multifactorial condition. Thus, the failure of current anti-AD drugs justifies the need towards alternative novel drug candidates with no to minimum side effects. Earlier studies have proven that embelin from *Embelia ribes* Burm has antioxidant, anti-inflammatory, anticonvulsant and neuroprotective properties. Thus, we hypothesized that embelin is neuroprotective against AD by targeting the multifactorial condition and can be further developed into drug for treatment and/or prevention of AD. To that end, the specific aims of this thesis were:

- (i) To evaluate the neuroprotective effect of embelin on nootropics condition as well as in scopolamine induced amnesia rats (**Paper I**)
- (ii) To study the neuroprotective effect of embelin in permanent bilateral common carotid artery occlusion (PBOCCA) model of AD (**Paper II**)
- (iii) To investigate the neuroprotective effects of embelin as well as mRNA expression of targeted genes in STZ-induced sporadic AD like condition in an *in-vitro* model (**Paper III**)
- (iv) To study the effect of embelin on blood-brain barrier integrity and permeability (**Paper IV**)

# Chapter 3

### **3.1 Introduction**

AD is a multifactorial neurodegenerative disease involving many risk factors etiology (19). However, the pathogenesis of AD is still ambiguous due to the complex nature of human brain (20). Many hypotheses about AD have been developed, including amyloid beta ( $A\beta$ ), Tau, cholinergic and vascular dysfunction and so forth. Thus, many efforts have been made to develop anti-AD drugs based on these hypotheses to stop or revert the progression of AD. This literature review will summarize the pathological hypotheses of AD; limitations of the current therapy and role of embelin in central nervous system (CNS) related disorders.

### **3.2 Pathology of AD**

#### **3.2.1 The Amyloid Hypothesis**

The amyloid hypothesis was first proposed by Hardy and Allsop in 1991 and become one of the main etiologic hypothesis in AD which serve as the top explanation on how disease develop until now (21). This hypothesis proposed that imbalance between production and clearance of both intracellular and extracellular amyloid beta is the triggering event for progressive neuronal damage which fully characterizes the AD (22-24). Amyloid beta is a small peptide (~4.5 kDa) with resistance to proteolytic degradation (24, 25). It comprises up to 43 amino acids, in which most of amyloid beta is in the form of 1-40 and less than 5% are generated in isoform 1-42 (25, 26). Since the long form of 1-42 amyloid beta peptide is known to be the most hydrophobic (24), it has the greater tendency to aggregate and initiate the pathological oligomers, fibril and plaques (26). Oligomers and fibrils are considered to have the greatest toxicity while the final stage of senile plaques are relatively inactive (26). Amyloid beta peptide is generated through sequential proteolytic breaks of the amyloid precursor protein (APP) (24). APP is a transmembrane glycoprotein of type I which is highly expressed in the brain (27) and associated with neuronal development, neurite outgrowth, and axonal transport (28). Pathogenetic mutations of APP at  $\beta$ -secretase or  $\gamma$ -secretase cleavage sites are associated with an increase in amyloid beta 1-42 production (29). The increase in APP amount 1.5 times more compared to normal group proportionally give rise in amyloid beta which leads to AD-like pathology (30). According to Šalković-Petrišić (26) these potent amyloid beta neurotoxicants initiate a whole range of pathological cascade including microgliosis and astrocytosis, oxidative stress, inflammatory

response, neuronal dysfunction, cell death, neurotransmitter deficits, and finally cognitive dysfunction.

### **3.2.2. Tau Hypothesis**

Tau is encoded by a single gene, microtubule-associated protein tau (MAPT), which is located on human chromosome 17 (31). Localization of tau mRNA has been found in the soma and dendrites of both normal and AD neurons of human post-mortem brains based on in situ hybridization techniques (32). In CNS, the low molecular weight tau which is highly soluble is responsible for assembly and stabilization of microtubules required for morphogenesis, axonal transport and neuronal growth (33, 34). The microtubules formation activity is known to be regulated by certain degree of tau phosphorylation whereas the hyperphosphorylation of tau suppresses this activity (35, 36).

Abnormal hyperphosphorylated tau undergoes conformational changes, aggregates, and eventually leads to formation of neurofibrillary tangles (NFTs) (37). The accumulation of NFT inhibit the formation of microtubule which eventually causes malfunction of axonal transport (38). In pathological condition, NFT trigger neurotoxic actions that affect neurons by synaptic dysfunction (39) and by transmission from affected neurons to connecting naive neurons (40) and finally leads to neuronal degeneration. On the other hand, according to Dickey, Dunmore (41) tau haplotypes that rise tau expression increase AD risk and therefore reducing tau levels might be an alternative approach to protect against AD (24, 42).

### **3.2.3. Cholinergic Hypothesis**

One of the most common symptoms in AD is cognitive impairment which has been linked to a deficiency of brain neurotransmitter known as acetylcholine (43, 44). Acetylcholine (ACh) is used by cholinergic neurons for sending signal or message for attention, learning, memory, sensory, wakefulness and sleep related physiological processes (20, 45, 46). In AD pathological condition, damaged cholinergic neurons generate a downregulation of acetyltransferase and increase in acetylcholinesterase activity (24). Acetylcholinesterase metabolizes acetylcholine to choline and acetic acid. Thus, this will eventually reduce ACh release, impaired binding of Ach to nicotinic and muscarinic receptors, dysfunctional neurotrophin support, and deficits in axonal transport (47). The cholinergic system plays an important role in the regulation of cognitive functions, as

evidenced by the extensive loss of cholinergic neurons observed in Alzheimer's patients (48) which finally led to the "cholinergic hypothesis". Furthermore, AChE inhibitors have become important alternatives in the treatment of AD (49).

#### **3.2.4. Vascular hypothesis**

The vascular hypothesis for AD was first proposed by De la Torre and Mussivan (50) identifying vascular risk factors for involving brain, the heart and the circulation (51). According to vascular hypothesis, reduction in cerebral blood flow (CBF), glucose metabolism and oxygen utilization is the primary factor driving to AD pathogenesis (50). This condition appear to be inversely proportional to the disease severity which is consistent with the pathogenesis and progression of AD. Chronic cerebral hypoperfusion (CCH) is one of the major mechanisms of cerebral vascular disorders (52). CCH can affect the cerebral vascular system and eventually cause decreased blood supply to the brain (53). This condition will finally lead to progressive neuronal damage, transmission failure and brain tissue death (50) with cognitive impairment (54).

### **3.3 Limitations of current therapy**

Unfortunately, there is no cure has been found for AD. The pharmacological treatments available today for AD have not been shown to slow, reverse or prevent the disease progression (19). This is likely due to many factors which are involved in initiation and progression of AD based on the hypotheses discussed above. So far the US Food and Drug Administration (FDA) approved two drug classes for the treatments of AD which are known as AChE inhibitors (rivastigmine, galantamine, donepezil, tacrine) and not competitive N-methyl-D-aspartate (NMDA) receptor antagonist (memantine) (55). The aforementioned AChE inhibitors drugs raise ACh levels and enhance cholinergic functions in the brain (56). Likewise, memantine stabilizes dysfunctional glutamatergic neurotransmission and protect neurons from glutamate toxicity (55). Both these drugs; AChE inhibitors and NMDA receptor antagonists can only improve symptoms and are mainly used for mild to moderate AD and moderate to severe AD respectively (57). Besides, these drugs cause undesired side effects. For instance, tacrine caused liver problem and loss of appetite whereas for NMDA receptor antagonist, it causes hallucination, confusion, and mood swings following medication (58). Thus, due to its adverse side effects, tacrine is now discontinued in the United States (3) and memantine is only licensed in several countries (57). Furthermore, both

effectiveness and tolerability of these drugs have an important interpatient variability and these represent a limitation to current treatment for AD.

### **3.4 Role of embelin in CNS related disorders**

Embelin (2, 5-dihydroxy-3-undecyl-1, 4-benzoquinone) is a naturally occurring alkyl substituted hydroxyl benzoquinone which identified as a major constituent of the fruits of *Embelia ribes* Burm (Family: Myrsinaceae). Traditionally, this plant has been utilized as brain tonic for treating mental disorders (59). In this chapter we feature our published literature review on the role of embelin in CNS related disorders. In the publication, we did a literature search on embelin in a systematic way using appropriate electronic database from January 2000 to February 2017. As a result of this systematic search, we then presented the efficacy of embelin against CNS related complications.



# Plant Derived Phytochemical, Embelin in CNS Disorders: A Systematic Review

Uday P. Kundap, Saatheeyavaane Bhuvanendran, Yatinesh Kumari, Iekhsan Othman and Mohd. Farooq Shaikh\*

Neuropharmacology Research Laboratory, Jeffrey Cheah School of Medicine and Health Sciences, Monash University Malaysia, Selangor, Malaysia

## OPEN ACCESS

### Edited by:

Adolfo Andrade-Cetto,  
National Autonomous University of  
Mexico, Mexico

### Reviewed by:

Benedict Green,  
Poisonous Plant Research Laboratory,  
ARS-USDA, USA  
Rajendra Karki,  
St. Jude Children's Research Hospital,  
USA

### \*Correspondence:

Mohd. Farooq Shaikh  
farooq.shaikh@monash.edu

### Specialty section:

This article was submitted to  
Ethnopharmacology,  
a section of the journal  
Frontiers in Pharmacology

**Received:** 09 September 2016

**Accepted:** 07 February 2017

**Published:** 27 February 2017

### Citation:

Kundap UP, Bhuvanendran S,  
Kumari Y, Othman I and Shaikh MF  
(2017) Plant Derived Phytochemical,  
Embelin in CNS Disorders: A  
Systematic Review.  
Front. Pharmacol. 8:76.  
doi: 10.3389/fphar.2017.00076

A Central nervous system (CNS) disease is the one which affects either the spinal cord or brain and causing neurological or psychiatric complications. During the nineteenth century, modern medicines have occupied the therapy for many ailments and are widely used these days. Herbal medicines have often maintained popularity for historical and cultural reasons and also considered safer as they originate from natural sources. Embelin is a plant-based benzoquinone which is the major active constituent of the fruits of *Embelia ribes* Burm. It is an Indo-Malaysian species, extensively used in various traditional medicine systems for treating various diseases. Several natural products including quinone derivatives, which are considered to possess better safety and efficacy profile, are known for their CNS related activity. The bright orange hydroxybenzoquinone embelin-rich fruits of *E. ribes* have become popular in ethnomedicine. The present systematic review summarizes the effects of embelin on central nervous system and related diseases. A PRISMA model for systematic review was utilized for search. Various electronic databases such as Pubmed, Springer, Scopus, ScienceDirect, and Google Scholar were searched between January 2000 and February 2016. Based on the search criteria for the literature, 13 qualified articles were selected and discussed in this review. The results of the report showed that there is a lack of translational research and not a single study was found in human. This report gives embelin a further way to be explored in clinical trials for its safety and efficacy.

**Keywords:** embelin, CNS disorders, neuropharmacology, neurodegenerative diseases, natural product

## INTRODUCTION

Central nervous system (CNS) is an integral part of the nervous system. It consists of the brain and spinal cord, and are associated with a number of important actions of the body. A CNS disease can be defined as one which affects either the spinal cord (myelopathy) or brain (encephalopathy) or both. The etiology of CNS involves a number of factors, for example, structural defects, infections, trauma, autoimmune disorders, tumors, neurodegeneration, and others, which may lead to neurological or neuropsychiatric or neurodegenerative or neurodevelopment disorders (Cannas et al., 2002; Upadhyay, 2014). The prevalence of CNS diseases is at least two times higher in developing countries than developed countries. According to World Health Organization (WHO), traditional medicines have become a topic of global importance. In many developing countries, a large proportion of the population relies heavily on traditional healers and phytomedicine for

primary health care requirements. Concurrently, many people in developed countries have begun to turn to alternative or complementary therapies, including medicinal herbs (World Health Organization, 1999; Saraf, 2012).

Embelin is chemically known as 2,5-dihydroxy-3-undecyl-1,4-benzoquinone, which is the major active constituent of the fruits from *Embelia ribes* Burm (Family: Myrsinaceae), commonly known as “False Black Pepper” (Figure 1). It is an Indo-Malaysian species, reported from India, Sri Lanka, Singapore, Malaysia, and South China. *Embelia ribes* Burm is extensively used in Indian, Folk, Homeopathy, Tibetan, Unani, and Siddha traditional medicinal systems for treating various ailments like chronic inflammatory disorders, heart and urinary conditions, snake and insect bites, and tumor (Radhakrishnan et al., 2012). The dried fruit is considered anthelmintic, astringent, carminative, alterative, and stimulant (Nadkarni, 1996). Embelin is already studied for its safety and toxicity profile in rodents and non-rodents. It is reported that embelin is safe up to 3 g/kg orally when tested in rodents after acute exposure. Another report on subacute toxicity after repeated administration of embelin at 10 mg/kg dose found to be safe in rats (Poojari, 2014).

Fruits of *E. ribes* have been used for the treatment of central nervous system (CNS) disorders, mental disorders and as a brain tonic in the traditional systems of medicine. Embelin was found to be useful in decreased cerebral infarction area and histopathological alteration, such as normal glial density, decreased edema, absence of lymphocytes, congestion of blood vessels, and necrosis. These reports suggest that embelin would be useful as an adjunct therapy for cerebral stroke and as a potent neuroprotective agent (Thippeswamy et al., 2011). Embelin possesses all the characteristics of a compound which can cross the blood-brain barrier (BBB) and elicit an effect on the CNS (Pathan et al., 2009). Embelin reported for its CNS effect by diverse mechanisms, namely by scavenging free radicals and antioxidant effect, by inhibiting pro-inflammatory cytokines like NF- $\kappa$ B and p53, by modulating sodium channel, chloride conductance, and GABA<sub>A</sub> receptor, by inhibiting STAT3, XIAP, and PPAR $\gamma$  pathways (Figure 2).

Embelin has been explored and reported for various CNS disorders using cell lines and animal models. There is no single study which summarizes the effectiveness of embelin in CNS associated disorders. Although embelin proved to be effective in laboratories against various CNS disorders, but it is not being

translated to humans yet. In the present systematic review, an effort is being made to systematically review all the literature available with embelin in animal and clinical research.

## MATERIALS AND METHODS

### Search Technique

The extensive literature search was done to conduct a systematic review summarizing the effects of embelin on central nervous system and related diseases. Various electronic databases were used, namely Pubmed, Springer, Scopus, ScienceDirect, and Google Scholar between the period January 2000 and February 2016. The following keywords were searched individually and in combination with the embelin: brain, trauma, CNS, neurological disorder, neurodegenerative disease, and psychological disorder.

### Study Selection and Exclusion/Inclusion Criteria

The search was limited to, articles published in English language and original research articles only. Abstracts of symposiums and conferences, review articles, books, and patents were excluded due to insufficient information for evaluation and comparison. Articles which were not related to CNS diseases were excluded. Any clinical, pre-clinical, *ex-vivo*, and *in-vitro* studies were also the part of the inclusion criteria.

### Data Extraction

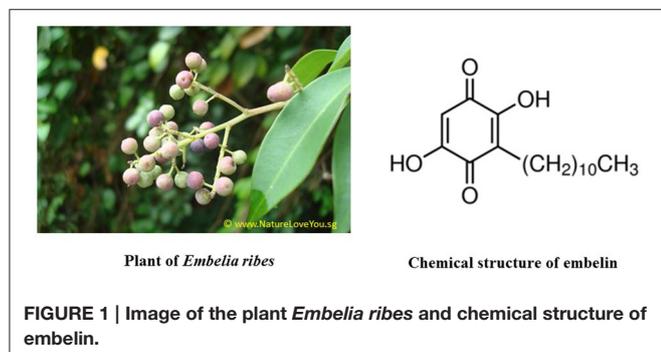
Two separate researchers obtain data independently, and then the titles and abstracts of each article were compared to delete duplication of the data. Based on the mentioned eligibility criteria for the literature search, 14 articles were excluded and 13 qualified articles were evaluated in this study. The aim of using PRISMA statement is to help authors to understand and improve the reporting of systematic reviews and meta-analyses related to use of embelin in CNS related disorders (Moher et al., 2015). Flow diagram was prepared according to the guidelines of PRISMA-Transparent reporting of systematic reviews and meta-analyses (Moher et al., 2009).

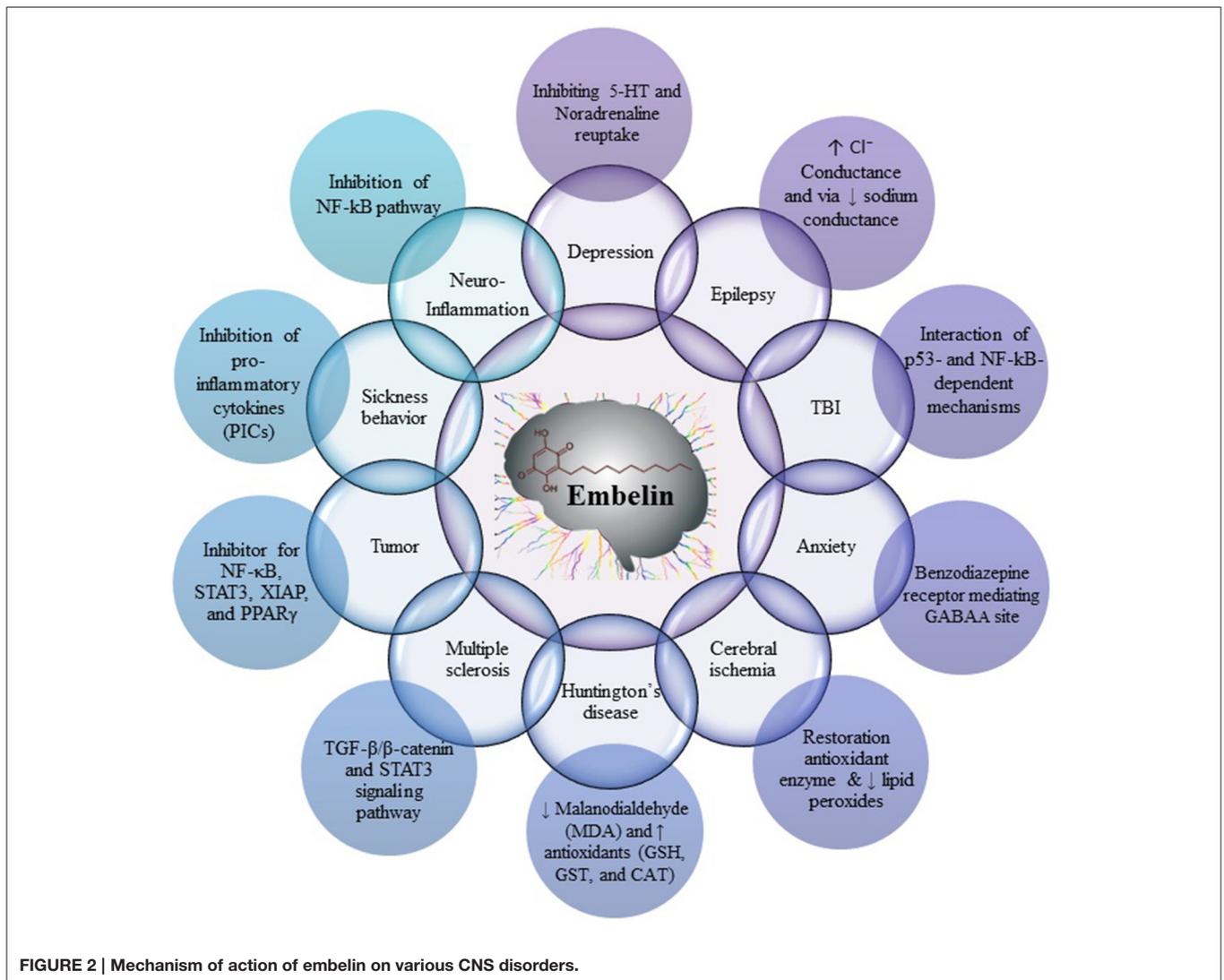
## RESULTS AND DISCUSSION

The search based on the keywords mentioned in the methodology yielded 6,448 records. After applying exclusion criteria, total articles removed were 6,435, which includes; (a) 3,470 reviews, book and patents, (b) 2,090 did not meet review criteria, (c) 402 abstracts, (d) 459 duplicates, and (e) 14 not relevant to the aim of the review based as they deal with formulations of the embelin (Figure 2). Thirteen eligible articles were included, compiled in Table 1 and discussed in the present systematic review (Figure 3).

### Anticonvulsant Activity

Mahendran et al. (2011b) isolated embelin from the berries of *E. ribes* and reported on the anticonvulsant activity of the embelin using maximal electroshock (MES) and pentylenetetrazole (PTZ). MES cause the spread of seizure similar to grandmal epilepsy. In MES method, brief high-intensity shock is applied to the head through corneal or ear electrodes with a stimulator





that either delivers a constant current to constant voltage at a frequency of 50–60/s. The MES convulsions are divided into five phases such as the phase of tonic limb flexion, the phase of tonic limb extension, the phase of clonic convulsions, stupor, and recovery or death (Castel-Branco et al., 2009). Protection against hind leg tonic extension (HLTE) in MES predicts the ability of embelin to prevent the spread of seizure discharge from the epileptic focus in the brain and suppressing generalized tonic-clonic and partial seizures. Phenytoin is said to protect against seizures by causing blockage of voltage-dependent, voltage-gated sodium channels. This block sustains repetitive high-frequency firing of action potentials. The results show that there is a complete absence of HLTE in MES model when treated with embelin 10 mg/kg dose in comparison with phenytoin. It shows that embelin at 10 mg/kg dose might act via blockade of voltage-gated sodium channels. Embelin at 2.5, 5, and 10 mg/kg shows, dose-dependent activity against MES model which act of sustaining repetitive high-frequency firing of action potential

exhibiting anticonvulsant activity. Pentylentetrazole (PTZ) is used to induce clones seizure-like behavior with increased locomotor activity. The increased chloride conductance drives the membrane potential toward the reversal potential of the  $\text{Cl}^-$  ion which is about  $-65$  mV in neurons, inhibiting the firing of new action potentials. This mechanism is responsible for the anti-epileptic effects of  $\text{GABA}_A$  allosteric agonists. Embelin might increase chloride conductance which drives membrane potential, inhibiting the firing of action potential delaying onset of a clonic-tonic seizure in PTZ induced epilepsy. PTZ basically acts at the picrotoxin site of the  $\text{GABA}_A$  receptor and reduces chloride conductance which further leads to glutamate excitation (Desmond et al., 2012).

For each seizure model, embelin was administered intraperitoneally 30 min prior to the induction of MES and PTZ. Administration of embelin (2.5 and 5 mg/kg, i.p.) showed significant ( $P < 0.001$ ) reduction in the duration of HLTE compared to the control. Based on the results, the study

**TABLE 1 | Pharmacological activities reported with embelin in central nervous system related disorders.**

Sr. no.	CNS activity	Study sample	Source of embelin	Dose	Result	Number of citations	Author (year)
1	Anticonvulsant activity	Swiss albino rats (150–200 g; <i>n</i> = 6) Swiss albino mice (20–25 g; <i>n</i> = 6)	Embelin isolated from berries of <i>Embelia ribes</i>	2.5, 5, and 10 mg/kg	<ul style="list-style-type: none"> <li>• ↓ in the duration of HLTE in MES (2.5 and 5 mg/kg, i.p.).</li> <li>• Electroshock—100% protection against mortality.</li> <li>• ↑ Clonic and tonic onsets at all dose.</li> </ul>	54	Mahendran et al., 2011b
2	Antidepressant activity	Swiss albino mice (20–25 g; <i>n</i> = 6)	Embelin isolated from berries of <i>Embelia ribes</i>	2.5 and 5 mg/kg	<ul style="list-style-type: none"> <li>• Antidepressant-like effect in Tail suspension test (TST).</li> <li>• ↓ Immobility in the Forced swimming test (FST).</li> <li>• Exhibited significant activity in mice TST and FST experimental models.</li> </ul>	6	Gupta et al., 2013
3	Anxiolytic activity	Swiss albino mice ( <i>n</i> = 6)	Embelin isolated from berries of <i>Embelia ribes</i>	2.5 and 5 mg/kg	<ul style="list-style-type: none"> <li>• ↑ Time spent and number of entries in open arm (elevated plus maze).</li> <li>• ↓ Decrease in the duration of immobility in light box (light and dark model).</li> <li>• ↑ Increase rearing assisted rearing and number of square crossed (open field test).</li> <li>• Embelin showed its anxiolytic effect in dose-dependent manner.</li> </ul>	6	Afzal et al., 2012
4	Sickness behavior	Male Swiss albino mice (25–30 g; <i>n</i> = 8)	Embelin isolated from berries of <i>Embelia ribes</i>	10 and 20 mg/kg	<ul style="list-style-type: none"> <li>• Embelin prevented anhedonia, anorexia.</li> <li>• Ameliorated brain oxidative stress markers.</li> <li>• Protective effect of embelin in LPS-induced sickness behavior in mice.</li> </ul>	0	Shaikh et al., 2016
5	Huntington's disease	Adult Wistar rats (190–220 g; <i>n</i> = 8)	Embelin isolated from berries of <i>Embelia ribes</i>	10 and 20 mg/kg/day	<ul style="list-style-type: none"> <li>• Loss of body weight.</li> <li>• Decreased the oxidative stress.</li> <li>• Decrease of 69–76% brain lesion.</li> <li>• Protect the neurons from 3-NP toxicity.</li> </ul>	1	Dhadde et al., 2016
6	Multiple sclerosis (Autoimmune encephalomyelitis, CNS inflammation)	Female C57BL/6 mice, aged 6–8 weeks ( <i>n</i> = 6)	Embelin pure form	25 and 50 mg/kg	<ul style="list-style-type: none"> <li>• -↓ Human CD14+ monocyte-derived dendritic cell differentiation.</li> <li>• -↓ Duction in the EAE (experimental autoimmune encephalomyelitis) clinical score.</li> <li>• -↓ Inflammatory Th1 and Th17 cells in EAE.</li> </ul>	9	Xue et al., 2014
7	Traumatic brain injury	Female Sprague–Dawley rats, male C57BL/6 mice ( <i>n</i> = 10)	Embelin pure form	200 nM	<ul style="list-style-type: none"> <li>• Inhibition of NF-κB expression of XIAP increases in PFT-treated animals.</li> <li>• p53 and NF-κB dependent mechanisms delayed neurodegeneration</li> </ul>	62	Plesnila et al., 2007
8	Hypoxia-ischemia (HI) induced neurological injury	Female and male Wistar rats pups ( <i>n</i> = 15)	Embelin pure form (Sigma-Aldrich, USA)	20 mg/kg embelin	<ul style="list-style-type: none"> <li>• Confirm sex differences in behavioral and anatomical outcome.</li> <li>• XIAP acts to protect the female brain from the early HI injury.</li> </ul>	18	Hill et al., 2011

(Continued)

TABLE 1 | Continued

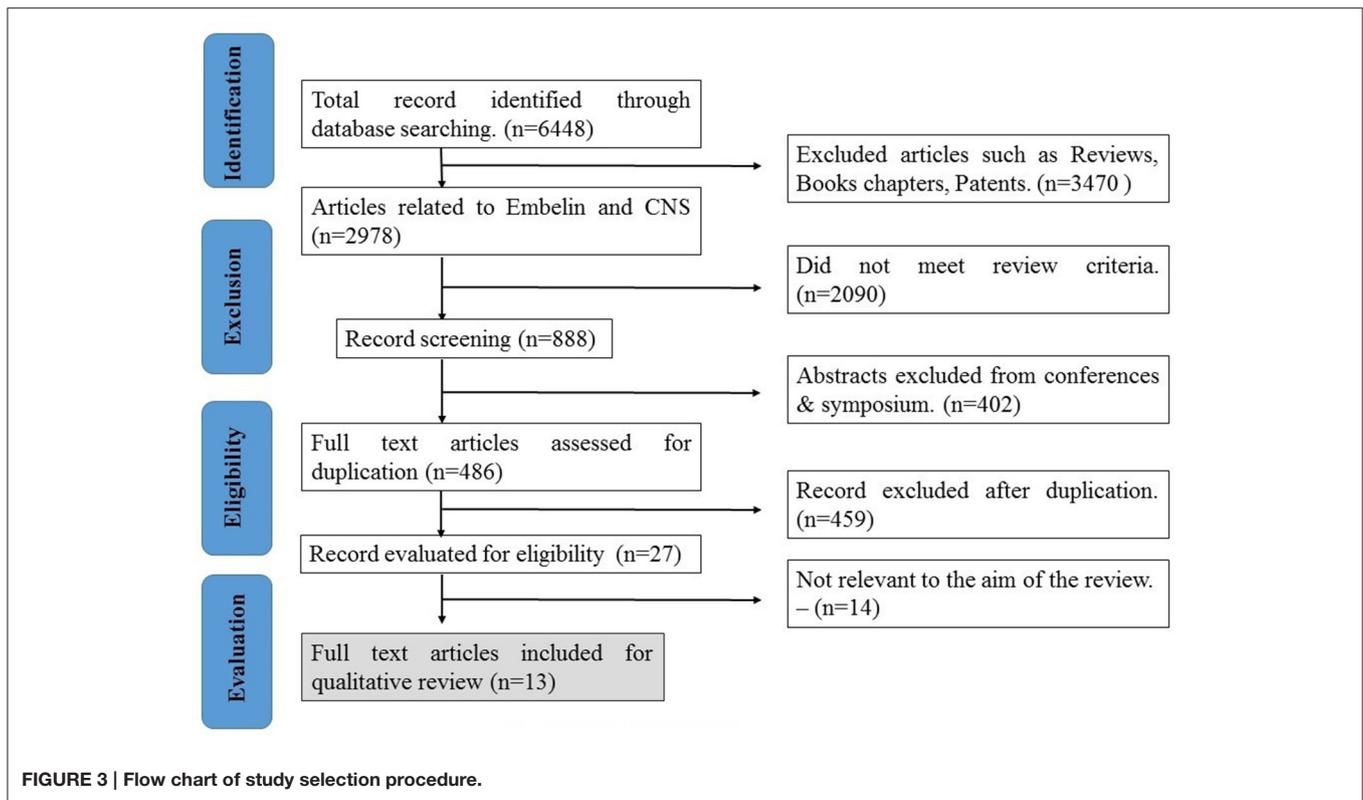
Sr. no.	CNS activity	Study sample	Source of embelin	Dose	Result	Number of citations	Author (year)
9	Global ischemia/reperfusion-induced brain injury	Male Wistar rats (200–260g; $n = 6$ )	Extraction of embelin from <i>Embelia ribes</i>	25 and 50 mg/kg	<ul style="list-style-type: none"> <li>• ↑ Locomotor activity and hanging latency time.</li> <li>• ↓ Beam walking latency.</li> <li>• ↓ Lipid peroxidation.</li> <li>• ↑ Total thiol content and glutathione-S-transferase neuroprotective agent and useful in the treatment of stroke.</li> </ul>	22	Thippeswamy et al., 2011
10	Focal cerebral ischemia brain	Male Wistar rats (200-250 g; $n = 6$ )	Embelin isolated from berries of <i>Embelia ribes</i>	50, 75, 100 mg/kg	<ul style="list-style-type: none"> <li>• Decreased the infarction and edema (100 mg/kg).</li> <li>• Decreased MDA level (75 and 100 mg/kg).</li> <li>• ↑ SOD and CAT (100 mg/kg).</li> </ul>	0	Patel and Gohil, 2014
11	Cerebral ischemia	C57BL/6 male, Gl female, and Ovx female mice ( $n = 7$ )	Embelin pure form (Sigma-Aldrich, USA)	20 mg/kg	<ul style="list-style-type: none"> <li>• Inhibitor of XIAP exacerbated stroke-induced injury in females but had no effect in males.</li> </ul>	97	Siegel et al., 2011
12	Apoptosis in human glioma cells via NF- $\kappa$ B inhibition	Human glioma cell lines T98G, U87MG, and H4. Immortalized primary human fetal astrocytes (IM-PHFA)	Embelin pure form (Sigma-Aldrich, USA)	(0–50 $\mu$ M)	<ul style="list-style-type: none"> <li>• Embelin suppressed proliferation of human glioma cells.</li> <li>• Apoptosis in human glioma cells by inhibiting NF-<math>\kappa</math>B.</li> <li>• ↓ NF-<math>\kappa</math>B activity by reducing nuclear translocation of p65.</li> </ul>	20	Park et al., 2013
13	Apoptosis in human glioma cells via the mitochondrial pathway	Human brain glioma U87 cells	Embelin pure form (Sigma-Aldrich, USA)	(0, 50, and 100 $\mu$ g/ml)	<ul style="list-style-type: none"> <li>• Time- and dose-dependent apoptosis of brain glioma cells.</li> <li>• Arrest the cell cycle in the G0/G1 phase.</li> <li>• Changes in brain glioma cell mitochondrial membrane potential.</li> <li>• Shifting of Bax and Bcl-2 to cause apoptosis.</li> </ul>	6	Wang et al., 2013

demonstrated that embelin at 10 mg/kg dose could significantly reduce the duration of the HLTE in MES model. Embelin at all the three doses significantly ( $P < 0.001$ ) decreased the onset of stupor when compared to the control. Eventually, the percentage protection reported was 100% as no mortality was observed in all the embelin treated rats when challenged with maximum electroshock. On the other hand, embelin significantly delayed the onset of clonic and tonic seizures with an increased in a survival rate in a dose-dependent manner when checked against PTZ. Embelin also exhibited significant and dose-dependent delayed the onset of clonic-tonic actions and protection from PTZ induced mortality. At 5 and 10 mg/kg doses, it exhibited 50 and 83.33% protection against mortality. This study does not include the effectiveness of embelin in chronic models of epilepsy like lithium-Pilocarpine, kindling, or intracerebroventricular kainic acid. Based on the preliminary results, Mahendran et al. (2011a) postulated that embelin is a potent anticonvulsant phytochemical and the plausible mechanism is through GABAergic modulation. But, there was no supporting information like brain GABA estimation

or GABA receptor expression were included in the published report.

### Antidepressant Activity

Depression is one of the common neuropsychiatric disorder which contribute to the global burden of the diseases affects about 1 in 20 people across the world (Currie and World Health Organization Regional Office for Europe, 2000). Gupta et al. (2013) reported the anti-depressant effect of embelin in experimental animals using two universally accepted experimental models: mice tail suspension test (TST) and forced swimming test (FST). Embelin was isolated from fresh fruits of *E. ribes*. The TST is used as an experimental method in scientific research to measure stress in rodents. The FST—in a rodent is used for evaluation of the antidepressant efficacy of new compounds, antidepressant drugs, and experimental development that are aimed at translating or preventing depressive-like states. It is based on the observation that if an animal is subjected to short term inescapable stress then it will become immobile. It has been described as rendering



a situation in which “behavioral despair” is induced; that is, the animal loses hope to escape the stressful environment. It is well-known that compounds which selectively bind to high-affinity benzodiazepine receptors possess both anxiolytic and antidepressant effects. The anxiolytic effect of embelin was shown to be mediated through the effect on the GABA system. The similar mechanism of antidepressant action cannot be ruled out (Afzal et al., 2012). It is, therefore, reasonable to assume that the observed antidepressant-like activity of embelin could be attributed to its known antioxidant effect. GABA<sub>A</sub> receptors are allosteric modulatory sites for benzodiazepine. They are probably composed of five protein subunits, at least some of which belong to different subunit classes. So far GABA<sub>A</sub> receptors have been identified as six alpha-four beta-three gamma-, and delta-and two-rho =  $\rho$  subunits. A 3D-structural model similarity, further shows that embelin is closely related with the well-known antioxidant alpha-tocopherol (AT, vitamin E), especially in the long-chain non-polar tails and polar phenolic heads (Lobato et al., 2010). Embelin at 2.5 and 5 mg/kg dose administered orally and tested using tail suspension test (TST) and forced swimming test (FST) in mice. It was found to effectively reduce immobile time in both experimental models suggesting its antidepressant potential. Embelin (5 mg/kg) was reported to be comparably better than the standard antidepressant drug, imipramine (15 mg/kg), which is a tricyclic antidepressant drug (Gupta et al., 2013). As a criticism, Gupta et al. (2013) fail to mention about actual activity of embelin as an anti-depressant, which may be via inhibiting 5-hydroxytryptamine receptors (5-HT) and noradrenaline (NA) reuptake. In their study, imipramine was

used as a positive control which has the known antidepressant activity and it acts by inhibiting NA and 5-HT reuptake into neurons.

## Anxiolytic Activity

Anxiety is a feeling of discontent, such as fear or worry that can be intense or gentle. Everyone has feelings of anxiety at some point in their life for example, you may feel worried and anxious about a job interview or having a medical examination or sitting an exam. An anxiolytic is a medication or other intervention that inhibits anxiety. This effect is in contrast to anxiogenic agents, which increases anxiety. Afzal et al. (2012) revealed the anxiolytic potential of embelin using behavioral models of anxiety. The elevated plus maze (EPM) is a test used to measure anxiety in laboratory animals. The test uses two open and two enclosed arm apparatus with an elevated, plus-shaped (+). The behavioral model is based on the general disinclination of rodents to open spaces. This disinclination leads to the behavior termed thigmotaxis, a greater liking to remain in enclosed spaces or close to the edges of a bounded space. Reduction in anxiety is indicated in the plus-maze by an increase in the amount of time spent or entries in the open arms (time or entries in open arms/total time or total entries in open or closed arms; Walf and Frye, 2007). Open field test (OFT) is an experiment used to assay general locomotor activity levels and anxiety in rodents. Rodents display a natural aversion to brightly light areas. They also have an urge to explore a perceived threatening stimulus. The result of these two conflicting drives is anxiety. The increase in exploratory behavior leads to decreased anxiety. Increased

anxiety will result in less locomotor motion and the animal will have a preference to remain at the edges of the field (Ramos, 2008). The light/dark test is based on the natural version of rodents to brightly illuminated areas and on the spontaneous exploratory behavior of rodents in response to mild stress, that is, novel environment and light. The test apparatus consists of a small dark safe compartment (one-third) and a large illuminated preference compartment (two-thirds; Bourin and Hascoët, 2003). Embelin at 5 mg/kg dose significantly increased the percentage of time spent and the number of entries in open arm in EPM apparatus. Percentage of time spent in the open arms and number of open arm entries was significantly ( $P < 0.01$  and  $P < 0.001$ ) increased by embelin (2.5 and 5 mg/kg) and diazepam. Time spent in the open arm by animal treated with embelin 2.5 and 5 mg/kg dose was  $47.92 \pm 1.25$  and  $66.17 \pm 1.93$  and no. of entries in the open arm by animal treated with embelin 2.5 and 5 mg/kg dose was  $5.61 \pm 0.47$  and  $7.90 \pm 0.45$  significant. The result shows that embelin exhibited dose-dependent activity as an anxiolytic in mice EPM-test. In the open field test, embelin exhibited a significant increase in a number of rearing, assisted rearing and number of the crossing. A number of rearing in open field test by animal treated with embelin 2.5 and 5 mg/kg dose was  $17.68 \pm 0.52$  and  $20.33 \pm 0.59$  and number of assisted rearing in open field test by animal treated with embelin 2.5 and 5 mg/kg dose was  $16.20 \pm 1.00$  and  $21.12 \pm 1.2$ . In light and dark model, embelin produced a significant increase in time spent, the number of crossing and decrease in the duration of immobility in a light box. The animals treated with diazepam (1 mg/kg) and embelin (2.5 and 5 mg/kg) showed significant ( $P < 0.05$  and  $P < 0.001$ ) increase in the time spent in the lighted box and decrease in the time spent in the dark box. Time spent in the lighted box (s) by animal treated with embelin 2.5 and 5 mg/kg dose was  $94.92 \pm 1.73$  and  $116.8 \pm 4.24$  and time spent in a dark box (s) by animal treated with embelin 2.5 and 5 mg/kg dose was  $167.0 \pm 4.59$  and  $148.3 \pm 1.26$ . Embelin at 2.5 mg/kg dose, failed to produce any significant change in the number of crossing and duration of immobility. Afzal et al. (2012) concluded that embelin exhibits significant anxiolytic activity in a dose dependant manner. They proposed that the observed activity could be due to an antagonistic effect on GABA receptor complex as most of the anxiolytic and antidepressant molecules selectively bind to high-affinity benzodiazepine binding site, present on GABA receptor. Both Gupta et al. (2013) and Afzal et al. (2012) contradicted about vehicle used to dissolve embelin, they mentioned two different vehicles, olive oil and 1% Tween 80 (v/v) as embelin has poor water solubility.

## Sickness Behavior

During the course of an infection, the adaptive behavioral changes that develop in ill individuals is known as sickness behavior. It is relevant to understand depression and some aspects of the suffering in any disease. Sickness behavior is like a complex behavior induced by infections and immune trauma and mediated by pro-inflammatory cytokines. Some of the evidence state that sickness behavior is mediated through the effects of pro-inflammatory cytokines (PICs), such as IL-1, TNF $\alpha$ , and IL-6 (Maes et al., 2012). Embelin has been reported to possess

neuroprotective, anxiolytic and antiinflammatory assets and has been shown to inhibit Nf- $\kappa$ B pathway and cytokine production (Mahendran et al., 2011a). Few characteristics of the behavioral pattern including malaise, hyperalgesia, pyrexia, listlessness, and disinterest in social interactions with the environment, lethargy, behavioral inhibition, exploration and grooming, reduction of reproductive performance, anorexia and weight loss, failure to concentrate, and anxiety (Maes et al., 2012). The effect of embelin was evaluated in sickness behavior in mice by Shaikh et al. (2016). Adult male Swiss albino mice were pre-treated with embelin (10 and 20 mg/kg per oral) for 3 days and then challenged with lipopolysaccharide (LPS; 400  $\mu$ g/kg intraperitoneal). In EPM-test, pre-treatment with embelin (10 and 20 mg/kg) and dexamethasone (1 mg/kg) significantly reversed LPS-mediated effects and increased both the number of open arm entries ( $3.00 \pm 0.53$ ,  $3.12 \pm 0.58$  and  $3.12 \pm 0.47$ , respectively) and time spent in open arm ( $15.38 \pm 3.19$ ,  $14.00 \pm 2.67$  and  $13.63 \pm 1.94$  s, respectively) when compared with LPS-alone. In light-dark box test, Pre-treatment with both the tested doses of embelin and dexamethasone (1 mg/kg) prior to LPS-shot significantly increased the time spent in the light compartment ( $33.88 \pm 2.11$ ,  $43.75 \pm 6.81$  and  $34.13 \pm 4.38$  s, respectively). In the forced swim test, embelin (10 and 20 mg/kg) prior to LPS-injection significantly decreased the floating time ( $78.75 \pm 5.03$  and  $62.88 \pm 5.03$  s, respectively) when compared with LPS-alone-administered group. In social behavior tests, social exploration was measured just before the administration of LPS and again 2, 4, 8, and 24 h later. LPS-associated reduction in social behavior was attenuated by pre-treatment with embelin 10 mg/kg ( $20.61 \pm 4.15\%$ ,  $29.24 \pm 8.45\%$  and  $56.61 \pm 5.44\%$ , respectively) and 20 mg/kg ( $38.41 \pm 5.90\%$ ,  $44.78 \pm 5.17\%$  and  $63.55 \pm 5.95\%$ , respectively), dexamethasone 1 mg/kg ( $43.84 \pm 5.31\%$ ,  $49.12 \pm 2.95\%$  and  $64.87 \pm 4.42\%$ , respectively) when compared with LPS-alone-treated animals. In the open field test, pre-treatment with embelin (10 and 20 mg/kg) and dexamethasone (1 mg/kg) significantly attenuated LPS-induced changes and increased the peripheral, central and total number of line crossings and a number of climbs rear when compared with LPS-alone-treated group. Food and water intake test, pre-treatment of LPS-challenged mice with embelin (10 and 20 mg/kg) and dexamethasone (1 mg/kg) significantly reversed LPS-induced anorexia and adipisia in comparison to animals with LPS-alone-treated group. This all comparative finding eventually concluded that embelin is neuroprotective against LPS-induced sickness behavior in mice (Shaikh et al., 2016).

## Huntington's Disease

Huntington's disease (HD) is a progressive neurodegenerative disorder associated with severe degeneration of basal ganglia neurons, which affects muscle coordination and leads to mental decline and behavioral symptoms. Systemic administration of 3-nitropropionic acid (3-NP), an inhibitor of the mitochondrial citric acid cycle, results in a progressive locomotor deterioration resembling that of HD. It differs mechanistically from excitotoxic lesions in that 3-NP irreversibly inhibits the mitochondrial citric acid cycle and leads to depressed ATP levels and elevated lactate concentrations (Borlongan et al., 1997; Brouillet, 2014).

The study carried out by Dhadde et al. (2016) evaluated the neuroprotective potential of embelin against 3-nitropropionic acid (NP) induced experimental HD in rats. 3-NP significantly altered the behavioral and neuronal antioxidant status and caused significant neuronal damage in the striatal region. Elevated levels of malondialdehyde (MDA) and decreased levels of antioxidants (GSH, GST, and CAT) in the 3-NP treated rat brains supports the increased oxidative stress in HD. Behavioral tests were carried in the following order: neurological scoring, locomotor activity, EPM-test, beam walking test and hanging wire test. Biochemical estimation and brain lesion measurement were carried out in order to explore the molecular and structural differences of embelin in the brain. Administration of 3-NP alone shows motor abnormalities, decreased locomotor counts, loss of memory in EPM, decreased motor coordination in beam walking test, decreased hanging latency on hanging wire test and even 3-NP alone treatment resulted in highly significant ( $p < 0.001$ ) reduction in body weight. In neurological scoring, none of the rats in embelin treated groups (10 and 20 mg/kg) showed hind limb paralysis and inability to move indicating its potent activity in reversing 3-NP induced motor abnormalities. The treatment with embelin at both the doses (10 and 20 mg/kg) reversed the decrease in locomotor counts induced by 3-NP toward the normal, and it was found to be  $129.2 \pm 5.58$  and  $160 \pm 11.14$ , thus both the doses of embelin showed improvement in the locomotor count. In the EPM-test, embelin treatment at 10 mg/kg body weight significantly ( $p < 0.01$ ) Reversed the memory loss ( $27.73 \pm 3.92\%$ ) induced by 3-NP toward the normal, when compared with 3-NP alone treated animals. However, embelin at 20 mg/kg body weight dose showed a complete reversal of 3-NP induced memory loss same as a normal control group. At beam walking test, treatment dose of embelin (10 and 20 mg/kg) to 3-NP treated rats significantly ( $p < 0.001$ ) improved the motor coordination and body balance. These animals traversed the beam in  $5.11 \pm 0.66$  and  $5.51 \pm 0.72$  s, respectively. In hanging wire test, embelin at doses of 10 and 20 mg/kg increased 3-NP induced decrease in hanging latency period, with values  $36.66 \pm 1.78$  ( $p < 0.05$ ) and  $49.34 \pm 2.62$  s. The percentage decrease in the brain lesion area in both these groups was 69.59 and 76.21%, respectively. Embelin at 10 and 20 mg/kg to 3-NP treated animals significantly ( $p < 0.01$ ) reduced the brain lesion area to  $4.32 \pm 0.44$  and  $3.38 \pm 0.17\%$ , respectively. Embelin treatment significantly protected neurons against 3-NP induced toxicity and reduced brain lesion up to 76%. It also exhibited a significant antioxidant and improved behavioral alterations induced by 3-NP. It is postulated that effectiveness of embelin could be due to its antioxidant potential and ability of embelin to modulate  $Ca^{2+}$  influx associated with increased brain glutamate levels (Dhadde et al., 2016). In 3-NP induced HD like condition model in rats, embelin found to be effective neuroprotectant.

## Multiple Sclerosis (MS)

A chronic, typically progressive damage to the sheaths of nerve cells in the brain and spinal cord is termed as multiple sclerosis (MS). Symptoms may include numbness, impairment of speech and muscular coordination, blurred vision, and severe fatigue (Loma and Heyman, 2011). Animal models of brain inflammation are used to study autoimmune encephalomyelitis,

or experimental allergic encephalomyelitis (EAE). Dendritic cells (DCs) have a pivotal role in the immune response and in stimulating naïve T-lymphocytes. Induction and maintenance of self-tolerance is a critical role of DCs and the failure of which can lead to autoimmune/inflammatory diseases. Embelin concentrations of 10, 30, and 60  $\mu\text{M}$ , inhibits the differentiation and endocytosis of Human Monocyte-Derived dendritic cell (DCs). Compared with the day 5 untreated iDCs, a significant dose-dependent reduction in cell surface marker expression was observed in EB-treated cells. These results indicate that embelin inhibited the differentiation of human  $CD14^+$  monocytes into DCs in a dose-dependent manner. DC-derived cytokines are required for the polarization of the adaptive immune response. Therefore, Xue et al. (2014) investigated the potential effects of embelin on the regulation of the expression of the cell-polarizing cytokines. The production of the inflammatory cytokine tumor necrosis factor-alpha ( $\text{TNF-}\alpha$ ), the Th1 cell polarizing cytokine IL-12p35, the Th17 cell-polarizing cytokines IL-6 and IL-12/23p40, and the Th1 cytokine IFN- $\gamma$  is substantially inhibited by embelin. Embelin suppressed the DC-mediated polarization of Th1 and Th17 cells and that it may be useful for the treatment of autoimmune inflammatory diseases that are mediated by Th1 and Th17 cells. Embelin ameliorates the clinical severity of experimental autoimmune encephalomyelitis (EAE). Compared with PBS-treated mice, the incidence of clinical symptoms in the 25 and 50 mg/kg/day EB-treated mice were reduced. These data suggest that embelin significantly ameliorates the clinical outcome of EAE. TGF- $\beta$ / $\beta$ -catenin and STAT3 signaling pathway are used by embelin to inhibit DC function, which leads to a reduction in the EAE clinical score and in CNS inflammation and demyelination. The novel finding of this study is that the anti-inflammatory effect of embelin appears to require the presence of functional TGF- $\beta$ / $\beta$ -catenin and the absence of activated STAT3 in DCs. It was also found that embelin-induced inhibition of the differentiation of Th1 and Th17 cells was associated with a down regulation of the production of Th1-polarizing and Th17-polarizing. Embelin, a novel XIAP inhibitor, significantly increased TGF- $\beta$ / $\beta$ -catenin signaling and decreased STAT3 phosphorylation in DCs (Xue et al., 2014).

Embelin is a potent inhibitor of the activation of pro-inflammatory transcription factors, such as nuclear factor kappa B and signal transducer and activator of transcription 3 (STAT3; Heo et al., 2011). Embelin has been shown to inhibit the X-linked inhibitor of apoptosis protein and various inflammatory pathways (Ahn et al., 2007). In one of the study, Xue et al. (2014) demonstrated that embelin possess a strong therapeutic potential for autoimmune inflammatory conditions in MS. The study revealed the role of embelin in modulating newer regulatory mechanisms and molecular targets essential for the effectiveness in EAE. Therefore, these reports suggest that embelin could be used as a therapeutic agent to control pathological conditions, such as MS and other inflammatory autoimmune diseases, that are induced by the functional expansion of Th1 and Th17 cells.

## Traumatic Brain Injury

Traumatic brain injury (TBI) is one of the common causes of mortality in both children and young adults. Survivors have many complications like brain edema and programmed death of

neuronal cells following acute and chronic neurodegeneration. The study carried out by a team from five European institutes addresses the role and interaction of p53 and NF- $\kappa$ B-dependent mechanisms in TBI induced delayed neurodegeneration (Plesnila et al., 2007). Neuroprotection mediated by PFT is reversed by embelin in three different *in-vitro* models of neuronal cell death induced by camptothecin, glutamate, or oxygen-glucose deprivation (OGD). Embelin was used to evaluate whether enhanced X-chromosomal linked inhibitor of apoptosis (XIAP) levels is indeed involved in neuroprotection by pifithrin-a (PFT). Hence, they strongly suggest the involvement of NF- $\kappa$ B dependent regulation of XIAP in the observed neuroprotective effect (Plesnila et al., 2007).

### Hypoxia-Ischemia (HI) Induced Neurological Injury

Hypoxia-ischemia (HI) occurs when there is a deficiency in both oxygen and blood supply, which results in neonatal neurological impairment. Hill et al. (2011) tested on the caspase-dependent progression of apoptosis using embelin which is known as potent XIAP inhibitors in order to prove that sexes influences in differing pathways of cell death due to HI. So they found out that embelin inhibits XIAP by binding to BIR3 domain and thus eventually increase in cell death through a caspase-dependent pathway. Similarly, the behavioral outcomes showed that through XIAP inhibition, HI induced female rats possess severe behavioral deficits compared to HI males. These *in-vivo* data revealed that there were significant differences in severity of cognitive deficits in male infants compared to female infants with HI. This phenomenon supports the evidence of activation of caspase-independent cell death in males compared to females that activate caspase-dependent cascade following neonatal ischemia. By using embelin as XIAP inhibitor, they could conclude that gender influences cell death mechanism following HI injuries and suggest that it is very important to develop a sex-specific neuroprotection to cure HI.

### Ischemic Stroke

The majority of strokes occur when blood vessels to the brain become narrowed or clogged with fatty deposits called plaque. This cuts off blood flow to brain cells. A stroke caused by lack of blood reaching part of the brain is called an ischemic stroke. Stroke is the third major cause of mortality and the leading cause of long-term disability. Ischemic stroke accounts for ~80% of all strokes (Jauch et al., 2013). Ischemic stroke can be divided into two main types: thrombotic and embolic. Deprived of oxygen and other nutrients, the brain suffers damage as a result of the stroke. A thrombotic stroke occurs when diseased or damaged cerebral arteries become blocked by the formation of a blood clot within the brain (Rha and Saver, 2007). In order to investigate the mechanisms underlying injury after ischemic stroke as well as to develop effective therapeutic approaches to the disease, several ischemic stroke models have been developed in a variety of species. Models of stroke that can be used in rodents are becoming increasingly popular at the bench because (1) genetically-engineered animals; (2) a number of neurosensory and motor behavior outcomes; (3) fewer animal

welfare concerns. In general, there are four major types of animal models of ischemic stroke: (1) complete global cerebral ischemia; (2) incomplete global ischemia; (3) focal cerebral ischemia and (4) Multifocal cerebral ischemia (Liu and McCullough, 2011, **Figure 4**).

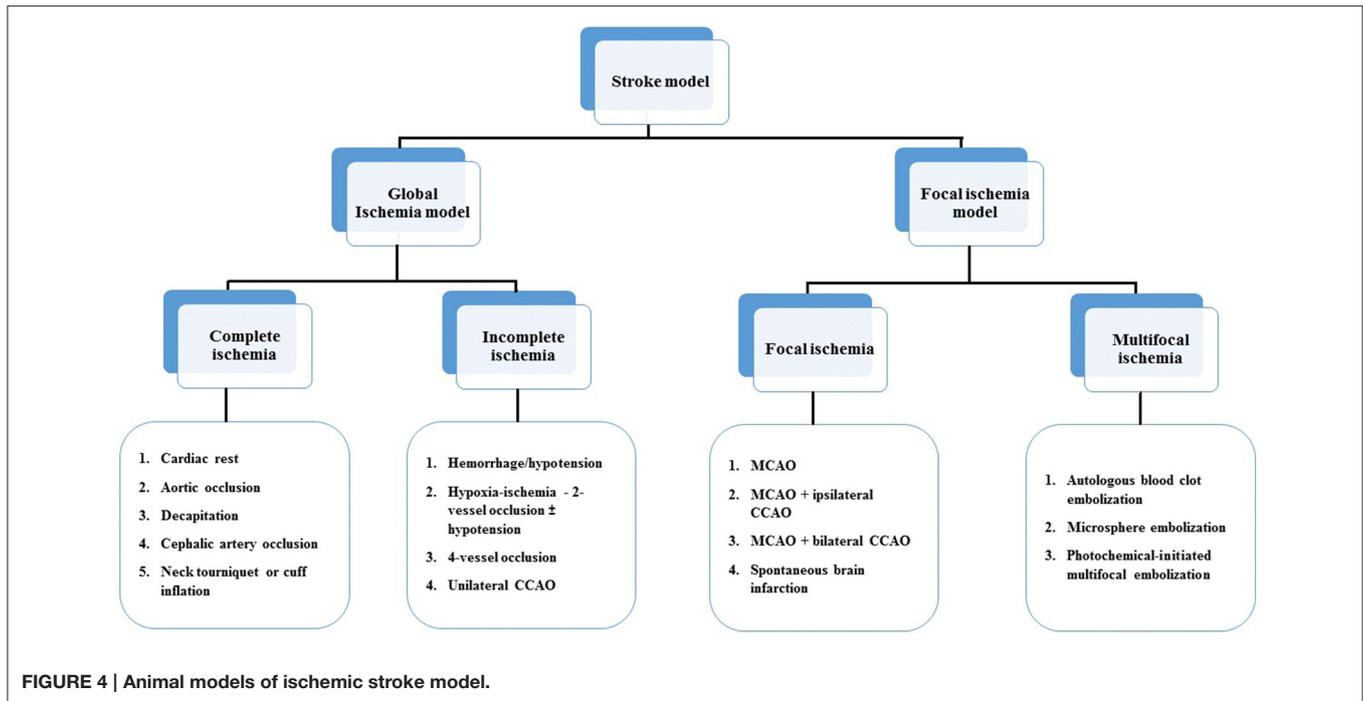
### Global Ischemia

Brain tissue and cells require oxygen and nutrients to sustain survival and contribute to standard neural operating procedures which are deprived in the a global ischemic event. This leads to the death of brain tissue or cerebral infarction/ischemic stroke which is due to poor oxygen supply or cerebral hypoxia. Siegel et al. (2011) also hypothesized that caspase dependent mechanism of ischemic cell death is also influenced by gender differences. They found that XIAP mRNA level was higher in the normal female mice brain compared to stroke induced female mice whereas no differences were observed in the male brain. They reported that embelin decreased the association between XIAP and Caspase-3 in both sexes and was acting as an XIAP inhibitor. Based on the results, larger brain infarcts were seen in embelin treated ovariectomized (Ovx) females compared to gonadally intact (GI) females. The effects of embelin on infarct exacerbation may be due to independent of circulating estrogen levels. Siegel et al. (2011) concluded that embelin treatment significantly increases stroke-induced injury in females but had no effect in males. This shows that XIAP is an important mediator of sex-specific responses after stroke.

Thippeswamy et al. (2011) investigated the protective role of embelin in transient global ischemia induced by occluding bilateral common carotid arteries followed by reperfusion. Embelin pre-treated rats significantly improved locomotion. Vestibulomotor function was assessed by beam walking test and pre-treatment with embelin significantly decreased beam walking latency when compared with ischemic control animals. Grip strength was measured using Hanging Wire test and embelin treated animals had better and longer hanging time compared to ischemic control. The behavioral observations were very well-supported by biochemical estimations, where embelin found to be modulating the lipid peroxidation, the total thiol content and glutathione-S-transferase activity in brain homogenates. Histopathological studies confirmed, decrease in the infarct size in embelin-treated animals.

### Focal Cerebral Ischemia

Cerebral ischemia is characterized by the inadequate oxygenated blood supply to the brain leads to the death of brain tissue It has been well-studied that reactive oxygen species play a key role in the pathogenesis of cerebral ischemia (Woodruff et al., 2011). Embelin is reported to have potent antioxidant activity and its chemical structure is similar to antioxidant coenzyme Q<sub>10</sub> (Matthews et al., 1998). Patel and Gohil, investigated the effect of embelin against focal cerebral ischemia using the middle cerebral artery occlusion (MCAO) model. The model of MCAO involves the insertion of a surgical filament into the external carotid artery and threading it forward into the internal carotid artery (ICA) until the tip occludes the origin of the MCA, resulting in a cessation of blood flow and subsequent brain



infarction in the MCA territory. Male Wistar rats were treated with embelin (50, 75, and 100 mg/kg, p.o.) for 20 days, followed by MCAO-induced focal cerebral ischemia and the parameters evaluated were infarct size and score. The antioxidant evaluation includes MDA, superoxide dismutase (SOD), and catalase (CAT) in brain homogenates. Embelin significantly decreased infarct size and improved infarct score. Embelin also decreased the MDA level whereas increased SOD and CAT level as compared to ischemic control group. The probable mechanisms by which embelin could be effective in cerebral ischemic condition is by the restoration of altered antioxidant enzyme activity as well as decreasing the production of lipid peroxides (Patel and Gohil, 2014). Some herbal medicines or their products having antioxidant activity have been suggested to protect against ischemic reperfusion injury, and thus justifying their use in cerebral ischemic patients.

## Brain Cancer/Apoptosis in Human Cancer Cells

### NF- $\kappa$ B Inhibition

Glioblastoma is known to be the most aggressive primary brain malignancy with median survival rates (Chou et al., 2015). Embelin is an active compound that acts an inhibitor for NF- $\kappa$ B, STAT3, XIAP, and PPAR $\gamma$  to induce growth suppression and apoptosis in human cancer cells (Park et al., 2013). Embelin is a small-molecule inhibitor of an XIAP, which has the ability to specifically inhibit XIAP of various types of a tumor cell to control and regulate the apoptosis (Wang et al., 2013). A recent finding shows that embelin also enhanced TRAIL-mediated apoptosis Allensworth et al. (2012) and thus on the basis of above reference study, Park et al. (2013) suggested that embelin may be a good anti-cancer agent with less toxicity in normal

cells. I $\kappa$ Bs regulate nuclear translocation and activation of NF- $\kappa$ B, embelin decreases phosphorylation of I $\kappa$ B $\alpha$  in a dose- and a time-dependent manner, which indicates that embelin activates I $\kappa$ B $\alpha$  that is a negative regulator of NF- $\kappa$ B. In addition, furthermore decreased NF- $\kappa$ B activity as a transcriptional activator and they found that embelin reduced nuclear translocation of NF- $\kappa$ B. Embelin suppressed proliferation of human glioma cells without affecting the normal, immortalized human astrocytes. It also has been reported to induce apoptosis in human glioma cells by inhibiting NF- $\kappa$ B which plays an important role in cell proliferation and survival of tumor. However, embelin has found to show no inhibitory effect on XIAP in glioma cells, although this active compound was discovered as an XIAP inhibitor. Besides that, overexpression of p65 was decreased in embelin induced apoptosis glioma cells. So, they concluded that embelin could be a potent novel therapeutic compound by blocking cancer cell proliferation and inducing apoptosis through NF- $\kappa$ B inhibition.

### Mitochondrial Pathway

On the other hand, in support of the above finding, Wang et al. (2013) investigated the role of mitochondrial pathway played in embelin-induced brain glioma cell apoptosis and the effect of embelin on the cell cycle. Expression of apoptosis-associated proteins, Bcl-2, Bcl-xL, Bax, and Bak, as well as cytochrome-c levels, were determined by performing western blot analysis. Embelin was found to be apoptotic to brain glioma cells in a time and dose-dependent manner. The observed effect could be due to arrest of the cell cycle in the G0/G1 phase. Changes in mitochondrial membrane potential were caused by embelin in brain glioma cell. Additionally, embelin regulated the shifting of Bax and Bcl-2 to promote the mitochondrial release

of cytochrome *c*, thus activating the caspase proteins to cause apoptosis. Thus, embelin induces apoptosis in brain glioma cells is closely associated with the mitochondrial pathway (Wang et al., 2013).

## Blood-Brain Barrier (BBB)—Cerebral Ischemia

Blood-brain barrier plays an important role in drug delivery to the CNS. Blood-brain barrier restricts, facilitates and regulates many substances from entering the CNS. It also secretes substances into the blood and the CNS (Banks, 2009). The entry of compounds across the BBB depends on their lipid solubility based on the estimation of oil/water partition coefficient (Laterra and Betz, 1999). Besides that, molecular weight, charge, tertiary structure and degree of protein binding are also among the factors in addition to lipid solubility affecting the ability of a drug to cross the BBB (Banks, 2009).

According to Pathan et al. (2009) a drug is likely to be able to transport across the BBB, if it possesses some important properties like, the compound should be in un-ionized form, partition coefficient (log P)-value should be near 2, molecular weight must be <400 Da and cumulative number of hydrogen bonds should not go beyond 8–10. According to this embelin is un-ionized molecule with log P-value of 4.83, the molecular weight is 294.38 and cumulative H bonds are 6. These properties of embelin make it permeable to BBB. So far, not a single study reported BBB permeability of embelin in *in-vitro* model. However, Siegel et al. (2011) performed *in-vivo* BBB permeability study and reported that embelin could cross the BBB. They performed liquid chromatography/tandem mass spectrometry (LC-MS/MS) on male and female sham and stroke brains. Embelin (20 mg/kg s.c.) was dosed for 3 days and it was found that the brain concentrations were elevated in both the sham and stroke mice, but the level was significantly higher in stroke mice which were close to reported IC<sub>50</sub> for embelin ( $4.1 \pm 1.1 \mu\text{M}$ ).

## Safety and Toxicity

Acute toxicity studies in mice treated with embelin 50 and 100 mg/kg oral dose showed no significant body weight change, mortality or apparent toxic effects, signifying its safety profile. This study suggests that embelin is safe on acute administration (Gupta et al., 1976). The LD<sub>50</sub>-value of embelin was reported as 44 mg/kg by i.p. route. Embelin in doses of 10 mg to 3 g/kg given orally to rats and mice did not show any toxic effects. Subacute toxicity on 10 weeks administration of 10 mg/kg of embelin to rats also indicated the drug to be free from toxic effects on heart, liver, kidney, and bone marrow, thereby having a high margin of safety in acute toxicity studies (Rathinam et al., 1976).

The toxicity of embelin has been assessed in female cyclic rats. Its administration at a dose of 120 mg/kg body weight did not cause any changes in the weight of liver, kidney, and spleen, however, the wet weight of the adrenals showed a remarkable increase. Biochemical constituents such as protein and glycogen did not show any change in these organs except in the adrenal where a significant increase was observed. The activity of acid and alkaline phosphatase was increased in the kidney and adrenal.

These toxic effects seem to be due to exposure of a very high dose i.e., 120 mg/kg, whereas LD<sub>50</sub> reported was around 44 mg/kg.

Administration of embelin for 6 weeks caused severe pathological changes in the liver and kidney which mainly included disintegration, necrotic changes, and perinuclear vacuolation. Marked tubular damage was observed in the kidneys. The adrenals showed hypertrophy and the histological features of the spleen remained unchanged (Prakash, 1994). In chronic toxicity study, the administration of embelin to Wistar rats at a dose of 50 mg/kg/day for 14 weeks did not cause any extreme drop in the blood counts but showed toxic effects on the hematopoietic cells (Sreepriya and Bali, 2006). Previous studies had also reported the non-toxic nature of embelin on hematopoietic cells when administered for 6 months in mice, rats, and monkeys (Radhakrishnan and Gnanamani, 2014).

For *in-vitro* cytotoxicity studies, embelin showed the toxic effect at 217  $\mu\text{g/ml}$  to lung fibroblasts (Feresin et al., 2003). IC<sub>50</sub> of 16.85 and 27.52  $\mu\text{M}$  of embelin was calculated against mouse lymphocytes and mouse macrophages, respectively (Sreepriya and Bali, 2006). Isolated ovarian cells were directly challenged with embelin and showed a direct effect on isolated ovarian cells (Simukoko, 2000). It did not show the toxic effect on human fibroblasts at 20  $\mu\text{g/ml}$  for 72 h in an *in-vitro* setting. Embelin was most active against sarcoma (XC) cells after 72 h of incubation (ED<sub>50</sub> 8  $\mu\text{g/ml}$ ) and slightly less active against Murine melanoma (B16) cells (ED<sub>50</sub> 13  $\mu\text{g/ml}$ ). An encouraging observation is a fact, that at these concentrations, embelin did not affect normal cells (HSF; Podolak et al., 2005).

Overall toxicity studies revealed that embelin at therapeutic doses found to be non-toxic and safe to use. Higher doses of embelin exhibit some sort of toxicity, but these doses are well above LD<sub>50</sub>-value and toxic effects are very much expected. There is also a need to carry out detailed toxicity study of embelin as per the International Council for Harmonization (ICH) safety guidelines.

## CONCLUSION AND FUTURE DIRECTIONS

Embelin is the main constituent found in the plant *E. ribes*. Embelin possesses favorable physical and chemical properties and its ability to cross the blood brain barrier make it a suitable candidate for the treatment of CNS disorders. In the present systematic review, an attempt was made to compile and discuss the efficacy of embelin against CNS complications. Embelin had been studied using various *in-vitro* prototypes and *in-vivo* animal models. It is well-reported that embelin exhibit strong anticonvulsant, anxiolytic, antidepressant properties and also improve conditions like sickness behavior, Huntington's disease, multiple sclerosis, cerebral ischemia and TBI.

Although a vast number of activities have been reported with embelin in experimental settings, there is not a single human study found on embelin related to CNS activity. None of the animal experimental outcomes was translated into human clinical research. One of the potential reasons for the non-translational research could be a lack of detailed safety and toxicity profile. Future pre-clinical and clinical trials are required

to support the safety and efficacy of this active compound. Once safety profile is established, embelin should be taken up for clinical trials. As embelin is being studied for a rich number of CNS activities, a controlled human clinical trial will open up a new horizon for this promising molecule.

## AUTHOR CONTRIBUTIONS

UK and SB has equal contribution for first author. MS, UK, and SB contributed in perceiving and designing the study. UK and

SB equally contributed with literature search and collection of data for the study. Data analysis and draft of the manuscript were completed by all authors. All the authors approved the content of the manuscript.

## ACKNOWLEDGMENTS

This work is supported by the eScience Fund of Ministry of Science, Technology and Innovation (MOSTI), Malaysia (Grant No. 06-02-10-SF0250).

## REFERENCES

- Afzal, M., Gupta, G., Kazmi, I., Rahman, M., Upadhyay, G., Ahmad, K., et al. (2012). Evaluation of anxiolytic activity of embelin isolated from *Embelia ribes*. *Biomed. Aging Pathol.* 2, 45–47. doi: 10.1016/j.biomag.2012.03.003
- Ahn, K. S., Sethi, G., and Aggarwal, B. B. (2007). Embelin, an inhibitor of X chromosome-linked inhibitor-of-apoptosis protein, blocks nuclear factor- $\kappa$ B (NF- $\kappa$ B) signaling pathway leading to suppression of NF- $\kappa$ B-regulated antiapoptotic and metastatic gene products. *Mol. Pharmacol.* 71, 209–219. doi: 10.1124/mol.106.028787
- Allensworth, J. L., Aird, K. M., Aldrich, A. J., Batinic-Haberle, I., and Devi, G. R. (2012). XIAP inhibition and generation of reactive oxygen species enhances TRAIL sensitivity in inflammatory breast cancer cells. *Mol. Cancer Ther.* 11, 1518–1527. doi: 10.1158/1535-7163.MCT-11-0787
- Banks, W. A. (2009). Characteristics of compounds that cross the blood-brain barrier. *BMC Neurol.* 9:S3. doi: 10.1186/1471-2377-9-S1-S3
- Borlongan, C. V., Koutouzis, T. K., and Sanberg, P. R. (1997). 3-Nitropropionic acid animal model and Huntington's disease. *Neurosci. Biobehav. Rev.* 21, 289–293. doi: 10.1016/S0149-7634(96)00027-9
- Bourin, M., and Hascoët, M. (2003). The mouse light/dark box test. *Eur. J. Pharmacol.* 463, 55–65. doi: 10.1016/S0014-2999(03)01274-3
- Brouillet, E. (2014). The 3-NP model of striatal neurodegeneration. *Curr. Protoc. Neurosci.* 67, 9.48.1–14. doi: 10.1002/0471142301.ns0948s67
- Cannas, A., Spissu, A., Floris, G., Congia, S., Saggi, M., Melis, M., et al. (2002). Bipolar affective disorder and Parkinson's disease: a rare, insidious and often unrecognized association. *Neurol. Sci.* 23, s67–s68. doi: 10.1007/s100720200073
- Castel-Branco, M. M., Alves, G. L., Figueiredo, I. V., Falcão, A. C., and Caramona, M. M. (2009). The maximal electroshock seizure (MES) model in the preclinical assessment of potential new antiepileptic drugs. *Methods Find. Exp. Clin. Pharmacol.* 31, 101–106. doi: 10.1358/mf.2009.31.2.1338414
- Chou, Y. C., Chang, M. Y., Wang, M. J., Harnod, T., Hung, C. H., Lee, H. T., et al. (2015). PEITC induces apoptosis of Human Brain Glioblastoma GBM8401 cells through the extrinsic- and intrinsic-signaling pathways. *Neurochem. Int.* 81, 32–40. doi: 10.1016/j.neuint.2015.01.001
- Currie, C., and World Health Organization Regional Office for Europe. (2000). *Health and Health Behaviour among Young People: Health Behaviour in School-Aged Children: A WHO Cross-National Study (HBSC)*. International Report, WHO.
- Desmond, D., Kyzar, E., Gaikwad, S., Green, J., Riehl, R., Roth, A., et al. (2012). Assessing epilepsy-related behavioral phenotypes in adult zebrafish. *Zebrafish Protoc. Neurobehav. Res.* 66, 313–322. doi: 10.1007/978-1-61779-597-8\_24
- Dhadde, S. B., Nagakannan, P., Roopesh, M., Anand Kumar, S. R., Thippeswamy, B. S., Veerapur, V. P., et al. (2016). Effect of embelin against 3-nitropropionic acid-induced Huntington's disease in rats. *Biomed. Pharmacother.* 77, 52–58. doi: 10.1016/j.biopha.2015.11.009
- Feresin, G. E., Tapia, A., Sortino, M., Zacchino, S., de Arias, A. R., Inchausti, A., et al. (2003). Bioactive alkyl phenols and embelin from Oxalis erythrorhiza. *J. Ethnopharmacol.* 88, 241–247. doi: 10.1016/S0378-8741(03)00258-7
- Gupta, G., Kazmi, I., Afzal, M., Upadhyay, G., Singh, R., and Habtemariam, S. (2013). Antidepressant-like activity of Embelin isolated from *Embelia ribes*. *Phytopharmacology* 4, 87–95.
- Gupta, O., Ali, M. M., Ray, G. B., and Atal, C. (1976). Some pharmacological investigations of embelin and its semisynthetic derivatives. *Indian J. Physiol. Pharmacol.* 21, 31–39.
- Heo, J. Y., Kim, H. J., Kim, S. M., Park, K. R., Park, S. Y., Kim, S. W., et al. (2011). Embelin suppresses STAT3 signaling, proliferation, and survival of multiple myeloma via the protein tyrosine phosphatase PTEN. *Cancer Lett.* 308, 71–80. doi: 10.1016/j.canlet.2011.04.015
- Hill, C. A., Alexander, M. L., McCullough, L. D., and Fitch, R. H. (2011). Inhibition of X-linked inhibitor of apoptosis with embelin differentially affects male versus female behavioral outcome following neonatal hypoxia-ischemia in rats. *Dev. Neurosci.* 33, 494–504. doi: 10.1159/000331651
- Jauch, E. C., Saver, J. L., Adams, H. P. Jr., Bruno, A., Connors, J. J., Demaerschalk, B. M., et al. (2013). Guidelines for the early management of patients with acute ischemic stroke: a guideline for healthcare professionals from the American Heart Association/American Stroke Association. *Stroke* 44, 870–947. doi: 10.1161/STR.0b013e318284056a
- Laterra, J. K. R., and Betz, L. A. (1999). *Blood—Cerebrospinal Fluid Barrier*, 6th Edn. Philadelphia, PA: Lippincott-Raven.
- Liu, F., and McCullough, L. D. (2011). Middle cerebral artery occlusion model in rodents: methods and potential pitfalls. *J. Biomed. Biotechnol.* 2011:464701. doi: 10.1155/2011/464701
- Lobato, K. R., Cardoso, C. C., Binfare, R. W., Budni, J., Wagner, C. L., Brocardo, P. S., et al. (2010).  $\alpha$ -Tocopherol administration produces an antidepressant-like effect in predictive animal models of depression. *Behav. Brain Res.* 209, 249–259. doi: 10.1016/j.bbr.2010.02.002
- Loma, I., and Heyman, R. (2011). Multiple sclerosis: pathogenesis and treatment. *Curr. Neuropharmacol.* 9, 409–416. doi: 10.2174/157015911796557911
- Maes, M., Berk, M., Goehler, L., Song, C., Anderson, G., Galecki, P., et al. (2012). Depression and sickness behavior are Janus-faced responses to shared inflammatory pathways. *BMC Med.* 10:66. doi: 10.1186/1741-7015-10-66
- Mahendran, S., Badami, S., Ravi, S., Thippeswamy, B. S., and Veerapur, V. P. (2011a). Synthesis and evaluation of analgesic and anti-inflammatory activities of most active free radical scavenging derivatives of embelin-A structure-activity relationship. *Chem. Pharm. Bull.* 59, 913–919. doi: 10.1248/cpb.59.913
- Mahendran, S., Thippeswamy, B., Veerapur, V., and Badami, S. (2011b). Anticonvulsant activity of embelin isolated from *Embelia ribes*. *Phytopharmacol.* 18, 186–188. doi: 10.1016/j.phymed.2010.04.002
- Matthews, R. T., Yang, L., Browne, S., Baik, M., and Beal, M. F. (1998). Coenzyme Q10 administration increases brain mitochondrial concentrations and exerts neuroprotective effects. *Proc. Natl. Acad. Sci. U.S.A.* 95, 8892–8897. doi: 10.1073/pnas.95.15.8892
- Moher, D., Liberati, A., Tetzlaff, J., and Altman, D. G. (2009). Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. *Ann. Int. Med.* 151, 264–269. doi: 10.7326/0003-4819-151-4-200908180-00135
- Moher, D., Shamseer, L., Clarke, M., Ghersi, D., Liberati, A., Petticrew, M., et al. (2015). Preferred reporting items for systematic review and meta-analysis protocols (PRISMA-P) 2015 statement. *Syst. Rev.* 4:1. doi: 10.1186/2046-4053-4-1
- Nadkarni, K. M. (1996). *[Indian Materia Medica]; Dr. KM Nadkarni's Indian Materia Medica: with Ayurvedic, Unani-Tibbi, Siddha, Allopathic, Homeopathic,*

- Naturopathic & Home Remedies, Appendices & Indexes. 1*, Vol. 1. Mumbai: Popular Prakashan.
- Park, S.-Y., Lim, S.-L., Jang, H.-J., Lee, J.-H., Um, J.-Y., Kim, S.-H., et al. (2013). Embelin induces apoptosis in human glioma cells through inactivating NF- $\kappa$ B. *J. Pharmacol. Sci.* 121, 192–199. doi: 10.1254/jphs.12137FP
- Patel, R. S., and Gohil, P. (2014). Effect of embelin in middle cerebral artery occlusion-induced focal cerebral ischemia in rats. *Oxid. Antioxid. Med. Sci.* 3, 135–139. doi: 10.5455/oams.020714.or.068
- Pathan, S. A., Iqbal, Z., Zaidi, S. M., Talegaonkar, S., Vohra, D., Jain, G. K., et al. (2009). CNS drug delivery systems: novel approaches. *Recent Pat. Drug Deliv. Formul.* 3, 71–89. doi: 10.2174/187221109787158355
- Plesnila, N., Von Baumgarten, L., Retiounskaia, M., Engel, D., Ardeshiri, A., Zimmermann, R., et al. (2007). Delayed neuronal death after brain trauma involves p53-dependent inhibition of NF- $\kappa$ B transcriptional activity. *Cell Death Differ.* 14, 1529–1541. doi: 10.1038/sj.cdd.4402159
- Podolak, I., Galanty, A., and Janeczko, Z. (2005). Cytotoxic activity of embelin from *Lysimachia punctata*. *Fitoterapia* 76, 333–335. doi: 10.1016/j.fitote.2005.02.006
- Poojari, R. (2014). Embelin-a drug of antiquity: shifting the paradigm towards modern medicine. *Expert Opin. Investig. Drugs* 23, 427–444. doi: 10.1517/13543784.2014.867016
- Prakash, A. O. (1994). Short term toxicity of embelin in female rats. *Phytother. Res.* 8, 257–264. doi: 10.1002/ptr.2650080502
- Radhakrishnan, N., Gnanamani, A., Prasad, N. R., and Mandal, A. B. (2012). Inhibition of UVB-induced oxidative damage and apoptotic biochemical changes in human lymphocytes by 2, 5-dihydroxy-3-undecyl-1, 4-benzoquinone (embelin). *Int. J. Radiat. Biol.* 88, 575–582. doi: 10.3109/09553002.2012.697644
- Radhakrishnan, N., and Gnanamani, A. (2014). 2, 5-dihydroxy-3-undecyl-1, 4-benzoquinone (Embelin)-A second solid gold of India-A Review. *Int. J. Pharm. Pharmacol. Sci.* 6, 23–30.
- Ramos, A. (2008). Animal models of anxiety: do I need multiple tests? *Trends Pharmacol. Sci.* 29, 493–498. doi: 10.1016/j.tips.2008.07.005
- Rathinam, K., Santhakumari, G., and Ramiah, N. (1976). Studies on the antifertility activity of embelin. *J. Res. Ind. Med.* 11, 84–90.
- Rha, J. H., and Saver, J. L. (2007). The impact of recanalization on ischemic stroke outcome: a meta-analysis. *Stroke* 38, 967–973. doi: 10.1161/01.STR.0000258112.14918.24
- Saraf, S. (2012). Legal regulations of complementary and alternative medicines in different countries. *Pharmacogn. Rev.* 6:154. doi: 10.4103/0973-7847.99950
- Shaikh, A., Dhadde, S. B., Durg, S., Veerapur, V. P., Badami, S., Thippeswamy, B. S., et al. (2016). Effect of embelin against lipopolysaccharide-induced sickness behaviour in mice. *Phytother. Res.* 30, 815–822. doi: 10.1002/ptr.5585
- Siegel, C., Li, J., Liu, F., Benashski, S. E., and McCullough, L. D. (2011). miR-23a regulation of X-linked inhibitor of apoptosis (XIAP) contributes to sex differences in the response to cerebral ischemia. *Proc. Natl. Acad. Sci. U.S.A.* 108, 11662–11667. doi: 10.1073/pnas.1102635108
- Simukoko, H. (2000). *The Effects of Embelin (A Benzoquinone Compound of Plant Origin) on Some Reproductive Parameters of Female Sprague-Dawley Rats* (Doctoral dissertation).
- Sreepriya, M., and Bali, G. (2006). Effects of administration of Embelin and Curcumin on lipid peroxidation, hepatic glutathione antioxidant defense and hematopoietic system during N-nitrosodiethylamine/Phenobarbital-induced hepatocarcinogenesis in Wistar rats. *Mol. Cell. Biochem.* 284, 49–55. doi: 10.1007/s11010-005-9012-7
- Thippeswamy, B. S., Nagakannan, P., Shivasharan, B. D., Mahendran, S., Veerapur, V. P., and Badami, S. (2011). Protective effect of embelin from *Embelia ribes* Burm. against transient global ischemia-induced brain damage in rats. *Neurotox Res.* 20, 379–386. doi: 10.1007/s12640-011-9258-7
- Upadhyay, R. K. (2014). Drug delivery systems, CNS protection, and the blood brain barrier. *Biomed. Res. Int.* 2014:869269. doi: 10.1155/2014/869269
- Walf, A. A., and Frye, C. A. (2007). The use of the elevated plus maze as an assay of anxiety-related behavior in rodents. *Nat. Protoc.* 2, 322–328. doi: 10.1038/nprot.2007.44
- Wang, A., Zhang, B., Zhang, J., Wu, W., and Wu, W. (2013). Embelin-induced brain glioma cell apoptosis and cell cycle arrest via the mitochondrial pathway. *Oncol. Rep.* 29, 2473–2478. doi: 10.3892/or.2013.2369
- Woodruff, T. M., Thundyil, J., Tang, S.-C., Sobey, C. G., Taylor, S. M., and Arumugam, T. V. (2011). Pathophysiology, treatment, and animal and cellular models of human ischemic stroke. *Mol. Neurodegener.* 6:11. doi: 10.1186/1750-1326-6-11
- World Health Organization (1999). *WHO Monographs on Selected Medicinal Plants*, Vol. 2. Geneva: World Health Organization.
- Xue, Z., Ge, Z., Zhang, K., Sun, R., Yang, J., Han, R., et al. (2014). Embelin suppresses dendritic cell functions and limits autoimmune encephalomyelitis through the TGF- $\beta$ / $\beta$ -catenin and STAT3 signaling pathways. *Mol. Neurobiol.* 49, 1087–1101. doi: 10.1007/s12035-013-8583-7

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2017 Kundap, Bhuvanendran, Kumari, Othman and Shaikh. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# Chapter 4

## 4.1 Introduction

Cholinergic deficit is a neuropathological condition associated with memory loss and it is interconnected with the severity of Alzheimer's disease (60). Therefore, restoration of cholinergic function by increasing the availability of acetylcholine remains a rational target for treating AD symptoms (61). Based on the cholinergic hypothesis, brain cholinergic function can be enhanced by inhibiting AChE, which will increase ACh activity (62). Scopolamine, a muscarinic antagonist had been extensively used to induce experimental models of Alzheimer's disease that affect learning and memory functions (63). A clinical observation on scopolamine treatment as a premedication for anesthesia has been reported to cause amnesia, (64) most likely caused by a blockade of cholinergic signaling. Studies have reported that scopolamine-induced amnesia model could be used to evaluate the efficacy of compounds that have the potential to be developed as therapeutic agents for AD (65).

In our study, we evaluated the neuroprotective effect of embelin on nootropics condition, as well as in scopolamine-induced amnesia rat model. We described the findings of this study in the following publication entitled 'Amelioration of Cognitive Deficit by Embelin in a Scopolamine-Induced Alzheimer's Disease-Like Condition in a Rat Model'.



# Amelioration of Cognitive Deficit by Embelin in a Scopolamine-Induced Alzheimer's Disease-Like Condition in a Rat Model

Saatheeyavaane Bhuvanendran, Yatinesh Kumari\*, Iekhsan Othman and Mohd Farooq Shaikh\*

Neuropharmacology Research Laboratory, Jeffrey Cheah School of Medicine and Health Sciences, Monash University Malaysia, Bandar Sunway, Malaysia

## OPEN ACCESS

### Edited by:

Muhammad Ayaz,  
University of Malakand, Pakistan

### Reviewed by:

Sagheer Ahmed,  
Shifa Tameer-e-Millat University,  
Pakistan  
Alla B. Salmina,  
Krasnoyarsk State Medical University  
named after

Prof. V.F.Voino-Yasenetski, Russia

### \*Correspondence:

Yatinesh Kumari  
yatinesh.kumari@monash.edu  
Mohd Farooq Shaikh  
farooq.shaikh@monash.edu;  
shaikhmohdfarooq@gmail.com

### Specialty section:

This article was submitted to  
Ethnopharmacology,  
a section of the journal  
Frontiers in Pharmacology

**Received:** 16 March 2018

**Accepted:** 04 June 2018

**Published:** 25 June 2018

### Citation:

Bhuvanendran S, Kumari Y, Othman I and Shaikh MF (2018) Amelioration of Cognitive Deficit by Embelin in a Scopolamine-Induced Alzheimer's Disease-Like Condition in a Rat Model. *Front. Pharmacol.* 9:665. doi: 10.3389/fphar.2018.00665

Embelin (2,5-dihydroxy-3-undecyl-1,4-benzoquinone) is one of the active components (2.3%) found in *Embelia ribes* Burm fruits. As determined via *in vitro* AChE inhibition assay, embelin can inhibit the acetylcholinesterase enzyme. Therefore, embelin can be utilized as a therapeutic compound, after further screening has been conducted for its use in the treatment of Alzheimer's disease (AD). In this study, the nootropic and anti-amnesic effects of embelin on scopolamine-induced amnesia in rats were evaluated. Rats were treated once daily with embelin (0.3 mg/kg, 0.6 mg/kg, 1.2 mg/kg) and donepezil (1 mg/kg) intraperitoneally (i.p.) for 17 days. During the final 9 days of treatment, a daily injection of scopolamine (1 mg/kg) was administered to induce cognitive deficits. Besides that, behavioral analysis was carried out to assess the rats' learning and memory functions. Meanwhile, hippocampal tissues were extracted for gene expression, neurotransmitter, and immunocytochemistry studies. Embelin was found to significantly improve the recognition index and memory retention in the novel object recognition (NOR) and elevated plus maze (EPM) tests, respectively. Furthermore, embelin at certain doses (0.3 mg/kg, 0.6 mg/kg, and 1.2 mg/kg) significantly exhibited a memory-enhancing effect in the absence of scopolamine, besides improving the recognition index when challenged with chronic scopolamine treatment. Moreover, in the EPM test, embelin treated rats (0.6 mg/kg) showed an increase in inflection ratio in nootropic activity. However, the increase was not significant in chronic scopolamine model. In addition, embelin contributed toward the elevated expression of BDNF, CREB1, and scavengers enzymes (SOD1 and CAT) mRNA levels. Next, pretreatment of rats with embelin mitigated scopolamine-induced neurochemical and histological changes in a manner comparable to donepezil. These research findings suggest that embelin is a nootropic compound, which also possesses an anti-amnesic ability that is displayed against scopolamine-induced memory impairment in rats. Hence, embelin could be a promising compound to treat AD.

**Keywords:** embelin, Alzheimer's disease, cognition, neuroprotective, anti-amnesic effect

## INTRODUCTION

Alzheimer's disease (AD) is known as the leading cause of dementia amongst people aged 65 and older (Ghumatkar et al., 2015). This age-related disease affects millions of individuals, and it is estimated that by 2050, 1 in 85 people worldwide will be suffering from AD (Brookmeyer et al., 2007). According to Tanzi and Bertram (2005), AD is a progressive and chronic neurodegenerative disorder which displays global cognitive decline involving memory, orientation, judgment, and reasoning. The key features of AD's pathogenesis are the gradual amassing of the protein fragment beta-amyloid (plaques) and twisted fibers of the protein tau (tangles), outside and inside neurons in the brain, respectively (Alzheimer's Association, 2017). Beta-amyloid plaques function as a neurotoxin by intervening in neuron-to-neuron communication at synapses. On the other hand, tau tangles prevent the passage of essential molecules and nutrients inside neurons, which causes axonal transport dysfunction and neuronal loss (Ali et al., 2015; Alzheimer's Association, 2017).

Apart from that, memory impairment is associated with cholinergic system dysfunction, which involves cholinergic neurons, neurotransmitters, and their receptors (Bartus et al., 1982; Lee et al., 2015). Cholinergic system dysfunction results from a loss of cholinergic neurons in the basal forebrain and hippocampus, which diminishes cognitive capability (Bartus et al., 1982; Lee et al., 2015). In healthy individuals, activation of the central cholinergic system enhances hippocampal neurogenesis through the cAMP response element-binding protein/brain-derived neurotrophic factor (CREB/BDNF) pathway (Lee et al., 2015). At present, one of the treatments for AD is a dispensation of acetylcholinesterase (AChE) inhibitors like tacrine or donepezil that increase the availability of acetylcholine at cholinergic synapses (Pandareesh et al., 2016). Moreover, oxidative stress plays an important role in AD, with some studies suggesting that beta-amyloid toxicity is linked to an increment in reactive oxygen species (ROS), including H<sub>2</sub>O<sub>2</sub> (Butterfield and Lauderback, 2002), and lipid peroxidation in neuronal cultures (Yatin et al., 1999). High oxidative stress can cause memory deficits via impairment of hippocampal synaptic plasticity (Serrano and Klann, 2004) and oxidative damage in neurodegenerative diseases (Ding et al., 2007).

Current pharmacological options for AD, only have a partial effect and poor control over the disease-causing neurons linked with Alzheimer's symptoms and lethal complications (Alzheimer's Association, 2017). As such, the available drugs in the market mainly focus on the improving memory by inhibiting the AChE enzyme (Ghumatkar et al., 2015). However, AD is not a result of a single factor like AChE, but rather is a multifactorial condition and this needs to be considered when designing a drug. Other factors such as oxidative stress and synaptic dysfunction play a significant role in the cognitive deficits in AD. Natural products could be a source of neuroprotective drugs as they can maintain normal cellular interaction in the brain and reduce the loss of neuronal functions in pathological circumstances (Hritcu et al., 2014).

Presently, many AD research groups have already explored the potential of using natural products as neuroprotective agents.

One such potential natural product is embelin (2,5-dihydroxy-3-undecyl-1,4-benzoquinone), which is the main active constituent in the fruits of *Embelia ribes* Burm (Family: Myrsinaceae), commonly known as "False Black Pepper" (Kundap et al., 2017a). The bright orange fruits of *E. ribes* have been utilized in traditional medicinal practice for treating central nervous system (CNS) disorders such as mental disorders and as a brain tonic (Poojari, 2014). Moreover, embelin has displayed anti-inflammatory, antioxidant, analgesic, antifertility, antitumor, wound healing, hepatoprotective, and antibacterial activities (Mahendran et al., 2011). Additionally, it has been reported that embelin is neuroprotective and possesses anticonvulsant ability when tested using animal models (Mahendran et al., 2011).

Embelin possesses all the features of a compound that can traverse the blood-brain barrier (BBB) and prompt a reaction in the CNS (Pathan et al., 2009; Kundap et al., 2017a). Even though embelin has various uses, there have been no studies of its neuropharmacological activities against AD-like conditions. Thus, in the present study, the anti-amnesic potential of embelin on memory deficits in a rat model of cognitive impairment caused by scopolamine was examined.

## MATERIALS AND METHODS

### Animal Care

In-house bred Sprague Dawley rats weighing between 180–200 g and between 6–8 weeks old were housed in the animal facility of the Jeffrey Cheah School of Medicine and Health Sciences, Monash University Malaysia. The rats were kept in cages and maintained under standard husbandry conditions (12:12 h light/dark cycle, controlled room temperature (23 ± 2°C), stress-free, *ad libitum* water, standard diets, and sanitary conditions). Before commencing the experiment, the rats were allowed to acclimatize for a period of 1 week to reduce stress. The Monash Animal Research Platform (MARF) Animal Ethics Committee in Australia approved all the animal experimentations conducted in this study.

### Experimental Design

#### Drug Treatment

Embelin (98%) batch number (Yucca/EM/2015/01/01) was purchased from Yucca Enterprises, Mumbai, India. The range of doses for embelin was determined based on pre-screening results. Embelin was solubilized in DMSO and then dissolved in saline. Donepezil and scopolamine were prepared in saline. Normal control rats were administered saline throughout the experiment. The treatments were given intraperitoneally (i.p) at a volume corresponding to 0.1 ml/100 g of body weight.

All experiments were performed in a balanced design (9 animals/group) to avoid being influenced by order and time. The behavioral studies were divided into two categories namely the nootropic and scopolamine models.

## Nootropic Model

- (i) **Group 1:** Control (Saline) ( $n = 9$ );
- (ii) **Group 2:** Positive control (donepezil (DPZ) 1 mg/kg);
- (iii) **Group 3:** Low dose of embelin (EMB) 0.3 mg/kg;
- (iv) **Group 4:** Medium dose of EMB 0.6 mg/kg;
- (v) **Group 5:** High dose of EMB 1.2 mg/kg

For nootropic activity, all the groups received pretreatment via the intraperitoneal route, for 8 days. All these rats were subjected to a battery of behavioral tests from day six onward until day eight for NOR and EPM (Figure 1).

## Scopolamine Model

- (i) **Group 1:** Control (Saline) ( $n = 9$ );
- (ii) **Group 2:** Negative control [scopolamine (SCP) 1 mg/kg] ( $n = 9$ );
- (iii) **Group 3:** Positive control [donepezil (DPZ) 1 mg/kg] + (SCP 1 mg/kg) ( $n = 9$ );
- (iv) **Group 4:** Low dose of embelin (EMB) 0.3 mg/kg + (SCP 1 mg/kg) ( $n = 9$ );
- (v) **Group 5:** Medium dose of EMB 0.6 mg/kg + (SCP 1 mg/kg) ( $n = 9$ );
- (vi) **Group 6:** High dose of EMB 1.2 mg/kg + (SCP 1 mg/kg) ( $n = 9$ )

For scopolamine, amnesia was induced in all the groups except the control group by daily intraperitoneal injections of scopolamine (1 mg/kg) for 9 days after embelin pretreatment (day nine to day 17). Half an hour after scopolamine administration, NOR was conducted on day 15, and EPM was carried out on day 16 and 17 of the study. At the end of the experiment, the rats were sacrificed, and their brains were isolated for further biochemical and immunohistochemistry analysis.

## Novel Object Recognition (NOR)

For the object recognition task, an open field box (40 × 40 × 20 cm) composed of black acrylic material was utilized as the experimental apparatus. This method is similar to that used by Ennaceur and Delacour (1988), with minor modifications. Besides that, behavioral testing was carried out between 9:00 am and 6:00 pm under red light illumination. The scrutinized objects were two similar transparent culture flasks containing water and a Lego toy of similar height as that of the flask (new object). Both objects types presented during the test session varied in texture, color, and size. This assessment

has three phases: (i) habituation; (ii) training, and (iii) test. On the first day, each rat was allowed to become familiarized with the open field box without the presence of an object for about 10 min. On the second day, each rat was placed in the open field for 5 min and allowed to freely explore the two identical objects (transparent cultured flask with water). After an interval of 90 min post-training session, one of the old objects used was substituted with a new object and the rats were subjected to a 2 min test run. The time spent with each object was recorded and evaluated using SMART software version 3.0. The open field box was cleaned with 70% ethanol between runs to minimize scent trails. The recognition index was calculated using the formula  $[TB/(TA + TB)*100]$  where TA and TB are time spent exploring familiar object A and novel object B respectively (Batool et al., 2016). Exploration of an object was noted when a rat sniffed or touched the object with its nose and/or forepaws.

## Elevated Plus Maze (EPM)

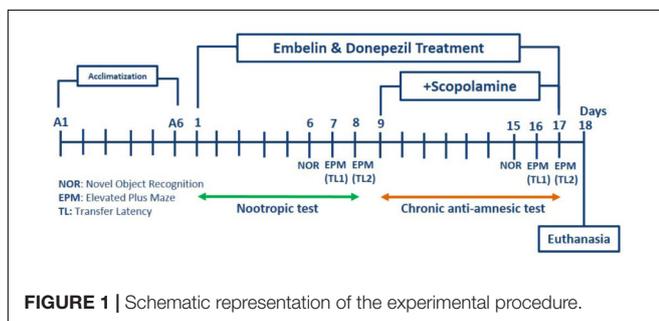
The EPM device was comprised of four arms sharing the same dimensions, i.e., two open arms (50 × 10 cm) that crossed over two closed arms with 40 cm high walls. These arms were connected using a central square (10 × 10 cm), thus giving the apparatus plus sign look. Furthermore, the EPM was elevated 50 cm above floor level. This technique is almost similar to one reported by Halder et al. (2011). The behavioral testing was conducted between 9:00 am and 6:00 pm under dim red light illumination. Assessment of memory via EPM was done in two sessions. During the training phase, each rat was placed at the end of an open arm and by using a stopwatch, transfer latency time (s), which is the time each rat took to enter (with all four paws) into either closed arm, was noted. The maze was cleaned with 70% ethanol between runs to minimize scent trails. To evaluate memory retention, a test phase was conducted 24 h (retention) after a training session. The cut-off time for each rat to explore the maze in both the phases (training and test) was 90 s. A drop in transfer latency time during test sessions was taken as an index of memory improvement.

## Tissue Processing

All the rats were sacrificed under ketamine and xylazine anesthesia 1 h after completing the behavioral test. In each group, five rat brains were fixed in 4% paraformaldehyde, and hippocampi of remaining four rats were used for real-time PCR and neurotransmitter analysis. One part of the hippocampus was used for isolation of RNA and another part of the hippocampus was homogenized on ice using methanol containing formic acid.

## Total RNA Extraction and Real-Time PCR

Total RNA was extracted from the rat brain's hippocampal region and was similar to the method used by Kundap et al. (2017b), with some minor modifications. One part of the hippocampus tissue was momentarily homogenized in Trizole solution. The mixture was extracted using chloroform and centrifuged at 13,500 rpm at 4°C. Then, the aqueous phase was precipitated with isopropanol and followed by centrifugation at 13,500 rpm at 4°C. The volume of isopropanol added was same as the volume



of the supernatant from the aqueous phase. After that, the alcohol was removed. The pellet on the other hand, was rinsed twice with 70% ethanol and resuspended in 20  $\mu$ L of RNase free water. RNA concentration was ascertained via absorbance at 260 nm using a Nanodrop machine. The total RNA (500 ng) was then reverse transcribed to synthesize cDNA using a QuantiTect<sup>®</sup> Reverse Transcription Kit, according to the manufacturer's protocol. Next, the mRNA expression of genes encoding cAMP response element-binding protein (CREB1), brain-derived neurotrophic factor (BDNF), superoxide dismutase 1 (SOD1), catalase (CAT), and IMPDH2 in the hippocampus, was measured by real-time PCR using the StepOne Real-Time PCR system. Subsequently, cDNA from the reverse transcription reaction was subjected to real-time PCR using a QuantiNova<sup>™</sup> SYBR<sup>®</sup> Green PCR kit according to manufacturer's protocol. A comparative threshold ( $C_T$ ) cycle method was applied to normalize cDNA content of samples, which involves of normalization of a number of target gene copies against the endogenous reference gene, IMPDH2.

### Neurotransmitter Analysis Using Liquid Chromatography-Tandem Mass Spectrometry (LC-MS/MS)

The brain levels of neurotransmitters like dopamine (DA), glutamate (Glu), norepinephrine (NE), and acetylcholine (ACh) were estimated using LC-MS/MS in a similar manner to that used by Kundap et al. (2017b), with some modifications. For all these standard neurotransmitters, stock solutions of 1 mg/ml were prepared in methanol (0.1% formic acid) and then stored at 4°C until use. Four calibration standards with the concentration ranges of 0.25–200.00, 250.00–20,000.00, 0.50–200.00, and 0.25–200.00 ng/mL were used for validation of DA, Glu, NE, and ACh respectively. In brief, hippocampal tissue was homogenized in ice-cold methanol containing formic acid. Then, the homogenate was vortex-mixed followed by centrifugation at 14,000 rpm for 10 min at 4°C. Finally, the supernatant was subjected to LC-MS/MS analysis, which was run on an Agilent 1290 Infinity UHPLC, coupled with an auto-sampler system comprising of Agilent 6410 Triple Quad LC/MS, ZORBAXEclipse plus C18 RRHD 2.1  $\times$  150.0 mm and 1.8-micron (P/N959759-902) column (Agilent Technologies, Santa Clara, CA, United States). The mobile phase consisted of 0.1% formic acid in (i) water (Solvent A) and (ii) acetonitrile (Solvent B). It was used with a gradient elution: (i) 0–3 min, 50% B; (ii) 3–6 min, 95% B; (iii) 6–7 min, 95% B at a flow rate of 0.1 mL/min.

### Immunohistochemical Stain Analysis

Immunohistochemical stain analysis was conducted via assessment of neurogenesis using Doublecortin (DCX) and lipid peroxidation with 4-hydroxy-2-nonenal (4HNE) staining in the hippocampus. Five brain samples from each group were immersed in 4% paraformaldehyde overnight. The samples were methodically cryoprotected in 10, 20, and 30% sucrose for 24 h. Next, the brains were embedded in 15% polyvinylpyrrolidone (PVP), frozen using dry ice, and cut

into 40  $\mu$ m frozen coronal sections using a Leica CM3050 cryostat. All sections were then stored in an anti-freeze buffer. Endogenous quenching using 1% H<sub>2</sub>O<sub>2</sub> in methanol for 30 min was performed on the free-floating sections. After washing with phosphate buffered saline (PBS), the tissues were treated with blocking buffer (1.0% bovine serum albumin in PBS and 0.3% Triton X-100) for 1 h, followed by incubation with primary DCX (1:500, Abcam) and 4HNE (1:250, Abcam) antibodies overnight at 4°C. The tissues were then incubated with a biotinylated goat anti-rabbit secondary antibody (Abcam) for 2 h after being washed with PBS. Subsequently, the tissues were exposed for 2 h to an avidin-biotin-peroxidase complex (Vectastain ABC kit, Vector). Peroxidase activity was visualized using a stable diaminobenzidine solution (DAB, Sigma). All immunoreactions were monitored via a microscope (BX41, Olympus) and using the DigiAcquis 2.0 software, results were calculated.

### Statistical Analysis

All findings were expressed as mean  $\pm$  standard error of the mean (SEM). These data were analyzed using one-way analysis of variance (ANOVA) followed by Dunnett's tests. The *P*-values of \**P* < 0.05, \*\**P* < 0.01, and \*\*\**P* < 0.001 were considered as statistically significant. All the experimental groups were compared with the SCP 1 mg/kg group.

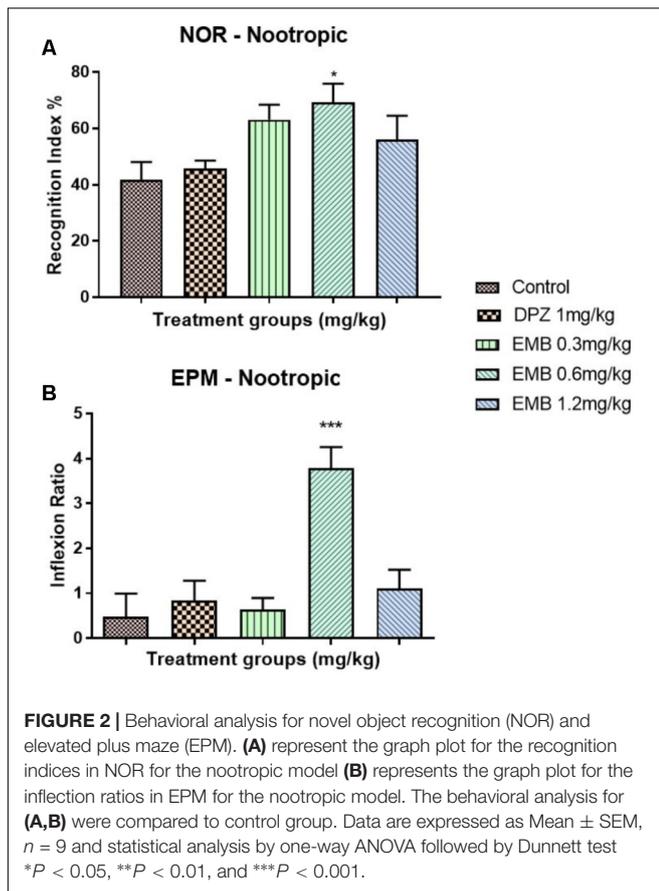
## RESULTS

### Nootropic effect of Embelin

Findings obtained from the NOR test for embelin nootropic activity are illustrated in **Figure 2A**. The effect of different embelin doses on memory function were assessed following 7 days of pretreatment. The results were expressed as recognition index (%) for the novel object. Based on the outcomes, the pretreated groups of embelin showed an increase in recognition index for novel object compared with the control group and donepezil groups. Only 0.6 mg/kg of embelin showed statistically significant results with *p* value of <0.05. In EPM, the inflection ratio was significantly increased in 0.6 mg/kg embelin treated groups when compared with the control (**Figure 2B**). There was no significant difference in other treated groups.

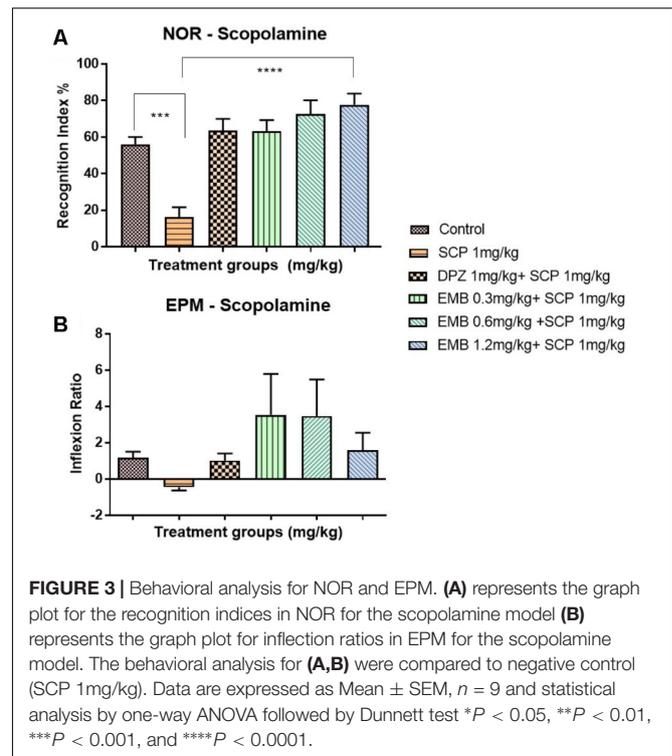
### Anti-amnesic Effect of Embelin in Rats With Scopolamine-Induced Amnesia

The NOR test showed a reduction in recognition index percentage for the negative group (SCP 1 mg/kg) in the chronic scopolamine model (**Figure 3A**). Moreover, the recognition index percentage for all the embelin treated groups were high and comparable with donepezil (1 mg/kg) group. A significant difference in the recognition index percentage was observed between all embelin treated and the negative group (*P* < 0.05). In EPM, inflection ratio analysis showed that there was an increase in retention memory in all embelin treated groups compared with the negative control group; however, it was statistically not significant (**Figure 3B**).



## Changes in mRNA Levels in the Hippocampus

BDNF mRNA levels were significantly down-regulated, approximately twofold in the hippocampus of the scopolamine group, compared with the positive control,  $P < 0.01$ . This down-regulation was ameliorated by embelin, in a dose-dependent manner, in comparison with the negative control, and a significant difference was revealed for the 1.2 mg/kg dose of embelin (Figure 4A). In addition, multiple exposures to scopolamine significantly down-regulated (twofold) the mRNA expression level of CREB1 in the negative control, compared with the positive control ( $P < 0.001$ ). Embelin treatment increased CREB1 expression level in a dose-dependent manner, compared with the negative control, and it was significant for the 1.2 mg/kg embelin dose (Figure 4B). Furthermore, scopolamine depleted antioxidant mRNA in hippocampal tissues, including (CAT) (Figure 4C) and SOD1 gene expression (Figure 4D). The down-regulation of CAT mRNA was significantly ameliorated through embelin treatment compared to the negative control for the 1.2 mg/kg embelin group ( $P < 0.05$ ). In SOD1, these changes were reversed by embelin pretreatment for all embelin treated groups, and the result was significant in the 0.6 mg/kg embelin group, with approximately a 1.5-fold change in comparison with the scopolamine treated group.

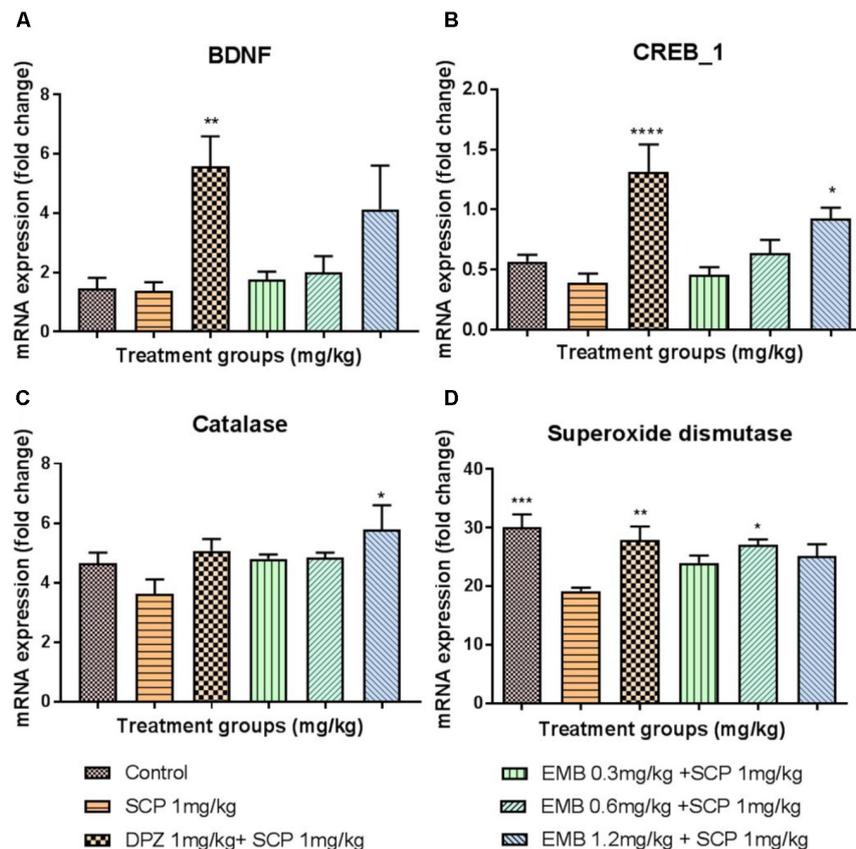


## Estimation of Neurotransmitters by LC-MS/MS

Administration of scopolamine significantly altered the levels of ACh, DA, NE, and Glu in the rat brain's hippocampus. Specifically, the level of ACh ( $P < 0.05$ ) decreased substantially whereas other neurotransmitters' levels increased significantly ( $P < 0.05$  for DA and Glu;  $P < 0.01$  for NE). Nevertheless, embelin treatment significantly normalized the level of all these neurotransmitters, and it was in a dose-dependent manner for ACh and DA (Figures 5A–D) (\* $P < 0.05$ , \*\* $P < 0.01$ , and \*\*\* $P < 0.001$ ).

## Neurogenesis and Lipid Peroxidation in the Hippocampus

Scopolamine significantly inhibited adult neurogenesis via a reduction in the distribution of dendrites and neuron bodies in the dentate gyrus (DG) region, as shown by DCX staining in the subgranular zone (SGZ) (Figure 6A). Pretreatment with embelin totally ameliorated adult neurogenesis by enhancing immature neurons in the SGZ in a dose-dependent approach in comparison with the negative control ( $P < 0.05$  for 0.3 mg/kg,  $P < 0.01$  for 0.6 mg/kg and  $P < 0.001$  for 1.2 mg/kg; Figure 6B). On the other hand, scopolamine injection significantly induced lipid peroxidation in the hippocampus, as represented by a deep brown color in the cornu ammonis 3 (CA3) regions through 4HNE staining. Pretreatment with embelin significantly lowered 4HNE-positive staining in the CA3 (threefold change) compared with the negative group ( $P < 0.0001$  for all embelin groups; Figures 7A,B). Besides that, donepezil ameliorated these



**FIGURE 4 |** Gene expression in the rat hippocampi determined by real time-PCR. The genes included are **(A)** BDNF, **(B)** CREB1, **(C)** Catalase, and **(D)** Superoxide Dismutase. All changes in the expressions levels were compared to the negative control group (SCP 1 mg/kg). Data are expressed as Mean  $\pm$  SEM,  $n = 4$  and statistical analysis by one-way ANOVA followed by Dunnett test \* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.001$ , and \*\*\*\* $P < 0.0001$ .

alterations triggered by scopolamine, as displayed through both DCX and 4-HNE staining.

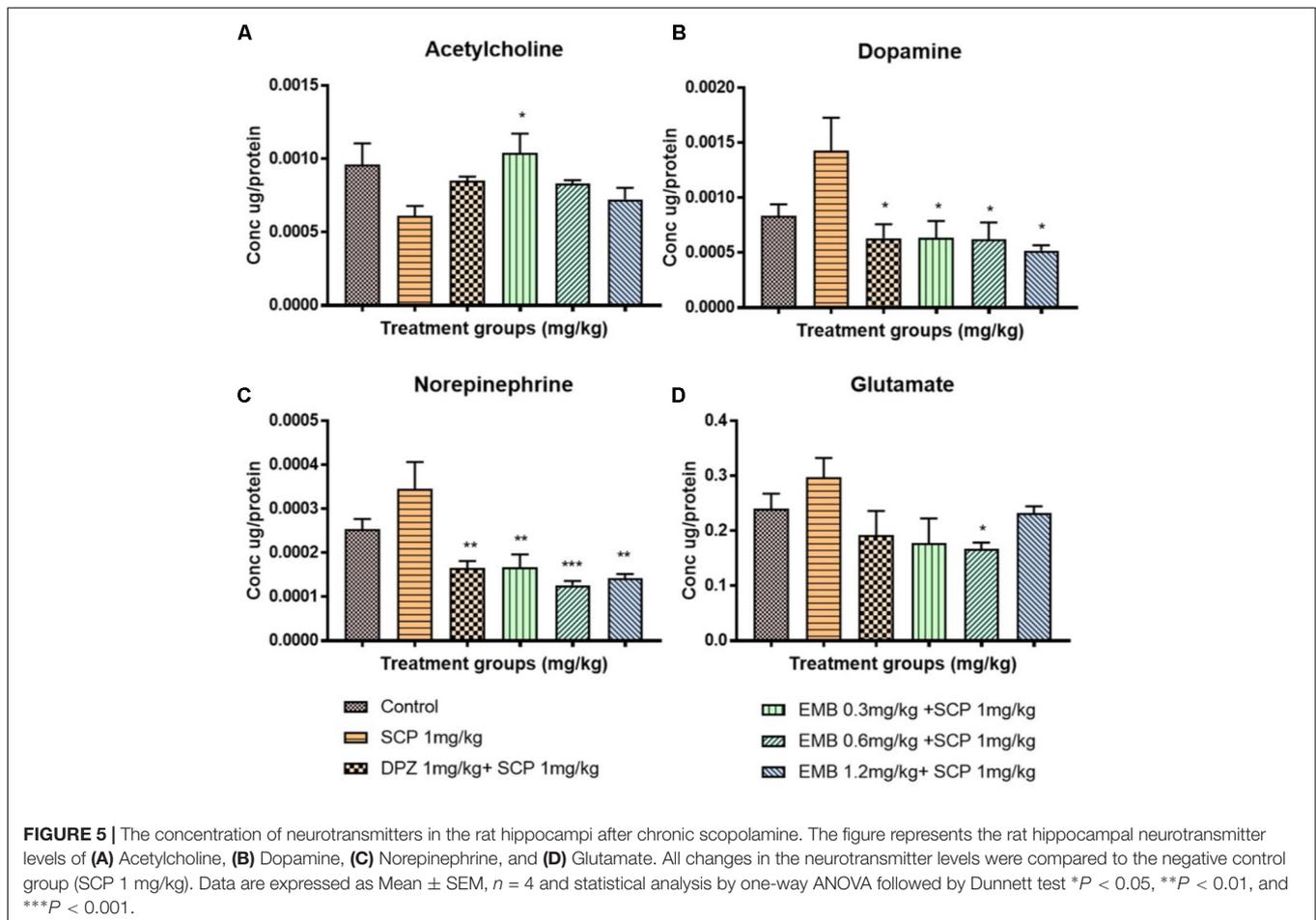
## DISCUSSION

This work aims to determine whether embelin has an anti-amnesic effect by modulating the cholinergic pathway. An animal model of hippocampal memory damage due to intraperitoneal injection of scopolamine was adopted to verify this hypothesis. The experiments comprised of two parts: Experiment 1 (pretreatment with embelin without scopolamine injection during training) to test embelin's nootropic effects on learning and memory process, and Experiment 2 (multiple exposures of scopolamine injection) to assess the effect of embelin on anti-amnesic activities and biochemical aspects during learning and memory process.

At the beginning of this experiment, we conducted a dose deciding study to find the therapeutic dose of embelin. A prior literature search determined that the range of embelin dose was between 2.5 mg/kg to 10 mg/kg for the intraperitoneal route in CNS related animal models (Mahendran et al., 2011; Afzal et al., 2012). However, our preliminary study using these

range of embelin doses resulted in a neurobehavioral effect on coordination and motor activity whereby the treated rats were immobile and kept falling from the behavioral apparatus. Thus, we decided 1.2 mg/kg as the highest dose as the LD<sub>50</sub> value for embelin was 44 mg/kg for intraperitoneal administration reported by Poojari (2014). Furthermore, we decided 0.3 mg/kg and 0.6 mg/kg would be the low dose and medium dose respectively, and all these 3 doses were effective therapeutic doses for our study as we noticed no side effects.

In this experiment, NOR and EPM were applied as behavioral models to evaluate learning and memory. The NOR test is particularly relevant in AD research as it allows the assessment of visual recognition memory, which is affected early in AD progression, involving brain regions similar to those affected by this devastating and debilitating neurodegenerative disease (Grayson et al., 2015). On the other hand, EPM is a behavioral test employed to study long-term spatial memory (Uddin et al., 2016b). Certain EPM parameters like retention transfer latency are utilized for the evaluation of memory. A decrease in transfer latency on the second day, which is after 24 h, indicates an improvement of memory and vice-versa (Dhingra and Kumar, 2012). The findings of this study showed that embelin at 0.6 mg/kg displayed nootropic activity in both the recognition

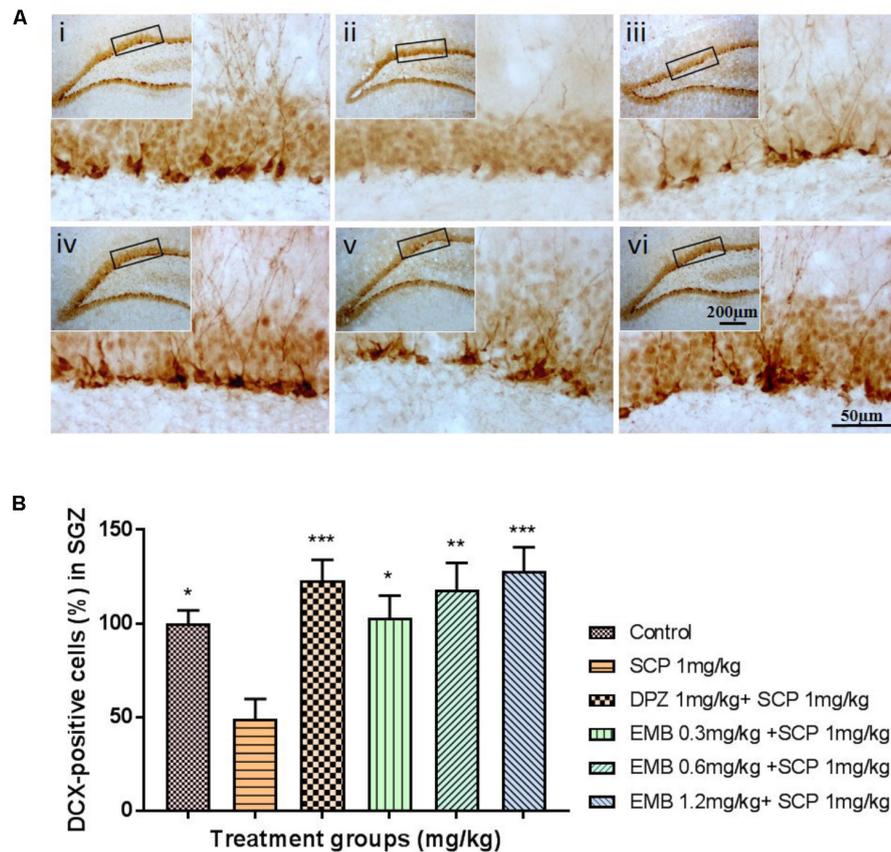


index and inflection ratio in the NOR and EPM tests, respectively (Figures 2A,B). However, the nootropic activity of embelin in both behavioral paradigms was found to be dose independent. This could be explained that at a higher dose, the drug reaches its maximum effect so increasing the drug dosage does not increase its effectiveness, but on the contrary, effectiveness decreases. This theory is supported by the fact that CNS drugs such as antipsychotic drugs produce maximum dopaminergic blockage at high doses. However, further dose increments will not produce any dopamine blockage but eventually lead to other side effects such as anticholinergic activity (Bridges, 1981). It is possible that in this experiment, the 1.2 mg/kg embelin group has reached its maximum effect and therefore cognitive ability has declined. Based on the behavioral results obtained, it can be suggested that embelin is a nootropic drug that acts as a natural cognitive enhancer. These findings show that supplementation of embelin significantly amplified the rats' memory function and 0.6 mg/kg of embelin demonstrated significant nootropics effects. Nootropic drugs are used to treat cognition deficits in patients with AD, schizophrenia, stroke, attention deficit hyperactivity disorder (ADHD), and vascular dementia (VaD) (Birks and Grimley Evans, 2009; Froestl et al., 2012).

Scopolamine-induced dementia has been used extensively to assess potential therapeutic agents for treating AD

(Kwon et al., 2009). Scopolamine is a nonselective muscarinic cholinergic receptor antagonist associated with cholinergic dysfunction, which causes performance deficits in learning and memory (Heo et al., 2014). Therefore, in this study, scopolamine was administered to rodents for 1 week to induce cholinergic neurodegeneration along with cognitive deficits. Following 6 days of scopolamine administration, the scopolamine treated group had less than 20% of the recognition index of other groups. Pretreatment with embelin ameliorated memory impairment caused by scopolamine (Figure 3A), with the recognition index being twofold more, in comparison with scopolamine treated group in a dose-dependent manner. These results exposed that embelin was as effective as the donepezil-treated group. Moreover, the findings showed that embelin treatment attenuated amnesic behavior in EPM, but it was insignificant (Figure 3B). Hence, these outcomes suggest that embelin had an anti-amnesic effect in the scopolamine model.

The brain is susceptible to oxidative stress because it consumes huge amounts of oxygen, has an abundant lipid content, and a low antioxidant level compared than other organs (Serrano and Klann, 2004). Furthermore, it is well known that the hippocampus region in the brain is crucial for learning and memory, and the formation of spatial memory (Huang et al., 2015; Lee et al., 2016). The scopolamine-induced memory



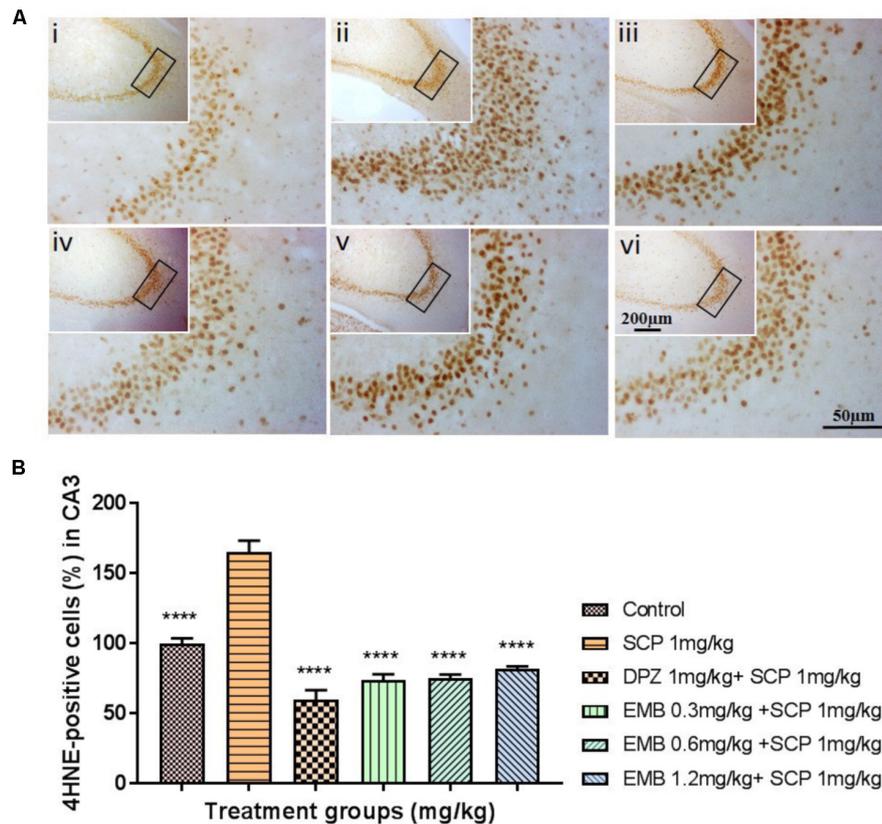
**FIGURE 6 |** DCX immunohistochemical analysis of the effects of embelin in improving scopolamine-induced suppression of neurogenesis in the dentate gyrus.

**(A)** DCX-positive staining in immature neurons is shown in the subgranular zone of the dentate gyrus. Photomicrographs of the hippocampal section of treatment groups was (i) Control (ii) SCP 1 mg/kg alone (iii) DPZ 1 mg/kg + SCP 1 mg/kg (iv) EMB 0.3 mg/kg + SCP 1 mg/kg (v) EMB 0.6 mg/kg + SCP 1 mg/kg (vi) EMB 1.2 mg/kg + SCP 1 mg/kg. Representative photomicrographs were taken at magnifications of 40X and 200X. **(B)** Quantification of DCX population. Data are expressed as means Mean  $\pm$  SEM,  $n = 5$  and statistical analysis by one-way ANOVA followed by Dunnett test \* $P < 0.05$ , \*\* $P < 0.01$ , and \*\*\* $P < 0.001$ .

deficit model demonstrated that prominent oxidative stress and memory deficits in a rodent model is similar to that in AD patients, even though the mechanism of action remains unclear (Lee et al., 2015). The change in the mRNA levels of antioxidants in the hippocampus after embelin pretreatment was examined using the scopolamine model in this present study. Scopolamine injection induced oxidative stress in the hippocampus, as evident by the decreased levels of CAT and SOD1 mRNA levels in the scopolamine alone treated negative group. To prevent or slow down the progression of free radical-mediated oxidative stress, brain antioxidant defense enzymes such as CAT and SOD play a vital role in protecting tissues against oxidative damage (Uddin et al., 2016a). Antioxidant mRNA alteration caused by scopolamine injection was significantly ameliorated for SOD1 via pretreatment with embelin. However, CAT mRNA level was decreased by scopolamine induction, but it was not significant (Figures 4C,D). Additionally, scopolamine-induced lipid peroxidation in the hippocampus's CA3 was shown as positively stained 4HNE cells. Nonetheless, pretreatment with embelin completely attenuated the over-production of 4HNE cells (Figures 7A,B). These results propose that the protective

antioxidant gene response by embelin pretreatment reduced lipid peroxidation induced by scopolamine. An increase in 4HNE cells is a key histopathological feature of neurodegenerative diseases like AD (Serrano and Klann, 2004).

Expression of BDNF and CREB1 mRNA levels in scopolamine-induced hippocampal tissue were examined to investigate the role of embelin in neurogenesis and synaptic plasticity. In this study, hippocampal BDNF and CREB1 were markedly reduced due to scopolamine injection, and pretreatment with embelin increased the mRNA expression level of both BDNF and CREB1. A high dose of embelin at 1.2 mg/kg exhibited maximum protection by increasing the levels of BDNF and CREB1. Other than that, cAMP response element binding protein (CREB) plays a crucial role in neuronal growth, proliferation, differentiation, and survival (Lee et al., 2016). In our results, the explanation for the increased in dose dependency for both BDNF and CREB 1 could possibly be that embelin may be responsible for visual recognition memory in NOR through this BDNF/CREB pathway. We noticed that at the 1.2 mg/kg dose, embelin expressed high mRNA levels of BDNF and CREB1 and this could be the reason for a 60%



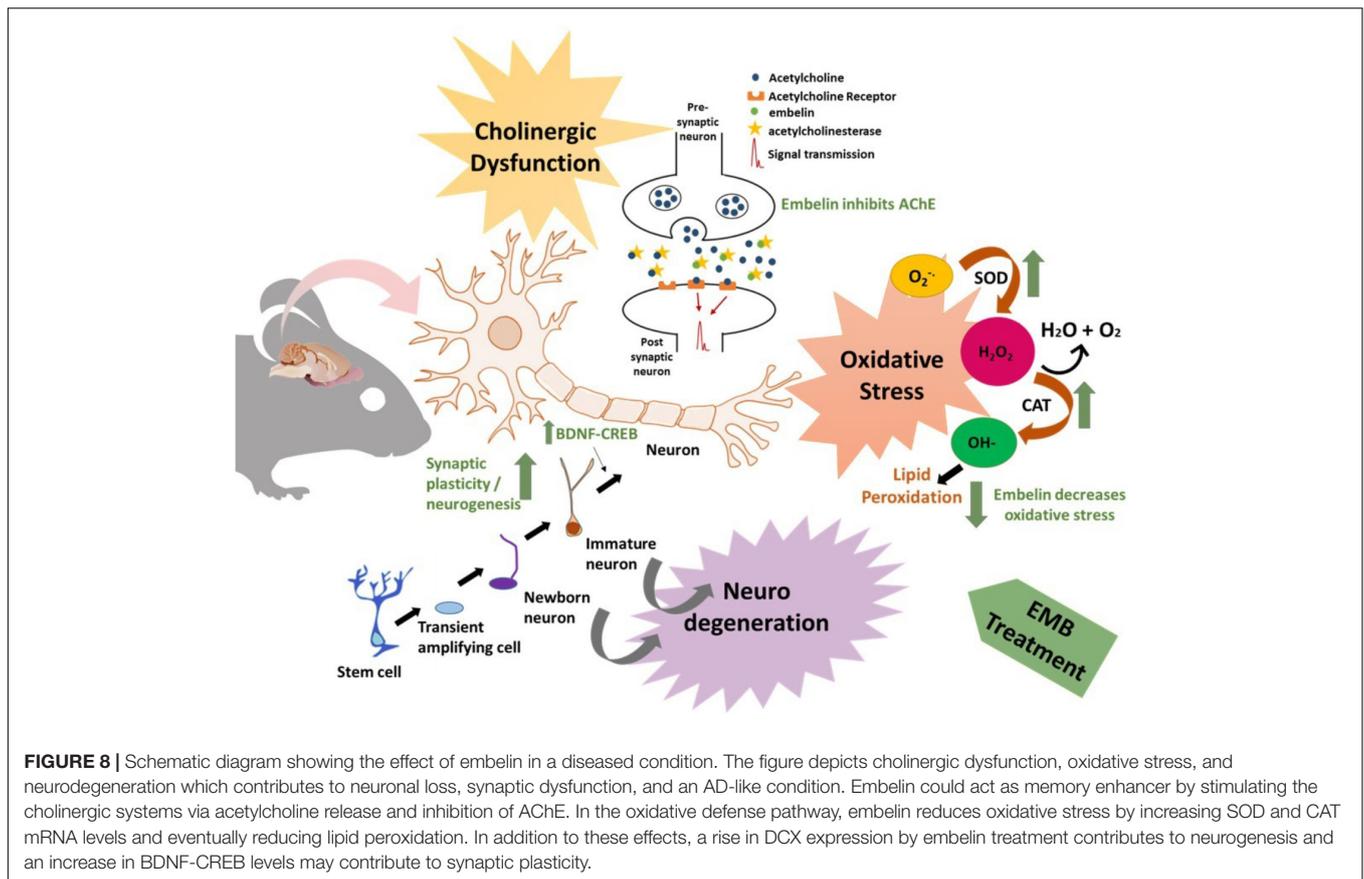
**FIGURE 7 |** 4HNE immunohistochemical analysis illustrating the inhibitory effects of embelin against scopolamine-induced lipid peroxidation in the hippocampus. **(A)** The 4HNE-positive stained cells in the CA3 region of the hippocampus are indicative of lipid peroxidation. Photomicrographs hippocampal section of treatment groups was (i) Control (ii) SCP 1 mg/kg alone (iii) DPZ 1 mg/kg + SCP 1 mg/kg (iv) EMB 0.3 mg/kg + SCP 1 mg/kg (v) EMB 0.6 mg/kg + SCP 1 mg/kg (vi) EMB 1.2 mg/kg + SCP 1 mg/kg. Representative photomicrographs were taken at magnifications of 40X, and 200X. **(B)** Quantification of 4HNE protein in CA3. Data are expressed as means Mean  $\pm$  SEM,  $n = 5$  and statistical analysis by one-way ANOVA followed by Dunnett test \* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.001$ , and \*\*\*\* $P < 0.0001$ .

increase in visual recognition index in NOR when compared with the scopolamine treated group. Thus, this validates the role of BDNF-CREB signaling in visual recognition memory, particularly for hippocampus-dependent learning.

Likewise, adult hippocampal neurogenesis plays a key role in hippocampal memory function (Mu and Gage, 2011). Altman and Das (1965) first reported on the continual production of new neurons in the adult hippocampus. These new neurons originated from adult neural stem cells (NSCs) residing in the SGZ of DG (Bonaguidi et al., 2011). In this present research, a significantly reduced level of immature neurons, revealed through DCX staining of scopolamine-induced rat hippocampus was determined, while pretreatment with embelin distinctly ameliorated repression of the SGZ region's neuronal precursor cells in a dose-dependent manner (Figures 6A,B).

Numerous studies have reported that most classical neurotransmitter systems such as ACh, NE, Glu, and DA, influence learning and memory (Myhrer, 2003). We adopted LC-MS/MS method as it is a simple, sensitive and simultaneously able to quantify the four major neurotransmitters from rat hippocampal tissue in a single run (Zheng et al., 2012). The

extraction of the neurotransmitters from rat hippocampus was done with utmost care and prior to LC-MS/MS analysis to avoid any possibilities of sample degradation and oxidation as described by He et al. (2013). The neurotransmitters' concentrations were expressed as a ratio of total protein concentration in order to get correct value and to avoid possible variation in sample when subjected to LC-MS/MS. In AD patients, pathological changes affecting glutamatergic, cholinergic, noradrenergic, and serotonergic systems have been revealed (Francis et al., 1999). In this study, the effect of embelin on brain neurotransmitter levels in rats administered scopolamine was investigated. ACh plays an essential role in learning process and memory as a key transmitter in the cholinergic system (Chen et al., 2016). A decrease in ACh levels is reported in this study as a biomarker of scopolamine-induced cognitive impairment in the rat hippocampus. Embelin administered at a dose of 0.3 mg/kg significantly increased ACh levels, subsequently improving cholinergic function. Interestingly, Arora and Deshmukh (2017) reported that embelin treatment in a streptozotocin-induced rat model decreased AChE activity, which is the enzyme that metabolizes ACh



into choline and acetate. Therefore, a reduction in AChE level indicates a high level of ACh as a result of embelin treatment, which is similar to our results. In the current research, Glu levels were raised after being treated with scopolamine. Similar outcomes were reported by Pandareesh et al. (2016) and Arora and Deshmukh (2017). Administration of 0.6 mg/kg embelin significantly lowered the level of Glu. A rise in Glu level has been reported to cause excitotoxic neuronal damage and loss of cognitive function (Arora and Deshmukh, 2017) and also associated with excitotoxicity in AD brains (Jackson, 2014). Scopolamine treatment also caused an increment in the levels of DA and NE in the hippocampus. Earlier reports suggests that an increase in DA and NE levels leads to amnesia and memory deficits. Wu et al.'s (2014) study, demonstrated that donepezil treatment can modulate the increase levels of DA and NE in disease control group. Interestingly, a similar protective effect was observed with embelin pre-treatment in amnesia condition.

In this scopolamine model, our results are unusual, with embelin causing different dose dependency in the behavioral model and neurotransmitters, particularly ACh when compared to other reported studies that utilized embelin. This could be explained by embelin being neuroprotective in a scopolamine-induced amnesia model via visual recognition memory but not in long-term spatial memory. This theory is supported by our results as there was a dose dependency in embelin treatment in NOR and the result of embelin is comparable with the donepezil group.

However, we could not see this pattern in EPM. Whilst embelin improved visual recognition in dose dependency manner, it also reduced the level of ACh in a dose-dependent manner as well. This discrepancy could be because at a dose of 0.3 mg/kg, embelin might be effectively increasing the level of ACh but stops further production of ACh at 1.2 mg/kg. At this particular dose, embelin probably plays a different role in inhibiting the enzyme AChE. This could be the reason that at 1.2 mg/kg of embelin, we observed a high recognition index in NOR of scopolamine-induced amnesia rats.

## CONCLUSION

In conclusion, the results from this study have demonstrated that embelin displays nootropic and neuroprotective abilities in scopolamine-induced amnesia in rats. Nootropic effects may be attributed to an increase in visual recognition and spatial memory in both NOR and EPM. Embelin possesses anti-amnesic effects, which could be mediated by an antioxidant gene response particularly through SOD1, the CREB-BDNF pathway, hippocampal neurogenesis, and cholinergic activity. The anti-amnesic effect of embelin is also comparable to that of donepezil at a specific concentration even though it is not in a dose-dependent manner in certain cases. Therefore, embelin could be a promising treatment for patients suffering from neurodegenerative diseases. **Figure 8** shows the potential

mechanism of action of embelin in scopolamine-induced memory impairment in rodents.

## ETHICS STATEMENT

The experimental protocol was approved by the Monash Animal Research Platform (MARF) Animal Ethics Committee, Monash University, Australia (MARF/2016/054).

## REFERENCES

- Afzal, M., Gupta, G., Kazmi, I., Rahman, M., Upadhyay, G., Ahmad, K., et al. (2012). Evaluation of anxiolytic activity of embelin isolated from *Embelia ribes*. *Biomed. Aging Pathol.* 2, 45–47. doi: 10.1016/j.biomag.2012.03.003
- Ali, T., Yoon, G. H., Shah, S. A., Lee, H. Y., and Kim, M. O. (2015). Osmotin attenuates amyloid beta-induced memory impairment, tau phosphorylation and neurodegeneration in the mouse hippocampus. *Sci. Rep.* 5:11708. doi: 10.1038/srep11708
- Altman, J., and Das, G. D. (1965). Autoradiographic and histological evidence of postnatal hippocampal neurogenesis in rats. *J. Comp. Neurol.* 124, 319–335. doi: 10.1002/cne.901240303
- Alzheimer's Association (2017). 2017 Alzheimer's disease facts and figures. *Alzheimers Dement.* 13, 325–373. doi: 10.1016/j.jalz.2017.02.001
- Arora, R., and Deshmukh, R. (2017). Embelin attenuates intracerebroventricular streptozotocin-induced behavioral, biochemical, and neurochemical abnormalities in rats. *Mol. Neurobiol.* 54, 6670–6680. doi: 10.1007/s12035-016-0182-y
- Bartus, R. T., Dean, R. L., Beer, B., and Lippa, A. S. (1982). The cholinergic hypothesis of geriatric memory dysfunction. *Science* 217, 408–414. doi: 10.1126/science.7046051
- Batool, Z., Sadiq, S., Liaquat, L., Tabassum, S., Madiha, S., Rafiq, S., et al. (2016). Repeated administration of almonds increases brain acetylcholine levels and enhances memory function in healthy rats while attenuates memory deficits in animal model of amnesia. *Brain Res. Bull.* 120, 63–74. doi: 10.1016/j.brainresbull.2015.11.001
- Birks, J., and Grimley Evans, J. (2009). Ginkgo biloba for cognitive impairment and dementia. *Cochrane Database Syst. Rev.* CD003120. doi: 10.1002/14651858.CD003120.pub3
- Bonaguidi, M. A., Wheeler, M. A., Shapiro, J. S., Stadel, R. P., Sun, G. J., Ming, G.-L., et al. (2011). In vivo clonal analysis reveals self-renewing and multipotent adult neural stem cell characteristics. *Cell* 145, 1142–1155. doi: 10.1016/j.cell.2011.05.024
- Bridges, P. K. (1981). Disturbed behavior induced by high-dose antipsychotic drugs. *Br. Med. J. (Clin. Res. Ed.)* 282:313.
- Brookmeyer, R., Johnson, E., Ziegler-Graham, K., and Arrighi, H. M. (2007). Forecasting the global burden of Alzheimer's disease. *Alzheimer's Dement.* 3, 186–191. doi: 10.1016/j.jalz.2007.04.381
- Butterfield, D. A., and Lauderback, C. M. (2002). Lipid peroxidation and protein oxidation in Alzheimer's disease brain: potential causes and consequences involving amyloid  $\beta$ -peptide-associated free radical oxidative stress1, 2. *Free Radic. Biol. Med.* 32, 1050–1060. doi: 10.1016/S0891-5849(02)00794-3
- Chen, L.-E., Wu, F., Zhao, A., Ge, H., and Zhan, H. (2016). Protection efficacy of the extract of *Ginkgo biloba* against the learning and memory damage of rats under repeated high sustained. *Evid. Based Complement. Alternat. Med.* 2016:6320586. doi: 10.1155/2016/6320586
- Dhingra, D., and Kumar, V. (2012). Memory-enhancing activity of palmatine in mice using elevated plus maze and morris water maze. *Adv. Pharmacol. Sci.* 2012:357368. doi: 10.1155/2012/357368
- Ding, Q., Dimayuga, E., and Keller, J. N. (2007). Oxidative damage, protein synthesis, and protein degradation in Alzheimer's disease. *Curr. Alzheimer Res.* 4, 73–79. doi: 10.2174/156720507779939788

## AUTHOR CONTRIBUTIONS

SB performed all the experiments and was responsible for the writing of the manuscript in its entirety. YK helped in designing gene expression study, result analysis and figures in the manuscript. IO helped in LC-MS/MS method. MS helped in conceptualizing, designing the study, result analysis, and manuscript writing. All authors gave their final approval for the submission of the manuscript.

- Ennaceur, A., and Delacour, J. (1988). A new one-trial test for neurobiological studies of memory in rats. I: behavioral data. *Behav. Brain Res.* 31, 47–59. doi: 10.1016/0166-4328(88)90157-X
- Francis, P. T., Palmer, A. M., Snape, M., and Wilcock, G. K. (1999). The cholinergic hypothesis of Alzheimer's disease: a review of progress. *J. Neurol. Neurosurg. Psychiatry* 66, 137–147. doi: 10.1136/jnnp.66.2.137
- Froestl, W., Muhs, A., and Pfeifer, A. (2012). Cognitive enhancers (nootropics). Part 1: drugs interacting with receptors. *J. Alzheimers Dis.* 32, 793–887. doi: 10.3233/JAD-2012-121186
- Ghumatkar, P. J., Patil, S. P., Jain, P. D., Tambe, R. M., and Sathaye, S. (2015). Nootropic, neuroprotective and neurotrophic effects of phloretin in scopolamine induced amnesia in mice. *Pharmacol. Biochem. Behav.* 135, 182–191. doi: 10.1016/j.pbb.2015.06.005
- Grayson, B., Leger, M., Piercy, C., Adamson, L., Harte, M., and Neill, J. C. (2015). Assessment of disease-related cognitive impairments using the novel object recognition (NOR) task in rodents. *Behav. Brain Res.* 285, 176–193. doi: 10.1016/j.bbr.2014.10.025
- Halder, S., Mehta, A. K., Kar, R., Mustafa, M., Mediratta, P. K., and Sharma, K. K. (2011). Clove oil reverses learning and memory deficits in scopolamine-treated mice. *Planta Med.* 77, 830–834. doi: 10.1055/s-0030-1250605
- He, B., Bi, K., Jia, Y., Wang, J., Lv, C., Liu, R., et al. (2013). Rapid analysis of neurotransmitters in rat brain using ultra-fast liquid chromatography and tandem mass spectrometry: application to a comparative study in normal and insomnic rats. *J. Mass Spectrom.* 48, 969–978. doi: 10.1002/jms.3243
- Heo, Y.-M., Shin, M.-S., Lee, J.-M., Kim, C.-J., Baek, S.-B., Kim, K.-H., et al. (2014). Treadmill exercise ameliorates short-term memory disturbance in scopolamine-induced amnesia rats. *Int. Neurobiol. J.* 18, 16–22. doi: 10.5213/inj.2014.18.1.16
- Hritcu, L., Noumedem, J. A., Cioanca, O., Hancianu, M., Kuete, V., and Mihasan, M. (2014). Methanolic extract of Piper nigrum fruits improves memory impairment by decreasing brain oxidative stress in amyloid beta (1–42) rat model of Alzheimer's disease. *Cell Mol. Neurobiol.* 34, 437–449. doi: 10.1007/s10571-014-0028-y
- Huang, T.-T., Leu, D., and Zou, Y. (2015). Oxidative stress and redox regulation on hippocampal-dependent cognitive functions. *Arch. Biochem. Biophys.* 576, 2–7. doi: 10.1016/j.abb.2015.03.014
- Jackson, W. S. (2014). Selective vulnerability to neurodegenerative disease: the curious case of Prion protein. *Dis. Models Mech.* 7, 21–29. doi: 10.1242/dmm.012146
- Kundap, U. P., Bhuvanendran, S., Kumari, Y., Othman, I., and Shaikh, M. F. (2017a). Plant derived phytochemical, embelin in CNS disorders: a systematic review. *Front. Pharmacol.* 8:76. doi: 10.3389/fphar.2017.00076
- Kundap, U. P., Kumari, Y., Othman, I., and Shaikh, M. F. (2017b). Zebrafish as a model for epilepsy-induced cognitive dysfunction: a pharmacological, biochemical and behavioral approach. *Front. Pharmacol.* 8:515. doi: 10.3389/fphar.2017.00515
- Kwon, S.-H., Kim, H.-C., Lee, S.-Y., and Jang, C.-G. (2009). Loganin improves learning and memory impairments induced by scopolamine in mice. *Eur. J. Pharmacol.* 619, 44–49. doi: 10.1016/j.ejphar.2009.06.062
- Lee, J.-S., Hong, S.-S., Kim, H.-G., Lee, H.-W., Kim, W.-Y., Lee, S.-K., et al. (2016). Gongjin-Dan enhances hippocampal memory in a mouse model of scopolamine-induced amnesia. *PLoS One* 11:e0159823. doi: 10.1371/journal.pone.0159823

- Lee, J.-S., Kim, H.-G., Lee, H.-W., Han, J.-M., Lee, S.-K., Kim, D.-W., et al. (2015). Hippocampal memory enhancing activity of pine needle extract against scopolamine-induced amnesia in a mouse model. *Sci. Rep.* 5:9651. doi: 10.1038/srep09651
- Mahendran, S., Thippeswamy, B., Veerapur, V., and Badami, S. (2011). Anticonvulsant activity of embelin isolated from *Embelia ribes*. *Phytomedicine* 18, 186–188. doi: 10.1016/j.phymed.2010.04.002
- Mu, Y., and Gage, F. H. (2011). Adult hippocampal neurogenesis and its role in Alzheimer's disease. *Mol. Neurodegener.* 6:85. doi: 10.1186/1750-1326-6-85
- Myhrer, T. (2003). Neurotransmitter systems involved in learning and memory in the rat: a meta-analysis based on studies of four behavioral tasks. *Brain Res. Rev.* 41, 268–287. doi: 10.1016/S0165-0173(02)00268-0
- Pandareesh, M., Anand, T., and Khanum, F. (2016). Cognition enhancing and neuromodulatory propensity of *Bacopa monniera* extract against scopolamine induced cognitive impairments in rat hippocampus. *Neurochem. Res.* 41, 985–999. doi: 10.1007/s11064-015-1780-1
- Pathan, S. A., Iqbal, Z., Zaidi, S., Talegaonkar, S., Vohra, D., Jain, G. K., et al. (2009). CNS drug delivery systems: novel approaches. *Recent Pat. Drug Deliv. Formul.* 3, 71–89. doi: 10.2174/187221109787158355
- Poojari, R. (2014). Embelin—a drug of antiquity: shifting the paradigm towards modern medicine. *Expert Opin. Investig. Drugs* 23, 427–444. doi: 10.1517/13543784.2014.867016
- Serrano, F., and Klann, E. (2004). Reactive oxygen species and synaptic plasticity in the aging hippocampus. *Ageing Res. Rev.* 3, 431–443. doi: 10.1016/j.arr.2004.05.002
- Tanzi, R. E., and Bertram, L. (2005). Twenty years of the Alzheimer's disease amyloid hypothesis: a genetic perspective. *Cell* 120, 545–555. doi: 10.1016/j.cell.2005.02.008
- Uddin, M. S., Al Mamun, A., Hossain, M. S., Ashaduzzaman, M., Noor, M. A. A., Hossain, M. S., et al. (2016a). Neuroprotective effect of *Phyllanthus acidus* L. on learning and memory impairment in scopolamine-induced animal model of dementia and oxidative stress: natural wonder for regulating the development and progression of Alzheimer's disease. *Adv. Alzheimers Dis.* 5, 53–72. doi: 10.4236/aad.2016.52005
- Uddin, M. S., Nasrullah, M., Hossain, M. S., Rahman, M. M., Sarwar, M. S., Amran, M. S., et al. (2016b). Evaluation of nootropic activity of *Persicaria flaccida* on cognitive performance, brain antioxidant markers and acetylcholinesterase activity in rats: implication for the management of Alzheimer's disease. *Am. J. Psychiatry Neurosci.* 4, 26–37. doi: 10.11648/j.ajpn.20160402.12
- Wu, C.-R., Lin, H.-C., and Su, M.-H. (2014). Reversal by aqueous extracts of *Cistanche tubulosa* from behavioral deficits in Alzheimer's disease-like rat model: relevance for amyloid deposition and central neurotransmitter function. *BMC Complement. Altern. Med.* 14:202. doi: 10.1186/1472-6882-14-202
- Yatin, S., Varadarajan, S., Link, C., and Butterfield, D. (1999). In vitro and in vivo oxidative stress associated with Alzheimer's amyloid  $\beta$ -peptide (1–42). *Neurobiol. Aging* 20, 325–330.
- Zheng, X., Kang, A., Dai, C., Liang, Y., Xie, T., Xie, L., et al. (2012). Quantitative analysis of neurochemical panel in rat brain and plasma by liquid chromatography-tandem mass spectrometry. *Anal. Chem.* 84, 10044–10051. doi: 10.1021/ac3025202

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Bhuvanendran, Kumari, Othman and Shaikh. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# Chapter 5

## **5.1 Introduction**

Neurovascular dysfunction had been linked to cognitive impairment and neurodegeneration especially in the early stages of AD (66). Chronic cerebral hypoperfusion (CCH) is a pathogenic factor in cerebrovascular diseases and neurodegenerative disorders, such as vascular dementia (67). CCH is characterized as a condition where there is insufficient blood flow to the brain, which aggravates AD-associated cognitive dysfunction (68). Rodent models of CCH were first established in experimental studies using occlusion or ligation of both common carotid arteries in rats, which allowed exploration on the early pathological events that may lead to vascular dementia (51). Moreover, studies had shown that this animal model is suitable for the further development of potential neuroprotective targets in neurodegenerative related disorders (69). Therefore in the second part of our study, we evaluated the neuroprotective effect of embelin in permanent bilateral common carotid artery occlusion (PBOCCA) model of AD.

**Detailed Status Information**

<b>Manuscript #</b>	<a href="#">SREP-18-34563</a>
<b>Current Revision #</b>	0
<b>Submission Date</b>	24th September 18
<b>Current Stage</b>	Manuscript Assigned to Peer-Reviewer/s
<b>Title</b>	Embelin Improves the Spatial Memory and Hippocampal Long-Term Potentiation in a Rat Model of Chronic Cerebral Hypoperfusion
<b>Manuscript Type</b>	Original Research
<b>Collection</b>	N/A
<b>Corresponding Author</b>	Dr. Mohd Farooq Shaikh (farooq.shaikh@monash.edu) (Monash University Malaysia)
<b>Contributing Authors</b>	Ms. Saatheeyavaane Bhuvanendran , Ms. Siti Najmi Syuhadaa Bakar , Dr. Yatinesh Kumari , Prof. Iekhsan Othman , Dr. Zurina Hassan
<b>Authorship</b>	Yes
<b>Abstract</b>	Alzheimer's disease (AD) is the second most occurring neurological disorder after stroke and is associated with cerebral hypoperfusion, possibly contributing to cognitive impairment. In the present study, neuroprotective and anti-AD effects of embelin were evaluated in chronic cerebral hypoperfusion (CCH) rat model using permanent bilateral occlusion of common carotid artery (PBOCCA) method. Rats were administered with embelin at doses of 0.3, 0.6 or 1.2 mg/kg (i.p) on day 14 post-surgery and tested for Morris water maze (MWM) followed by electrophysiological recordings to assess cognitive abilities and synaptic plasticity. The hippocampal brain regions were extracted for gene expression and neurotransmitters analysis. Treatment with embelin at the doses of 0.3 and 0.6 mg/kg significantly reversed the spatial memory impairment induced by CCH in rats. Embelin treatment has significantly protected synaptic plasticity impairment as assessed by hippocampal long-term potentiation (LTP) test. The mechanism of this study demonstrated that embelin treatment alleviated the decreased expression of BDNF, CREB1, APP, Mapt, SOD1 and NF- $\kappa$ B mRNA levels caused by CCH rats. Furthermore, treatment with embelin demonstrated neuromodulatory activity by its ability to restore hippocampal neurotransmitters. Overall these data suggest that embelin improve memory and synaptic plasticity impairment in CCH rats and can be a potential drug target for neurodegenerative disease-related cognitive disorders.
<b>Techniques</b>	Life sciences techniques, Gene expression analysis [PCR-based techniques]; Life sciences techniques [Experimental organisms]; Life sciences techniques, Experimental organisms [Rats]; Life sciences techniques, Biophysical methods [Field potential recordings];
<b>Subject Terms</b>	Biological sciences/Neuroscience/Diseases of the nervous system/Alzheimer's disease Health sciences/Molecular medicine
<b>Competing Interests Policy</b>	There is <b>NO</b> Competing Interest.

# 1 **Embelin Improves the Spatial Memory and Hippocampal Long-Term** 2 **Potentialiation in a Rat Model of Chronic Cerebral Hypoperfusion**

3 Saatheeyavaane Bhuvanendran<sup>1</sup>, Siti Najmi Syuhadaa Bakar<sup>2</sup>, Yatinesh Kumari<sup>1</sup>, Iekhsan  
4 Othman<sup>1</sup>, Mohd Farooq Shaikh<sup>1\*</sup>, Zurina Hassan<sup>2\*</sup>

5 <sup>1</sup>Neuropharmacology Research laboratory, Jeffrey Cheah School of Medicine and Health  
6 Sciences, Monash University Malaysia, Bandar Sunway, Selangor, Malaysia

7 <sup>2</sup>Centre for Drug Research, Universiti Sains Malaysia, Penang, Malaysia.

## 8 **Abstract**

9 Alzheimer's disease (AD) is the second most occurring neurological disorder after stroke and is  
10 associated with cerebral hypoperfusion, possibly contributing to cognitive impairment. In the  
11 present study, neuroprotective and anti-AD effects of embelin were evaluated in chronic cerebral  
12 hypoperfusion (CCH) rat model using permanent bilateral occlusion of common carotid artery  
13 (PBOCCA) method. Rats were administered with embelin at doses of 0.3, 0.6 or 1.2 mg/kg (i.p)  
14 on day 14 post-surgery and tested for Morris water maze (MWM) followed by electrophysiological  
15 recordings to access cognitive abilities and synaptic plasticity. The hippocampal brain regions  
16 were extracted for gene expression and neurotransmitters analysis. Treatment with embelin at the  
17 doses of 0.3 and 0.6 mg/kg significantly reversed the spatial memory impairment induced by CCH  
18 in rats. Embelin treatment has significantly protected synaptic plasticity impairment as assessed  
19 by hippocampal long-term potentiation (LTP) test. The mechanism of this study demonstrated that  
20 embelin treatment alleviated the decreased expression of BDNF, CREB1, APP, Mapt, SOD1 and  
21 NFκB mRNA levels caused by CCH rats. Furthermore, treatment with embelin demonstrated  
22 neuromodulatory activity by its ability to restore hippocampal neurotransmitters. Overall these  
23 data suggest that embelin improve memory and synaptic plasticity impairment in CCH rats and  
24 can be a potential drug target for neurodegenerative disease-related cognitive disorders.

25 **Keyword: embelin, PBOCCA, CCH, LTP, Vascular Dementia, cognitive dysfunction**

## 26 **Introduction**

27 Recognised as the second cause of age related cognitive deficits after Alzheimer's disease(AD),  
28 vascular dementia (VD) is generally a neurological disorder <sup>1</sup>. The hypothesis on vascular  
29 dementia proposed that the reduction in blood flow to the brain affects the glial and neuronal cells  
30 energy demands, thus causing neurodegeneration and brain dysfunction <sup>2</sup>. It has been discovered  
31 that this age-related neurodegenerative disorder contributed to 20% of all dementia patients, which  
32 is foreseen to be tripled by 2050 <sup>3,4</sup>. The decrease in cerebral blood flow namely chronic cerebral  
33 hypoperfusion (CCH) has been detected in cerebrovascular patients who later develop marked  
34 cognitive deficiency <sup>5</sup>.

35 In this study, CCH was induced in rodent using permanent bilateral occlusion of common carotid  
36 arteries (PBOCCA) method. According to Damodaran, et al. <sup>6</sup>, significant decrease in cerebral  
37 blood flow by 32% in the hippocampus and by 21% in the cortex was reported in PBOCCA rats  
38 which made it a suitable model for CCH <sup>7</sup>. For the duration of four days to three months (chronic  
39 phase), these rats demonstrated learning and memory impairments <sup>6</sup> followed by neuronal damage

40 and oxidative stress, which resemble the deficiencies that occur during dementia in humans <sup>8,9</sup>.  
41 Studies based on this animal model revealed the potential strategies to thwart, slow down and  
42 reverse the neurodegenerative disease progression associated with imbalance of cerebral blood  
43 flow <sup>10</sup>.

44 Synaptic integrity and plasticity are crucial for a healthy brain function especially in learning and  
45 memory <sup>11</sup>. Contrary, cognitive decline symptoms in neurological disorders including AD and  
46 dementia have been associated with synaptic plasticity impairment <sup>12</sup> by evidence of loss of  
47 synapse numbers and functions in the hippocampus <sup>13,14</sup>. Recently, numerous studies on long-term  
48 potentiation (LTP) have emerged as it is considered an indicator of synaptic plasticity at the  
49 cellular level that correlates with changes in cognitive function <sup>13</sup>. Experimental evidences have  
50 reported that by assessing long-term potentiation (LTP), neural plasticity dysfunction can be  
51 directly detected <sup>11</sup> as LTP inhibition has been observed in the CA1 region of the hippocampal in  
52 CCH rat models <sup>15,16</sup>.

53 Embelin is a promising benzoquinone compound (C<sub>17</sub>H<sub>26</sub>O<sub>4</sub>) with a molecular weight of 294.39  
54 g/mol belonging to the fruits of *Embelia ribes* Burm (Family: Myrsinaceae) <sup>17</sup>. In Indian traditional  
55 medicinal practice, the fruits of *Embelia ribes* are consumed as a brain tonic to cure disorders  
56 related to central nervous system (CNS) <sup>18</sup>. Kundap, et al. <sup>19</sup>, stated that the anticonvulsant,  
57 antidepressant and anxiolytic activities possessed by embelin have been demonstrated by many  
58 studies with the ability to improve neurological-related disorders such as Huntington's disease,  
59 multiple sclerosis, sickness behaviour, ischemia and traumatic brain injury. Recently, the  
60 neuroprotective effect of embelin in Alzheimer's disease-like condition model has been reported  
61 <sup>17,20</sup>, but specific neuroprotective mechanism of action against memory impairments in CCH-  
62 induced PBOCCA rats remain elusive.

63 Thus, this study aims at identifying whether the administration of embelin can pharmacologically  
64 ameliorate the memory impairment in PBOCCA rats. The first part of the experiment involved the  
65 behavioural effect of embelin on spatial cognitive performance using the Morris water maze  
66 (MWM), which was followed by the assessment on the effects of embelin on LTP in the CA1  
67 (Cornu ammonis 1) region of the hippocampus using *in vivo* electrophysiological recording. LTP  
68 of embelin in this study is the first to be reported, which may reveal the synaptic plasticity  
69 properties of this yellowish-orange compound as a potential therapy for Alzheimer's disease-like  
70 conditions. On the other hand, the last part of this experiment involved the extraction of rat's brain  
71 for studying their gene expression and neurotransmitter to aid in determining the potential  
72 mechanism responsible for the neuroprotective effect of embelin in vascular cognitive impairment  
73 and dementia conditions.

## 74 **Results**

### 75 **Effects of embelin on memory performance in MWM test in CCH-induced rats**

76 The MWM test was utilised in this study to evaluate the effects of embelin on the spatial memory  
77 impairment induced by CCH rats. Embelin was administered after each training session to study  
78 the post-training effect of this compound on learning and memory functions among PBOCCA rats.  
79 Figure 1A displays the swim traces of 5 groups on each test day where it was apparent that the  
80 PBOCCA rats treated with only vehicle (negative control) demonstrated longer latency to reach  
81 the submerged platform than that of the sham rats ( $p < 0.05$ ), thus indicating a poor learning  
82 disability following the PBOCCA surgery. Nevertheless, significant differences were confirmed

83 by two-way ANOVA analysis between escape latency and post-training administration of embelin  
84 due to treatment (post-training  $F_{4,86} = 5.92$ ;  $p < 0.001$ ) and test day (post-training  $F_{3,86} = 20.34$ ;  $p$   
85  $< 0.0001$ ). The relationship between embelin post-training treatment and test day was statistically  
86 significant ( $F_{12,86} = 2.121$ ;  $p < 0.05$ ). Besides, a gradual shortening of latency during the training  
87 stage in PBOCCA was noticed on rats with embelin, which showed significant enhancement with  
88 0.3 mg/kg embelin on training day 3 ( $p < 0.01$ ) and day 4 ( $p < 0.001$ ). Moreover, a significant  
89 decrease in escape latency was recorded in PBOCCA rats when receiving 0.6 mg/kg embelin on  
90 day 3 and 4 ( $p < 0.05$ ). However, the group with 1.2 mg/kg embelin displayed a decrease in escape  
91 latency on day two, but it was not significant as the performance to find the hidden platform  
92 remained constant for day 3 and 4 even after the training.

93 On the other hand, the untreated PBOCCA rats were seen unable to recall the location of platform  
94 in the probe trial on the fifth day, thus spending significantly less time in target quadrant compared  
95 to that of sham rats ( $p < 0.001$ ) (Figure 1B). A significant difference in post-training treatments  
96 with embelin ( $F_{4,19} = 8.477$ ;  $p < 0.001$ ) was noted between the groups in the percentage time spent  
97 in the target quadrant during the probe trial. PBOCCA rats with embelin of 0.3 mg/kg ( $p < 0.05$ )  
98 and 0.6 mg/kg ( $p < 0.01$ ) after the training session significantly spent longer time in the target  
99 quadrant compared to untreated PBOCCA rats, which indicates memory improvement. However,  
100 there was no significant difference in probe trial task on rats receiving 1.2 mg/kg embelin than  
101 those with PBOCCA ( $p > 0.05$ ).

#### 102 **LTP in the CA3-CA1 region of the hippocampus**

103 LTP recording was performed to investigate the synaptic plasticity in the CA3-CA1 region of the  
104 hippocampus. The normalised time course changes of fEPSP amplitude to one hr baseline period  
105 are presented in Figure 2A showing that the fEPSP amplitude of all the five groups was increased  
106 after TBS and stabilised to different levels above the baseline period. Meanwhile, two-way  
107 ANOVA analysis confirmed a significant difference in the synaptic activity in hippocampus due  
108 to treatment ( $F_{4, 288} = 57.14$ ;  $p < 0.0001$ ) and time ( $F_{17, 288} = 60.64$ ;  $p < 0.0001$ ) as well as the  
109 interaction between treatment and time ( $F_{4, 288} = 57.14$ ;  $p = 0.0148$ ). The results indicated reduced  
110 LTP in the PBOCCA group treated with vehicle alone compared to that of the sham group ( $p <$   
111  $0.0001$ ) with enhanced LTP in the PBOCCA rats treated with embelin 0.3 and 0.6 mg/kg groups  
112 ( $p < 0.0001$ ). Statistical mean values of the fEPSP amplitude for the last 2 hrs after TBS are  
113 graphically represented in Figure 2B. It was discovered that after high-frequency stimulation, LTP  
114 formation in the hippocampus has significantly inhibited PBOCCA group treated with vehicle  
115 ( $1.57 \pm 0.02$ ) than those in the sham group ( $1.96 \pm 0.04$ ;  $p < 0.0001$ ). In addition, one-way ANOVA  
116 showed significant restoration of the LTP inhibition in the vehicle group with 0.3 mg/kg ( $2.01 \pm$   
117  $0.04$ ;  $p < 0.0001$ ) and 0.6 mg/kg ( $1.83 \pm 0.06$ ;  $p = 0.001$ ) embelin in PBOCCA rats. Interestingly,  
118 1.2 mg/kg embelin treated PBOCCA ( $1.38 \pm 0.04$ ;  $p < 0.05$ ) group displayed a significant decrease  
119 in fEPSP amplitude after TBS compared to the PBOCCA group.

#### 120 **Changes in the mRNA level in the hippocampus by Real-time PCR**

121 The expression of mRNA in PBOCCA rat hippocampal tissues after embelin treatment was studied  
122 by Real-time PCR analysis showing a significant increase in APP mRNA expression of PBOCCA  
123 group ( $p < 0.0001$ ) than sham rats. In addition, a significant reduction in APP mRNA expression  
124 was observed in PBOCCA rats treated with 0.3 mg/kg ( $p < 0.001$ ) and 0.6 mg/kg ( $p < 0.01$ ) embelin  
125 compared to PBOCCA group. Meanwhile, group with 1.2 mg/kg embelin also demonstrated a  
126 significant reduction but a slight increase in APP mRNA expression compared to other embelin

127 groups ( $p < 0.05$ ) (Figure 3A). Moreover, the Mapt mRNA expression was seen increasing  
128 significantly for PBOCCA rats treated with vehicle alone compared to sham-vehicle treated group  
129 ( $p < 0.01$ ) along with a significant reduction in a dose-dependent manner for embelin treated  
130 groups (\* $p < 0.05$  and \*\*  $p < 0.01$ ). Figure 3B graphically displays the expression level of Mapt  
131 mRNA for each rat group.

132 The expression of CREB 1 mRNA level was observed significantly decreasing in PBOCCA rats  
133 than sham-vehicle treated rats ( $p < 0.01$ ). However, statistically insignificant change in the gene  
134 expression level of CREB 1 for all the embelin treated groups was recorded compared to PBOCCA  
135 group at a level of \* $\alpha = 0.05$ . Nevertheless, 0.3 and 0.6 mg/kg embelin treated groups had visibly  
136 increased in the expression level of CREB 1 compared to the PBOCCA-vehicle treated alone group  
137 (Figure 4B). On the other hand, insignificant downregulation of BDNF mRNA level was noted in  
138 PBOCCA group compared with sham groups where only 0.3 mg/kg of embelin treated PBOCCA  
139 rats presented a significant increase in BDNF mRNA expression ( $p < 0.05$ ). Besides, there was an  
140 insignificant increase in BDNF mRNA expression level in 0.6 and 1.2 mg/kg of embelin groups  
141 ( $p > 0.05$ ) when compared to PBOCCA group as shown in Figure 4A. Furthermore, Figure 5A  
142 depicts a significant depletion of the gene level of SOD1 in terms of antioxidant gene expression  
143 for the PBOCCA group compared to the healthy control group ( $p < 0.0001$ ). Additionally, all  
144 embelin treated PBOCCA rats demonstrated a significant increase in SOD1 expression with 3-fold  
145 change ( $p < 0.0001$ ) compared to PBOCCA group. On the other hand, there was a substantial rise  
146 in the gene expression level of NF $\kappa$ B for PBOCCA group compared to that of sham-vehicle treated  
147 rats ( $p < 0.05$ ). The mRNA level of NF $\kappa$ B was significantly ameliorated by embelin treatment ( $p$   
148  $< 0.05$ ) as presented in Figure 5B.

#### 149 **Estimation of neurotransmitter levels in the hippocampus by LC-MS/MS**

150 CCH-induced PBOCCA rats administered with vehicle caused a significant increase in the level  
151 of Glu ( $p < 0.0001$ ) compared to sham rats. PBOCCA rats treated with embelin (0.3, 0.6, and 1.2  
152 mg/kg, i.p.) significantly ameliorated the levels of Glu compared to PBOCCA vehicle-treated  
153 group (\*\* $p < 0.01$  and \*\*\* $p < 0.001$ ). On the other hand, levels of GABA, ACh, and 5HT in the  
154 hippocampal of PBOCCA rats were reduced in comparison to normal control group. However,  
155 only 5HT levels were detected to be significantly less in PBOCCA rats at a p-value  $< 0.0001$ .  
156 Nevertheless, embelin treatment has restored the decreased levels of GABA and 5HT compared  
157 to PBOCCA vehicle-treated rats, but were not significant. Interestingly, PBOCCA rats treated with  
158 embelin at dose 0.3 mg/kg and 0.6 mg/kg significantly attenuated the decrease in the level of ACh  
159 ( $p < 0.01$ ) compared to that of PBOCCA group. Meanwhile, embelin at dose 1.2 mg/kg group  
160 failed to restore the decreased level of ACh caused by CCH-induced PBOCCA rats as can be seen  
161 in Figure 6.

#### 162 **Discussion**

163 The therapeutic effects of embelin were found promising in many neurological-related disorders  
164 using various animal models<sup>19</sup>. Recently, embelin treatment in rodents has successfully reversed  
165 scopolamine<sup>17</sup> and streptozotocin-induced cognitive deficits<sup>20</sup> by modulating the antioxidant  
166 pathway, cholinergic activity, hippocampal neurogenesis and neuroinflammatory cytokines. Even  
167 though embelin has been found as a potential molecule in the previous findings against AD-like  
168 conditions, to date, no studies have reported the memory-improving effects of embelin in a CCH  
169 animal model. Thus, this paper is the first to report the acute effects of embelin in cognitive

170 impairment and pathophysiological transformation following two weeks of carotid arteries  
171 occlusion. The selection of dose range for embelin treatment used in this current study was  
172 determined based on previously published study<sup>17</sup>.

173 According to Farkas, et al.<sup>9</sup>, the cerebral blood flow to hippocampus was reduced by ~60%  
174 compared to control level, which progressively continued for a week with their effects remained  
175 for several months resulting from the PBOCCA surgery. Hence, PBOCCA-induced CCH in  
176 rodents could be extrapolated to human cerebral hypoperfusion resembling ageing or demented  
177 people<sup>21</sup>. Those with AD tend to have deficits in spatial abilities as they become lost in familiar  
178 places and unable to relocate the place<sup>22</sup>. Therefore, MWM has been considered as a standard  
179 method to study the spatial cognitive function in rats<sup>23</sup>. In the current study, it was discovered that  
180 the PBOCCA group showed longer escape latency during the 4-days training period, as well as  
181 during probe trials when the platform was removed from the pool. These data suggest that the  
182 PBOCCA rats had significantly impaired spatial learning and reference memory that were in line  
183 with previous studies<sup>6,24</sup>. In contrast to PBOCCA group, rats treated with embelin 0.3 and 0.6  
184 mg/kg groups were able to locate the platform much easier and faster and spent more time in the  
185 target quadrant. For the case of higher concentration, embelin at 1.2 mg/kg dose showed a longer  
186 escape latency similar to PBOCCA group. These results indicate that embelin administration with  
187 0.3 and 0.6 mg/kg doses has effectively alleviated the cognitive deficiency caused by chronic  
188 cerebral hypoperfusion.

189 Synaptic transmission and plasticity are the basis of learning and memory; thus, this present study  
190 examined whether or not the embelin administration can affect the long-term potentiation (LTP)  
191 in hippocampal of PBOCCA model. Based on results, synaptic plasticity was found significantly  
192 impaired in PBOCCA rats, which was in agreement with those of previous studies<sup>12,24,25</sup>.  
193 Interestingly, the adverse effect of chronic cerebral hypoperfusion on LTP was significantly  
194 attenuated after treatment with 0.3 mg/kg embelin. According to Rong, et al.<sup>26</sup>, enhanced LTP in  
195 the CA1 and CA3 fields of the dorsal hippocampus were due to a presynaptic mechanism of action.  
196 The result of LTP for 0.3 and 0.6 mg/kg was seen parallel with MWM performance supported by  
197 the idea that changes in synaptic efficacy underlie learning and memory processes<sup>27</sup>. Meanwhile,  
198 in the case of higher concentration, the present results are unusual with embelin 1.2 mg/kg dose  
199 displaying lower LTP than that of the PBOCCA model. The explanation for the decrease in the  
200 LTP for 1.2 mg/kg dose is similar to the previous study using scopolamine-induced memory  
201 impairment<sup>17</sup> in which the compound reached its maximum effect with the decline in cognitive  
202 ability at this dose for novel object recognition (NOR) and elevated plus maze (EPM). This  
203 experimental result is also supported by the fact that particular drug in which higher doses produce  
204 no effect or the opposite effect compared to intermediate doses<sup>28</sup>.

205 In the case of APP mRNA expression, it was found that chronic cerebral hypoperfused rats  
206 displayed significant upregulation due to occlusion of the common carotid arteries. Literature has  
207 reported that the chronic cerebral hypoperfusion resulted in an increase in protein level of APP in  
208 the hippocampus of rats<sup>29,30</sup>. Meanwhile, the APP mRNA expression was significantly  
209 downregulated by the embelin treatment in a dose-dependent manner. Moreover, significant  
210 increase in Mapt mRNA level was found in PBOCCA rats, which is consistent with the results  
211 reported by other studies<sup>31,32</sup>. Although the Mapt mRNA expression level was significantly  
212 decreased by all embelin treatments, the highest dose of embelin at 1.2 mg/kg group showed  
213 maximum protection against Mapt in CCH rats. On the other hand, the lowest dose of embelin at  
214 0.3 mg/kg group demonstrated maximum protection against APP. It can be explained from the

215 discrepancy that the downregulation of APP and Mapt by embelin maybe mediated through a  
216 different mechanism. This is another interesting outcome as this is the first time that embelin has  
217 been reported to downregulate the expression of APP and Mapt, which was directly linked to the  
218 AD.

219 To study the molecular basis of synaptic plasticity impairment, the expression of BDNF and CREB  
220 in the hippocampus of PBOCCA induced CCH rats was examined. BDNF is a crucial mediator  
221 involved in neuronal survival, development and synaptic plasticity<sup>33</sup>. Chronic cerebral  
222 hypoperfusion caused by blood flow insufficiency can cause progressive cognitive dysfunction  
223 with BDNF and CREB down-regulation. The cAMP-responsive element binding protein (CREB)-  
224 mediated transcription is needed in CNS for neuronal survival<sup>34</sup>. The transcription of the BDNF  
225 gene was CREB-regulated in an activity-dependent manner as demonstrated by several studies  
226 whereby its expression was involved in neuronal development, synaptic plasticity and  
227 neuroprotection<sup>35</sup>. In this present study, PBOCCA rats treated with vehicle alone showed a  
228 decrease in the BDNF level, which was significantly restored after treated with embelin. Moreover,  
229 it was reported that embelin at 0.3 mg/kg was able to enhance BDNF levels of the normal group  
230 in CCH rats. Besides, a similar pattern was recorded in CREB expression levels, but was  
231 insignificant compared to PBOCCA rats treated with vehicle alone. Furthermore, these outcomes  
232 were in accordance to MWM and LTP results whereby embelin at 0.3 mg/kg revealed BDNF and  
233 CREB1 upregulation as the reason for maximum improvement of spatial memory which is related  
234 to hippocampal and enhance the synaptic strength within the CA1 and CA3 neurons. Thus, it can  
235 be postulated based upon BDNF and CREB results that embelin is able to activate CREB pathway  
236 and improve BDNF mediated synaptic plasticity and neurogenesis.

237 There are several studies conducted emphasising the pathogenesis of chronic cerebral  
238 hypoperfusion and oxidative stress are caused by the implication of oxygen-derived free radicals,  
239 which play a significant role in cognitive dysfunction<sup>36</sup>. It has been proven by previous study that  
240 embelin has antioxidant properties in the hippocampus of the scopolamine model<sup>17</sup>. Therefore, it  
241 can be said that oxidative stress mechanism might have the potential to cause cognitive  
242 impairment. SOD in the mitochondrial matrix has been known as the prime line of antioxidant  
243 defence system. Hence, the mRNA expression level of SOD1 in the hippocampus was measured  
244 to assess whether or not the oxidative stress mechanism is involved in the effect of embelin. Data  
245 from this study displayed a good agreement with Zhang, et al.<sup>37</sup> from the observation of a  
246 significant decrease in SOD1 mRNA expression level of the PBOCCA rats' hippocampus.  
247 Interestingly, administrating embelin has significantly increased the SOD1 mRNA expression  
248 level, which indicates the antioxidant action of embelin that might be attributed to a direct receptor-  
249 mediated mechanism activating the downstream protein kinase signalling pathways and  
250 intracellular antioxidant enzyme systems<sup>25</sup>.

251 The NFκB was the last gene studied displaying a huge upregulation in the expression level  
252 following the CCH, which is in accordance to study reported by Fu, et al.<sup>38</sup>. Nuclear factor NFκB  
253 is a transcription factor serving a vital role in gene regulation and is applied in inflammation and  
254 oxidative stress<sup>39</sup>. In this study, the NFκB expression level in all groups was significantly reduced  
255 by embelin treatment.

256 The glutamatergic, GABAergic, cholinergic and monoaminergic neurotransmitters were observed  
257 to highly regulate the hippocampal activity, which play an imperative role in memory acquisition,  
258 consolidation and storage in the brain<sup>40,41</sup>. Kaundal, et al.<sup>42</sup> explained that significant alterations

259 in hippocampal neurotransmitters can be observed following PBOCCA surgery in rats in addition  
260 to cognitive dysfunction. In the present study, PBOCCA-induced CCH rats caused a significant  
261 elevation in hippocampal glutamate levels, which is in agreement with the results obtained in other  
262 AD-like conditions including scopolamine<sup>17,43</sup> and streptozotocin rat model<sup>20</sup>. On the contrary,  
263 the GABA levels in CCH rats were noticed to be slightly decreased but it was statistically  
264 insignificant. Moreover, previous studies recorded that imbalance levels in glutamate/GABA have  
265 caused excitotoxic neuronal damage and cognitive dysfunction in related neurological disorder<sup>41</sup>,  
266 which was similar to the present results. In the current study, embelin treatment significantly  
267 reduced hippocampal glutamate level and restored GABA level (insignificant) in CCH rats. It was  
268 also discovered that PBOCCA rats have caused a reduction in ACh level, thus indicating cognitive  
269 impairment due to the metabolism of ACh into acetate and choline by acetylcholinesterase (AChE)  
270<sup>20</sup>. Interestingly, PBOCCA rats treated with embelin 0.3 and 0.6 mg/kg significantly attenuated  
271 ACh level denoting the acetylcholinesterase inhibitor action of embelin. Moreover, these results  
272 are also comparable to probe trial performance in MWM. Furthermore, studies have revealed that  
273 a rise in the extracellular level of endogenous ACh by AChE inhibitor treatment can contribute to  
274 an increase in cerebral blood flow<sup>12,44,45</sup>. Hence, it can be explained that embelin is neuroprotective  
275 in PBOCCA-induced CCH rats via the up-regulation of cholinergic function by restoring cerebral  
276 blood flow.

277 Many studies conducted on knockout animals reported an increase in serotonin level, which  
278 improved memory performance, whereas the reduction in this neurotransmitter led to impairment  
279 in spatial memory<sup>46</sup>. Therefore, the primary function of serotonin in spatial learning and memory  
280 could be due to its involvement in cortical-hippocampal synaptic connections<sup>47</sup>. In this CCH rat  
281 model, significant decrement in serotonin level was found in the hippocampal of PBOCCA rats  
282 compared to sham rats. However, the present results showed that PBOCCA rats treated with  
283 embelin could not significantly ameliorate the serotonin levels to normal even though there was a  
284 slight increment in serotonin level when compared to that of PBOCCA rats. Hence, these outcomes  
285 suggest that embelin might modulate spatial memory through different mechanisms other than  
286 serotonin. Although the exact molecular mechanism of embelin on the expression of  
287 neurotransmitters level remained unclear, the results from this study are convincing in postulating  
288 that embelin might play a role as a neuromodulator. Thus, the improvement in cognitive functions  
289 based on MWM and LTP in this PBOCCA-induced CCH rats could be linked to the ability of  
290 embelin to modulate hippocampal neurotransmitters to a normal state. In conclusion, this study  
291 has provided novel finding where embelin treatment has mitigated the spatial memory and LTP  
292 impairment in a PBOCCA-induced CCH rat model. Embelin possessed neuroprotective and anti-  
293 AD effects that could be mediated by synaptic plasticity, antioxidant, anti-inflammatory, APP and  
294 Mapt gene response, cholinergic activity, neurochemical modulation and BDNF-CREB pathway.  
295 Thus, this current study suggests that embelin could become a potential therapeutics compound  
296 for treating cognitive disorders including VaD and AD. Figure 7 displays the potential mechanism  
297 of embelin action in CCH-induced memory impairment in rodents.

## 298 **Materials And Methods**

### 299 **Animals**

300 In-house bred male Sprague Dawley rats weighing 200-300 g and 6–8 weeks old were obtained  
301 from the Animal Research and Service Centre, Universiti Sains Malaysia (USM), Penang,  
302 Malaysia. The rodents were kept in cages maintained under standard husbandry conditions (12:12

303 h light/dark cycle, constant room temperature with no restriction on food and water supply). Prior  
304 to the employment of this study, the rats were allowed to acclimatise for one week in the transit  
305 house to reduce stress. We confirm that all animal experimentations were performed in accordance  
306 with relevant guidelines and regulations that have been approved by USM Animal Ethics  
307 Committee with the reference number USM/Animal Ethics Approval/2015/ (97) (707).

## 308 **Surgery**

309 Permanent bilateral common carotid arteries occlusion (PBOCCA) in rats was performed as  
310 previously described <sup>6</sup>. Briefly, a mixture of ketamine (80 mg/kg) and xylazine (10 mg/kg) was  
311 utilised to intraperitoneally anaesthetise all the rats. For PBOCCA surgery, the common carotid  
312 arteries were exposed by a surgical cut at the ventral midline. Both common carotid arteries were  
313 permanently ligated using a 5/0 silk suture. The skin incision was then closed, while the rats were  
314 kept in a well-ventilated room at a temperature of 25°C. The sham group was subjected to the same  
315 method without PBOCCA. All the rats were left for two weeks recovery period before being  
316 subjected to the Morris water maze and *in vivo* electrophysiology. Rats that subjected to PBOCCA  
317 surgery are susceptible to seizures, impaired vision and drastic weight loss. Thus, rats that showed  
318 any of these characteristics after PBOCCA surgery were excluded from this study. The mortality  
319 rate for rats that underwent PBOCCA surgery in this study was 20%.

## 320 **Experimental design and treatment**

321 Embelin preparation and the selection of doses for treatment were determined using the data from  
322 our previous study <sup>17</sup>. PBOCCA rats were divided into four groups. Group 1: Negative Control  
323 (PBOCCA + Vehicle) Saline with DMSO (n=6-7); Group 2: (PBOCCA + EMB 0.3mg/kg) low  
324 dose of EMB (n=6-7); Group 3: (PBOCCA + EMB 0.6mg/kg) medium dose of EMB (n=6-7);  
325 Group 4: (PBOCCA + EMB 1.2 mg/kg) high dose of EMB (n=6-7). The sham rats were labelled  
326 as Group 5 (n=6-7) and treated with the same vehicle as the negative control. Embelin and vehicle  
327 were given intraperitoneally (i.p.) at a volume corresponding to 0.1 ml/100 g of body weight from  
328 day 14 onwards. Figure 8 illustrates a schematic representation of the experimental procedure for  
329 CCH-induced PBOCCA rats.

## 330 **Behavioural Assessment**

### 331 **Morris Water Maze Test**

332 The experiment protocol for the Morris water maze was conducted adopting from Damodaran, et  
333 al. <sup>12</sup> with slight modifications. This study selected a black circular pool with 160 cm in diameter  
334 and 70 cm in height, which was placed in a test room surrounded by several visual cues. This was  
335 then followed by the addition of water into the pool to a depth of 39 cm. Besides, the pool was  
336 made opaque by the white paint added to it. The pool was divided into four quadrants, with a  
337 platform (10 cm diameter) situated 2 cm below the surface of the water in a fixed position in one  
338 quadrant while the opaque water was kept constant temperature at  $25 \pm 1^\circ\text{C}$ . Furthermore, the rats  
339 were given a pre-training session on the habituation day where they were allowed to swim freely  
340 in the pool for 60 s without a platform. All rats were put in the water at four starting points during  
341 the training session, respectively, and 60 s was set as the limit for the latency of escaping onto the  
342 platform to be recorded as a trial. Four trial were conducted daily within four consecutive days for  
343 each rat. Treatments were given intraperitoneally after each training session. It was observed on  
344 the fifth day that no platform was present; thus, each rat was subjected to a probe trial. The

345 percentage time spent in the target quadrant was used for obtaining the spatial reference memory  
346 for each rat.

### 347 **Long-Term Potentiation (LTP)**

348 The *in vivo* electrophysiological recording was conducted following the protocol proposed by  
349 Damodaran, et al.<sup>12</sup>. Briefly, urethane was utilised to anaesthetise the rats (2.0 g/kg, i.p., divided  
350 into four, 0.5 g/ kg doses every 20 min). An incision line was made to expose the skull of the rat  
351 on a stereotaxic frame. Standard stereotaxic measurements relative to bregma were utilised as the  
352 reference in drilling small holes in their skull to implant the recording electrode in the hippocampus  
353 CA1 region (AP: - 4.2 mm, mL: - 3.0 mm, V: - 3.0 mm). The bipolar stimulating electrode was  
354 placed into the Schaffer collaterals CA3 region of the hippocampus (AP: - 4.2mm, mL: + 3.0mm,  
355 V: - 4.0mm). Meanwhile, to find the stimulating intensity that could evoke 50-60 % response of  
356 its maximum extracellular field excitatory postsynaptic potentials (fEPSPs) amplitude, stimuli  
357 intensities between 0.1 and 1.0 mA with increment of 0.1 mA were delivered to the Schaffer  
358 collaterals. The stable baseline was recorded every 30 s for 1 hour. Then, single theta burst  
359 stimulation (TBS) comprising of ten bursts (each burst consisting of 5 pulses at 100 Hz) with bursts  
360 repeated every 200ms was delivered to induce LTP. The recording of fEPSPs were recorded every  
361 30 s for 2 hrs.

### 362 **Tissue Processing**

363 After the completion of the *in vivo* electrophysiological recording, all the rats were sacrificed under  
364 urethane anaesthesia with their brains extracted. Following this, the hippocampi isolation and the  
365 samples were stored at -80 °C until further analysis. For gene expression, one part of the  
366 hippocampus was transferred into 200µL ice-cold TRIzol®, whereas for neurotransmitter study,  
367 the other one was put into 200 µL ice-cold methanol with formic acid.

### 368 **Total RNA extraction and Real-Time PCR**

369 The method of Bhuvanendran, et al.<sup>17</sup> was seen similar to that of this study where the total RNA  
370 was extracted with identical Real-time PCR for the CCH- induced rat brain's hippocampal. The  
371 mRNA expression level of genes encoding brain-derived neurotrophic factor (BDNF), superoxide  
372 dismutase 1 (SOD1), amyloid precursor protein (APP), microtubule-associated protein tau (Mapt),  
373 nuclear factor kappa B (NF-κB) and IMPDH2 (Inosine Monophosphate Dehydrogenase 2)  
374 housekeeping gene was measured utilising Applied Biosystem real-time PCR. Threshold cycle  
375 (Ct) values of genes of interest was used against the Ct value of housekeeping gene to measure the  
376 expression level of five genes of interest using the formula:  $2^{(Ct \text{ value of housekeeping gene} - Ct \text{ value of gene of interest})}$ .  
377

### 378 **Neurotransmitter analysis using LC-MS/MS**

379 LC-MS/MS was used to estimate the level of neurotransmitters such as glutamate (Glu), γ-  
380 aminobutyric acid (GABA), acetylcholine (ACh) and serotonin (5HT) of CCH-induced rats. The  
381 protocol for neurotransmitter analysis has been described in details in the previous studies<sup>17</sup>. For  
382 validating Glu, GABA, ACh and 5HT, four calibration standards for neurotransmitters in  
383 concentration ranges of 250.00–20,000.00, 250.00–20,000.00, 0.25–600.00 ng/mL and 0.05–5.00  
384 ng/mL were used, respectively. The stock for the standards was prepared with methanol (0.1%  
385 formic acid) and kept at 4 °C until use. In short, one part of the hippocampus was homogenised in  
386 ice-cold methanol containing 1% formic acid followed by vortex-mixed for 60 s. The resulting

387 homogenate was then centrifuged at 14 000 rpm for 10 min at 4 °C. Finally, the resulting  
388 supernatant was put into vial inserts for the analysis of LC-MS/MS. ZORBAXE clipse plus C18  
389 RRHD 2.1 × 150.0 mm and 1.8-micron (P/N959759-902) column was utilised to separate the  
390 samples, which was then placed on Agilent 6410 Triple Quad (Agilent Technologies, Santa Clara,  
391 CA, United States) at 30 °C. The mobile phase comprising 0.1% formic acid in water (Solvent A)  
392 and acetonitrile with 0.1% formic acid (Solvent B) was used with a gradient elution of 0–3 min,  
393 50% B; 3–6 min, 95% B; 06–07 min and 95% B at a flow rate of 0.1 ml/min. MS acquisition of  
394 Glu, GABA, ACh, and 5HT was done in a positive electrospray ionisation multiple reaction  
395 monitoring (MRM) mode.

## 396 **Statistical Analysis**

397 Mean ± standard errors of the mean (SEM) were used to express the results obtained. Two-way  
398 ANOVA was used to analyse the results of acquisition trial of MWM and differences in fEPSP  
399 amplitude after TBS, which was then followed by Bonferroni's post hoc test. Statistical analysis  
400 for probe trial in MWM demonstrated the average fEPSP amplitude for 2 h recording, whereas  
401 mRNA expression and neurotransmitters levels were analysed using one-way ANOVA followed  
402 by Dunnett's post hoc test. Comparison was made for all the groups with the negative control  
403 group (PBOCCA) and found that \*p < 0.05, \*\*p < 0.01, \*\*\*p < 0.001 and \*\*\*\*p < 0.0001 were  
404 statistically significant. Additionally, the statistical analysis was carried out using GraphPad Prism  
405 software (version 7.0).

## 406 **Data Availability Statement**

407 The dataset generated from this study is available from the corresponding authors on reasonable  
408 request.

## 409 **Ethics Statement**

410 All animal experimentations conducted in this study have been approved by USM Animal Ethics  
411 Committee with the reference number USM/Animal Ethics Approval/2015/ (97) (707).

412

## 413 **References:**

- 414 1 Atzeni, F. *et al.* Rheumatic diseases and autoimmune vascular dementia. *Autoimmunity reviews*  
415 (2017).
- 416 2 de la Torre, J. C. Is Alzheimer's disease a neurodegenerative or a vascular disorder? Data,  
417 dogma, and dialectics. *The Lancet Neurology* **3**, 184-190 (2004).
- 418 3 Du, S.-Q. *et al.* Molecular mechanisms of vascular dementia: what can be learned from animal  
419 models of chronic cerebral hypoperfusion? *Molecular neurobiology* **54**, 3670-3682 (2017).
- 420 4 Rodriguez Garcia, P. L. & Rodriguez Garcia, D. Letter by Rodriguez-Garcia and Rodriguez-Garcia  
421 [corrected] regarding article, "Vascular contributions to cognitive impairment and dementia: a  
422 statement for healthcare professionals from the American Heart Association/American Stroke  
423 Association". *Stroke* **42**, e584, doi:10.1161/strokeaha.111.634279 (2011).
- 424 5 Tanaka, K.-i., Wada, N. & Ogawa, N. Chronic cerebral hypoperfusion induces transient reversible  
425 monoaminergic changes in the rat brain. *Neurochemical research* **25**, 313-320 (2000).
- 426 6 Damodaran, T. *et al.* Time course of motor and cognitive functions after chronic cerebral  
427 ischemia in rats. *Behavioural brain research* **275**, 252-258 (2014).

428 7 De la Torre, J., Fortin, T., Park, G., Pappas, B. & Richard, M. Brain blood flow restoration 'rescues'  
429 chronically damaged rat CA1 neurons. *Brain research* **623**, 6-15 (1993).

430 8 Farkas, E. *et al.* Diazoxide and dimethyl sulphoxide prevent cerebral hypoperfusion-related  
431 learning dysfunction and brain damage after carotid artery occlusion. *Brain research* **1008**, 252-  
432 260 (2004).

433 9 Farkas, E., Luiten, P. G. & Bari, F. Permanent, bilateral common carotid artery occlusion in the  
434 rat: a model for chronic cerebral hypoperfusion-related neurodegenerative diseases. *Brain*  
435 *research reviews* **54**, 162-180 (2007).

436 10 Jin, W. *et al.* Lipoxin A4 methyl ester ameliorates cognitive deficits induced by chronic cerebral  
437 hypoperfusion through activating ERK/Nrf2 signaling pathway in rats. *Pharmacology*  
438 *Biochemistry and Behavior* **124**, 145-152 (2014).

439 11 Zhao, Y. & Gong, C.-X. From chronic cerebral hypoperfusion to Alzheimer-like brain pathology  
440 and neurodegeneration. *Cellular and molecular neurobiology* **35**, 101-110 (2015).

441 12 Damodaran, T. *et al.* Clitoria ternatea L. root extract ameliorated the cognitive and hippocampal  
442 long-term potentiation deficits induced by chronic cerebral hypoperfusion in the rat. *Journal of*  
443 *Ethnopharmacology* (2018).

444 13 Xing, M., Sun, Q., Wang, Y., Cheng, Y. & Zhang, N. Hydroxysafflor yellow A increases BDNF and  
445 NMDARs in the hippocampus in a vascular dementia rat model. *Brain research* **1642**, 419-425  
446 (2016).

447 14 Scheff, S. W., Price, D. A., Schmitt, F. A., Scheff, M. A. & Mufson, E. J. Synaptic loss in the inferior  
448 temporal gyrus in mild cognitive impairment and Alzheimer's disease. *Journal of Alzheimer's*  
449 *Disease* **24**, 547-557 (2011).

450 15 Sekhon, L. H., Spence, I., Morgan, M. K. & Weber, N. C. Chronic cerebral hypoperfusion inhibits  
451 calcium-induced long-term potentiation in rats. *Stroke* **28**, 1043-1048 (1997).

452 16 Hattori, K., Naguro, I., Runchel, C. & Ichijo, H. The roles of ASK family proteins in stress  
453 responses and diseases. *Cell Communication and Signaling* **7**, 9 (2009).

454 17 Bhuvanendran, S., Kumari, Y., Othman, I. B. & Shaikh, M. Amelioration of cognitive deficit by  
455 embelin in a scopolamine-induced Alzheimer's disease-like condition in a rat model. *Frontiers in*  
456 *Pharmacology* **9**, 665 (2018).

457 18 Poojari, R. Embelin—a drug of antiquity: shifting the paradigm towards modern medicine. *Expert*  
458 *opinion on investigational drugs* **23**, 427-444 (2014).

459 19 Kundap, U. P., Bhuvanendran, S., Kumari, Y., Othman, I. & Shaikh, M. Plant derived  
460 phytocompound, embelin in CNS disorders: a systematic review. *Frontiers in pharmacology* **8**, 76  
461 (2017).

462 20 Arora, R. & Deshmukh, R. Embelin Attenuates Intracerebroventricular Streptozotocin-Induced  
463 Behavioral, Biochemical, and Neurochemical Abnormalities in Rats. *Molecular neurobiology* **54**,  
464 6670-6680 (2017).

465 21 Farkas, E. & Luiten, P. G. Cerebral microvascular pathology in aging and Alzheimer's disease.  
466 *Progress in neurobiology* **64**, 575-611 (2001).

467 22 Association, A. s. 2017 Alzheimer's disease facts and figures. *Alzheimer's & Dementia* **13**, 325-  
468 373 (2017).

469 23 D'Hooge, R. & De Deyn, P. P. Applications of the Morris water maze in the study of learning and  
470 memory. *Brain research reviews* **36**, 60-90 (2001).

471 24 Yang, C., Zhang, X., Gao, J., Wang, M. & Yang, Z. Arginine vasopressin ameliorates spatial  
472 learning impairments in chronic cerebral hypoperfusion via V1a receptor and autophagy  
473 signaling partially. *Translational psychiatry* **7**, e1174 (2017).

474 25 Yao, Y. *et al.* Bombesin attenuated ischemia-induced spatial cognitive and synaptic plasticity  
475 impairment associated with oxidative damage. *Biomedicine & Pharmacotherapy* **103**, 87-93  
476 (2018).

477 26 Rong, X., Chen, X. & Du, Y.-C. Potentiation of synaptic transmission by neuropeptide AVP4-8  
478 (ZNC (C) PR) in rat hippocampal slices. *Neuroreport* **4**, 1135-1138 (1993).

479 27 Martin, S. & Morris, R. New life in an old idea: the synaptic plasticity and memory hypothesis  
480 revisited. *Hippocampus* **12**, 609-636 (2002).

481 28 Roesler, R., Kent, P., Luft, T., Schwartzmann, G. & Merali, Z. Gastrin-releasing peptide receptor  
482 signaling in the integration of stress and memory. *Neurobiology of learning and memory* **112**,  
483 44-52 (2014).

484 29 Bennett, S. *et al.* Cleavage of amyloid precursor protein elicited by chronic cerebral  
485 hypoperfusion. *Neurobiology of aging* **21**, 207-214 (2000).

486 30 Ai, J. *et al.* MicroRNA-195 protects against dementia induced by chronic brain hypoperfusion via  
487 its anti-amyloidogenic effect in rats. *Journal of Neuroscience* **33**, 3989-4001 (2013).

488 31 Li, J.-F., Wang, Z., Sun, Q.-J. & Du, Y.-F. Expression of tau protein in rats with cognitive  
489 dysfunction induced by cerebral hypoperfusion. *International journal of clinical and*  
490 *experimental medicine* **8**, 19682 (2015).

491 32 Cao, Y. *et al.* The effect of *Scutellaria baicalensis* stem-leaf flavonoids on spatial learning and  
492 memory in chronic cerebral ischemia-induced vascular dementia of rats. *Acta Biochim Biophys*  
493 *Sin* **48**, 437-446 (2016).

494 33 Li, Q., Cui, J., Fang, C., Zhang, X. & Li, L. S-adenosylmethionine Administration Attenuates Low  
495 Brain-Derived Neurotrophic Factor Expression Induced by Chronic Cerebrovascular  
496 Hypoperfusion or Beta Amyloid Treatment. *Neuroscience bulletin* **32**, 153-161 (2016).

497 34 Fusco, S. *et al.* A role for neuronal cAMP responsive-element binding (CREB)-1 in brain responses  
498 to calorie restriction. *Proceedings of the National Academy of Sciences* **109**, 621-626 (2012).

499 35 Sakamoto, K., Karelina, K. & Obrietan, K. CREB: a multifaceted regulator of neuronal plasticity  
500 and protection. *Journal of neurochemistry* **116**, 1-9 (2011).

501 36 Bennett, S., Grant, M. M. & Aldred, S. Oxidative stress in vascular dementia and Alzheimer's  
502 disease: a common pathology. *Journal of Alzheimer's Disease* **17**, 245-257 (2009).

503 37 Zhang, J. *et al.* Puerarin attenuates cognitive dysfunction and oxidative stress in vascular  
504 dementia rats induced by chronic ischemia. *International journal of clinical and experimental*  
505 *pathology* **8**, 4695 (2015).

506 38 Fu, X. *et al.* Protective role of luteolin against cognitive dysfunction induced by chronic cerebral  
507 hypoperfusion in rats. *Pharmacology Biochemistry and Behavior* **126**, 122-130 (2014).

508 39 Schreck, R., Albermann, K. & Baeuerle, P. A. Nuclear factor  $\kappa$ B: an oxidative stress-responsive  
509 transcription factor of eukaryotic cells (a review). *Free radical research communications* **17**, 221-  
510 237 (1992).

511 40 Heii, A., Yosuke, I., Kenji, K., Takashi, M. & Reiji, I. Neurotransmitter changes in early-and late-  
512 onset Alzheimer-type dementia. *Progress in Neuro-Psychopharmacology and Biological*  
513 *Psychiatry* **16**, 883-890 (1992).

514 41 Dani, J. A. & Bertrand, D. Nicotinic acetylcholine receptors and nicotinic cholinergic mechanisms  
515 of the central nervous system. *Annu. Rev. Pharmacol. Toxicol.* **47**, 699-729 (2007).

516 42 Kaundal, M., Zameer, S., Najmi, A. K., Parvez, S. & Akhtar, M. Betulinic acid, a natural PDE  
517 inhibitor restores hippocampal cAMP/cGMP and BDNF, improve cerebral blood flow and recover  
518 memory deficits in permanent BCCAO induced vascular dementia in rats. *European journal of*  
519 *pharmacology* **832**, 56-66 (2018).

- 520 43 Pandareesh, M., Anand, T. & Khanum, F. Cognition enhancing and neuromodulatory propensity  
521 of Bacopa monniera extract against scopolamine induced cognitive impairments in rat  
522 hippocampus. *Neurochemical research* **41**, 985-999 (2016).
- 523 44 Blin, J. *et al.* Cholinergic neurotransmission has different effects on cerebral glucose  
524 consumption and blood flow in young normals, aged normals, and Alzheimer's disease patients.  
525 *Neuroimage* **6**, 335-343 (1997).
- 526 45 Scremin, O. U. *et al.* Prolonged effects of cholinesterase inhibition with eptastigmine on the  
527 cerebral blood flow-metabolism ratio of normal rats. *Journal of Cerebral Blood Flow &*  
528 *Metabolism* **13**, 702-711 (1993).
- 529 46 Glikmann-Johnston, Y., Saling, M. M., Reutens, D. C. & Stout, J. C. Hippocampal 5-HT1A receptor  
530 and spatial learning and memory. *Frontiers in pharmacology* **6**, 289 (2015).
- 531 47 Guo, K., Yin, G., Zi, X., Zhu, H. & Pan, Q. Effect of selective serotonin reuptake inhibitors on  
532 expression of 5-HT1AR and neurotransmitters in rats with vascular dementia. *Genet. Mol. Res* **15**  
533 (2016).

534

### 535 **Acknowledgments**

536 The authors are grateful to Monash University Malaysia and Universiti Sains Malaysia for the  
537 experimental laboratory support and facilities. SB was supported by Monash University Malaysia  
538 Merit Scholarship.

### 539 **Authors**

540 Saatheeyavaane Bhuvanendran  
541 Neuropharmacology Research laboratory, Jeffrey Cheah School of Medicine and Health Sciences,  
542 Monash University Malaysia, Bandar Sunway, Selangor, Malaysia  
543 Email: b.bhuvanendranpillai@monash.edu

544

545 Siti Najmi Syuhadaa Bakar  
546 Email: ctnajmi@usm.my  
547 Centre for Drug Research, Universiti Sains Malaysia, Penang, Malaysia.

548

549 Yatinesh Kumari  
550 Neuropharmacology Research laboratory, Jeffrey Cheah School of Medicine and Health Sciences,  
551 Monash University Malaysia, Bandar Sunway, Selangor, Malaysia  
552 Email: yatinesh.kumari@monash.edu

553

554 Iekhsan Othman  
555 Neuropharmacology Research laboratory, Jeffrey Cheah School of Medicine and Health Sciences,  
556 Monash University Malaysia, Bandar Sunway, Selangor, Malaysia  
557 Email: [iekhsan.othman@monash.edu](mailto:iekhsan.othman@monash.edu)

558

559 **\*Correspondence:**

560 Mohd Farooq Shaikh  
561 Neuropharmacology Research laboratory, Jeffrey Cheah School of Medicine and Health Sciences,  
562 Monash University Malaysia, 47500 Bandar Sunway, Selangor, Malaysia  
563 Tel: +603-55144483; Fax: +603-55146323  
564 E-mail: farooq.shaikh@monash.edu

565  
566 Zurina Hassan, PhD  
567 Centre for Drug Research, Universiti Sains Malaysia, 11800 Penang, Malaysia.  
568 Tel: +604-6532726; Fax: +604-6568669  
569 E-mail: zurina\_hassan@usm.my

570

## 571 **Contributions**

572 MFS, SB and ZH were involved in conceptualizing, designing the study, data analysis, and  
573 manuscript writing. SB performed all the experiments and supported by SNSB and ZH. IO and  
574 YK were involved in LC-MS/MS and gene expression study respectively. All authors gave their  
575 final approval for the submission of the manuscript.

## 576 **Competing interest**

577 The authors declare no competing interests

578 **Figure 1.** The effect of post-training administration of embelin on the performance of MWM in  
579 PBOCCA- induced CCH rats. (a) Escape latency in the MWM test of each training day. (b) Mean  
580 time spent on the platform zone in the MWM test. The behavioral analysis for (a) and (b) were  
581 compared to negative control group (P + Veh). Data are expressed as mean  $\pm$  S.E.M. from (n=7)  
582 with \* p < 0.05, \*\* p < 0.01, \*\*\* p < 0.001.

583 **Figure 2.** The effect of embelin on LTP in the CA1 hippocampus. (a) Time course changes in  
584 normalized fEPSP amplitude after TBS (b) The mean fEPSP amplitude during the 2-h interval  
585 after TBS. The analysis for (a) and (b) were compared to negative control group (P + Veh). Data  
586 are expressed as the mean  $\pm$  S.E.M. (n=6) with \* p < 0.05, \*\* p < 0.01, \*\*\* p < 0.001

587 **Figure 3.** The effect of embelin on amyloid-beta (a) APP and Tau (b) Mapt mRNA expression  
588 level in the rat hippocampus using real-time PCR. All changes in the expression levels were  
589 compared to negative control group (P + Veh). Data are expressed as the mean  $\pm$  S.E.M. (n=6)  
590 with \* p < 0.05, \*\* p < 0.01, \*\*\* p < 0.001, \*\*\*\* p < 0.001

591 **Figure 4.** The effect of embelin on synaptic plasticity mRNA expression level (a) BDNF and (b)  
592 CREB1 in the rat hippocampus using real-time PCR. All changes in the expression levels were  
593 compared to negative control group (P + Veh). Data are expressed as the mean  $\pm$  S.E.M. (n=6)  
594 with \* p < 0.05, \*\* p < 0.01, \*\*\* p < 0.001

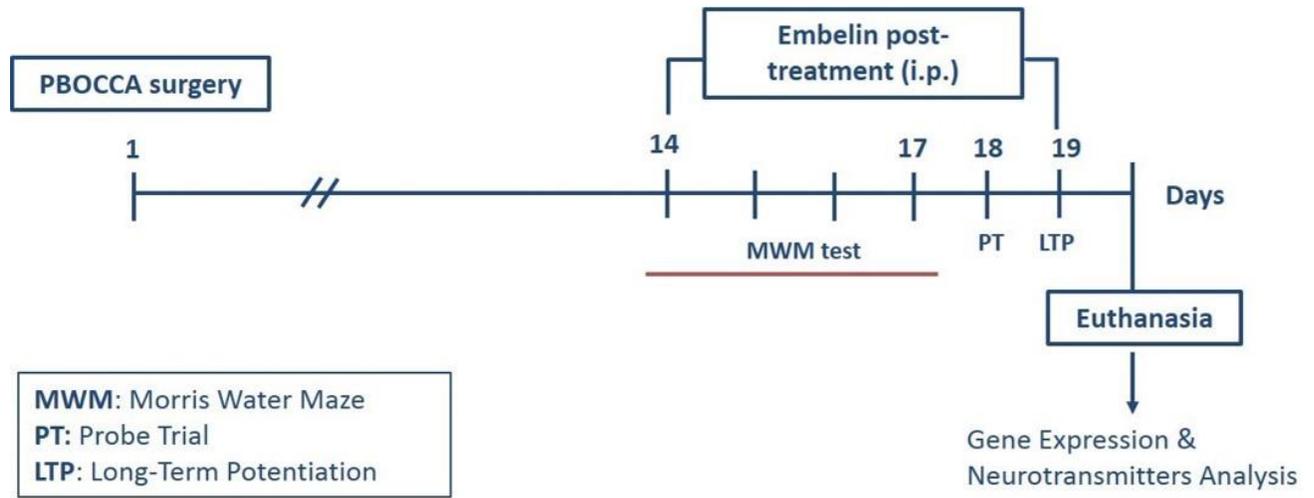
595 **Figure 5.** The effect of embelin on oxidative stress (a) SOD1 and neuroinflammation (b) NF- $\kappa$ B  
596 mRNA expression level in the rat hippocampus using real-time PCR. All changes in the expression  
597 levels were compared to negative control group (P + Veh). Data are expressed as the mean  $\pm$   
598 S.E.M. (n=6) with \* p < 0.05, \*\* p < 0.01, \*\*\* p < 0.001, \*\*\*\* p < 0.001

599 **Figure 6.** The effect of embelin on hippocampus neurotransmitters level of CCH-induced  
600 PBOCCA rats. The neurotransmitters included are (a) Glutamate (b) GABA (c) Acetylcholine (d)  
601 Serotonin. All changes in the expression levels were compared to negative control group (P +  
602 Veh). Data are expressed as the mean  $\pm$  S.E.M. (n=6) with \* p < 0.05, \*\* p < 0.01, \*\*\* p < 0.001,  
603 \*\*\*\* p < 0.001

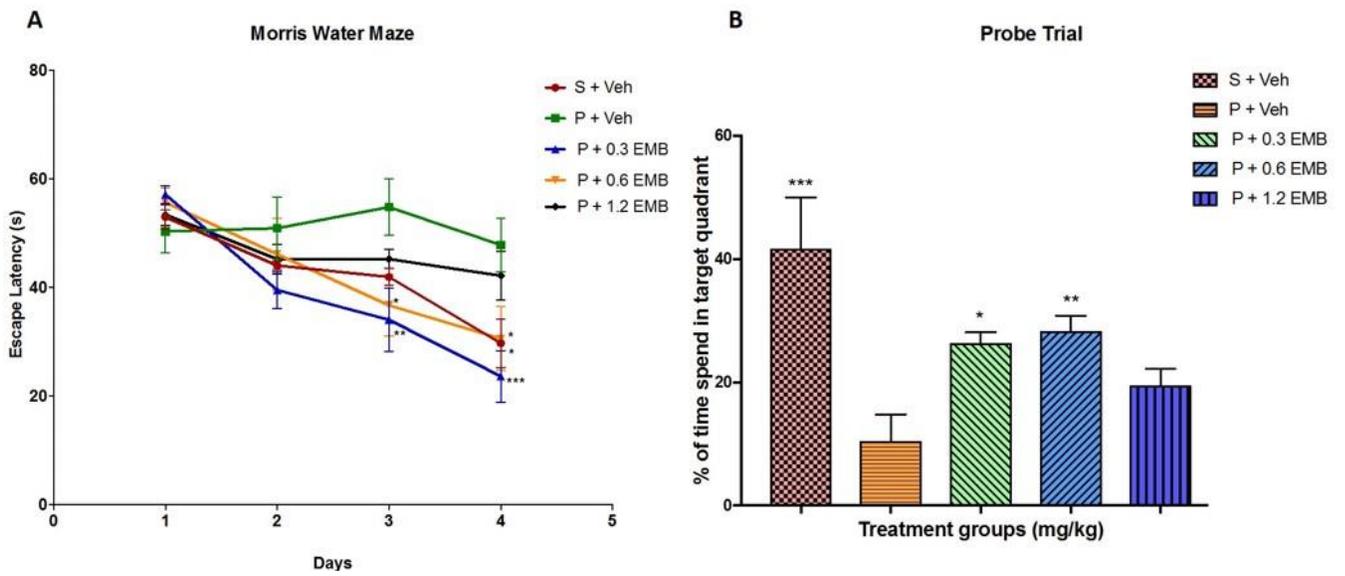
604 **Figure 7.** Mechanism of action of embelin in a Rat Model of Chronic Cerebral Hypoperfusion

605 **Figure 8.** Schematic representation of the experimental procedure and treatment schedule for  
606 CCH-induced PBOCCA rats

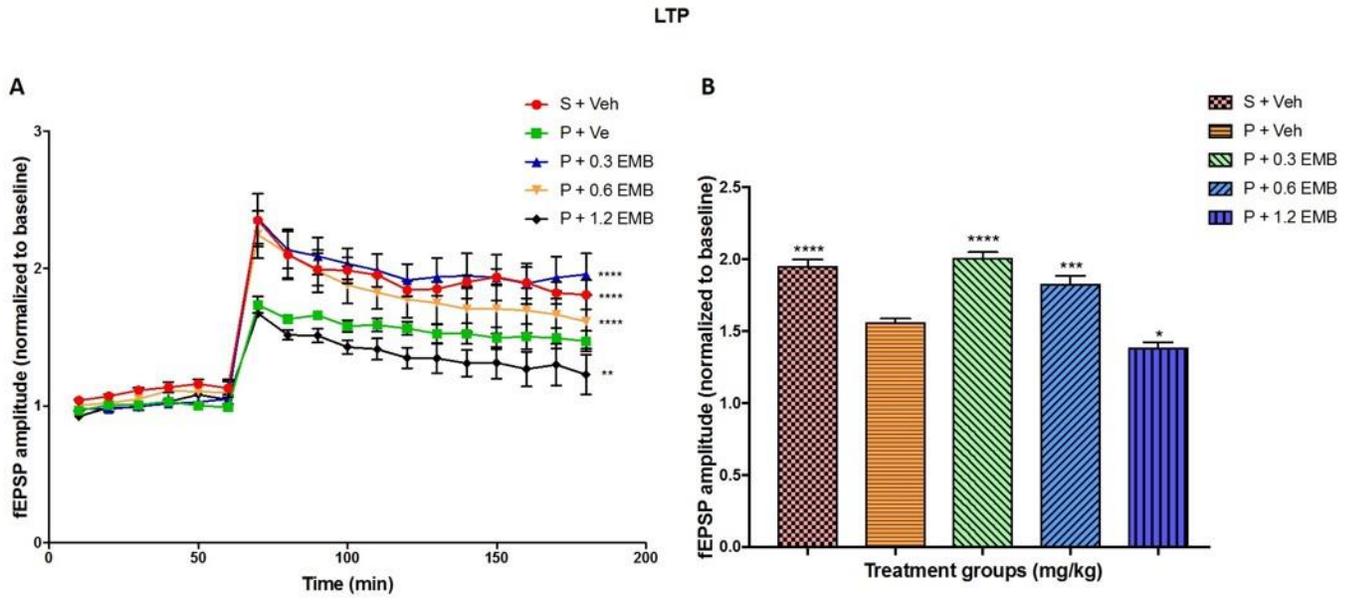
**Figure 1**



**Figure 2**



**Figure 3**



**Figure 4**

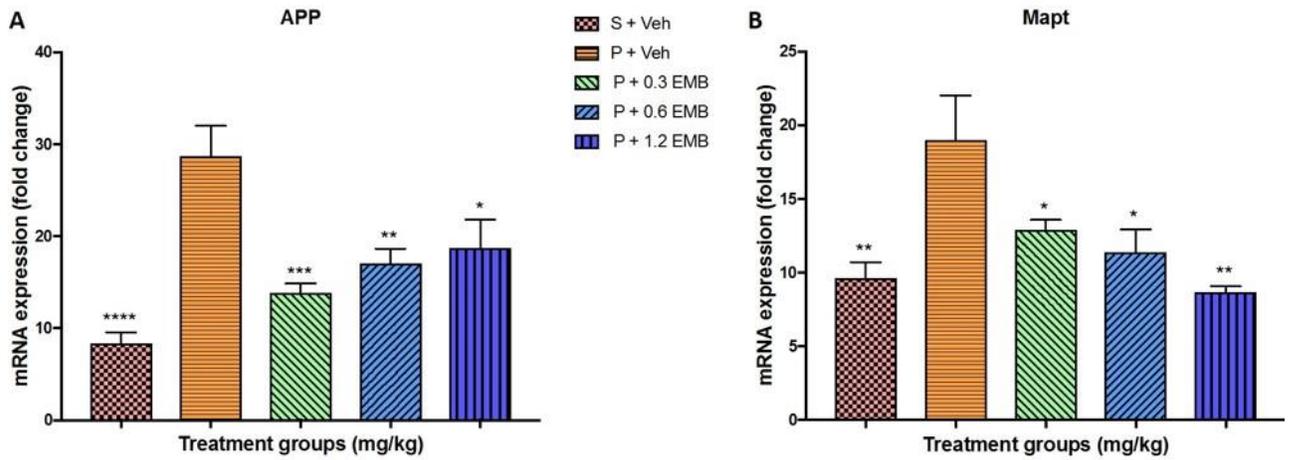


Figure 5

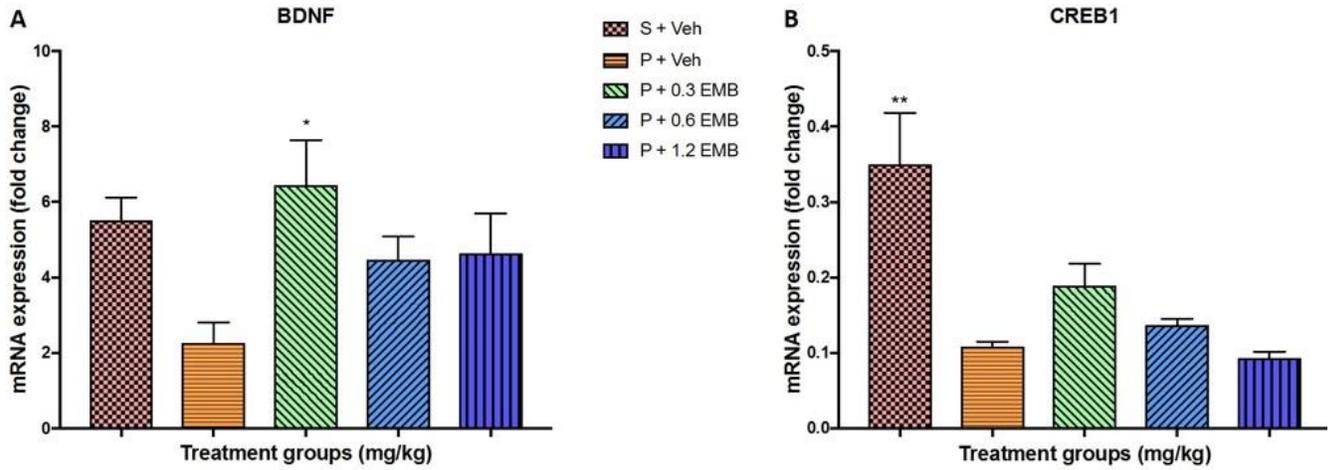


Figure 6

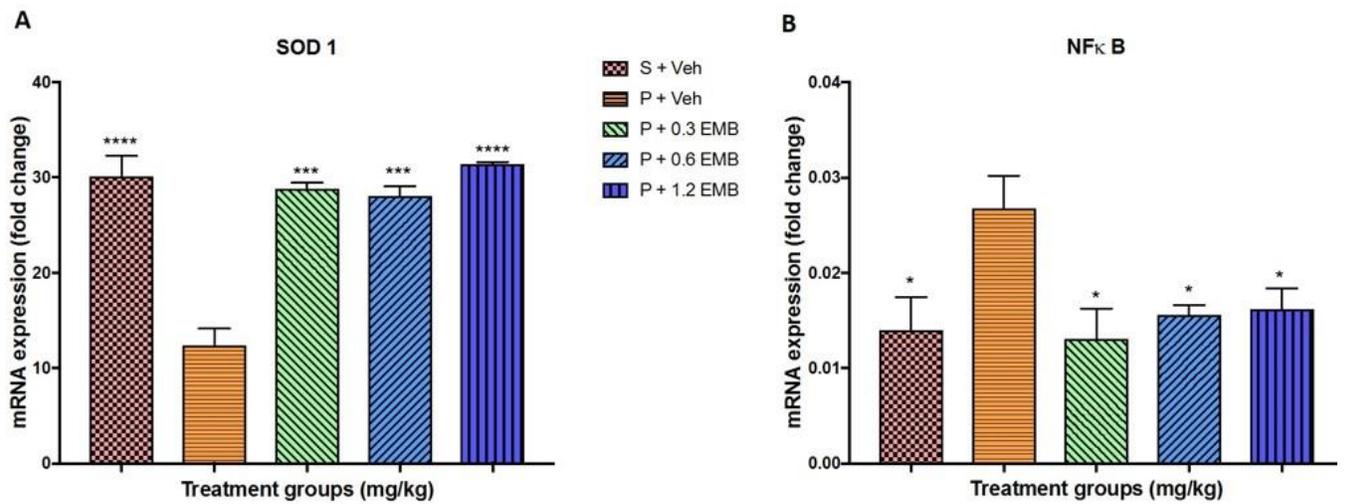


Figure 7

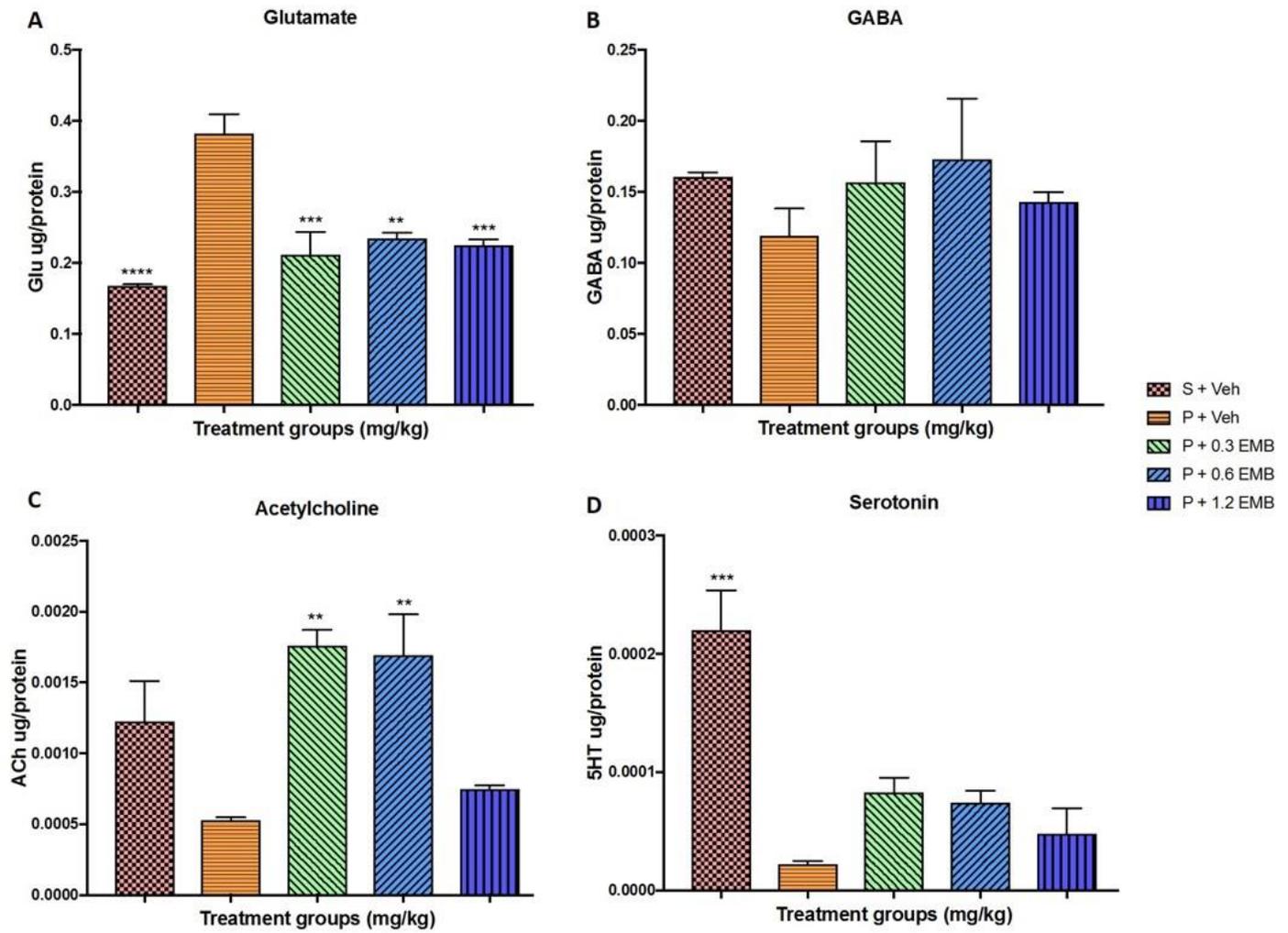
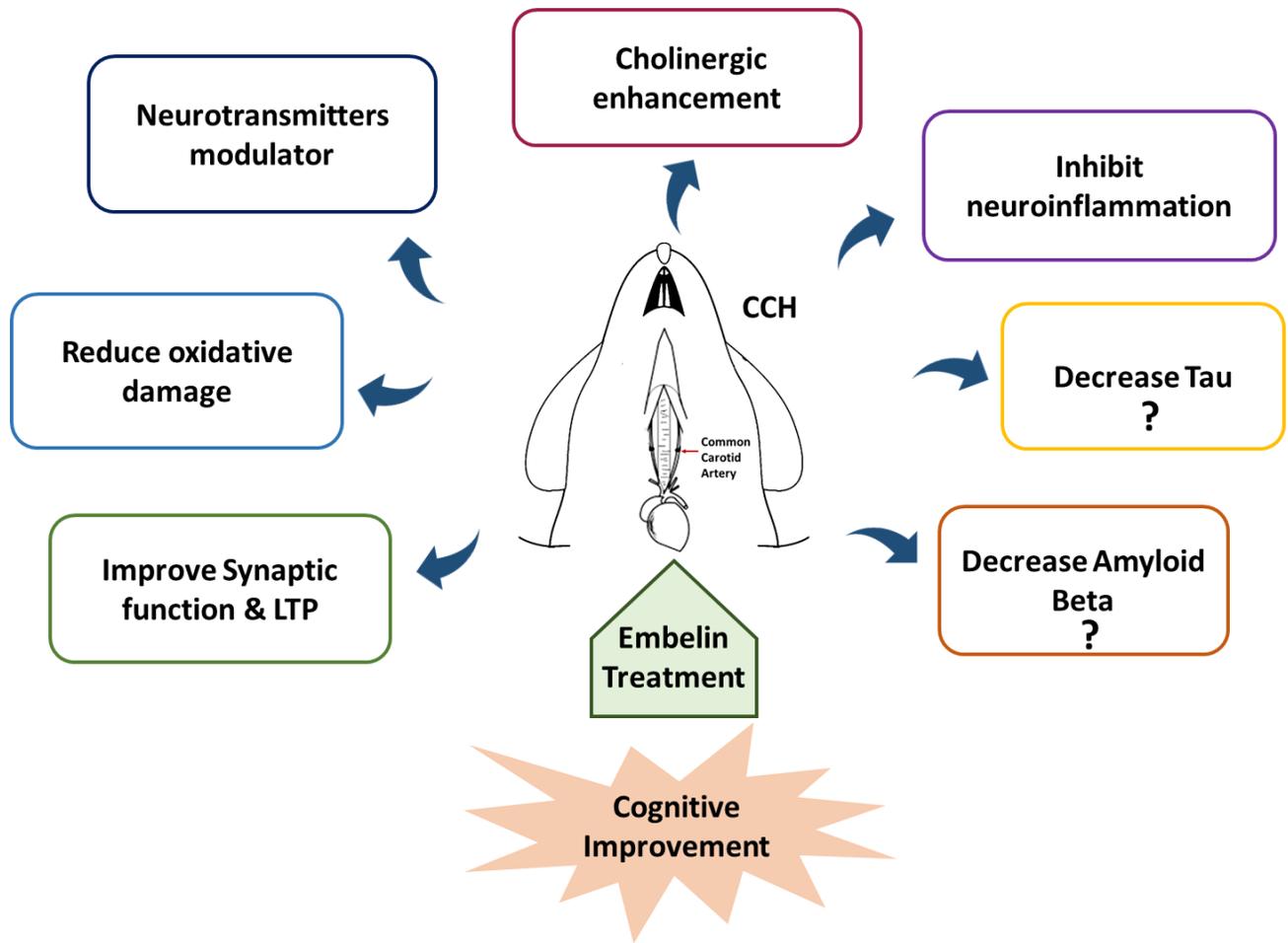


Figure 8



# Chapter 6

## 6.1 Introduction

Streptozotocin (STZ) (2-deoxy-2-(3-methyl-3-nitrosourea)-1-D-glucopyranose) is also known as glucosamine-nitrosourea. This compound is generally used to induce experimental diabetes in animals (70). On the other hand, administration of STZ through intracerebroventricular (icv) or intraperitoneal (ip) routes is able to induce sporadic AD-like condition with brain insulin resistance, accumulation of A $\beta$ , tau, oxidative stress and progressive decline in cognitive function (71). According to Šalković-Petrišić (26), similar pathological features can be observed between sporadic AD in humans and STZ-icv treated rats. Accumulation of laboratory evidences demonstrated that STZ-icv animal models had been extensively used to assess the therapeutic potential of both old and novel compounds and drugs for the development of potential anti-AD drug (72). Herein, the third part of our study, we evaluated the neuroprotective effects of embelin in STZ-induced sporadic AD like condition in an *in-vitro* model using rat primary hippocampal neuronal cultures.

Manuscript Number: EJP-49604

Title: Embelin prevents amyloid beta accumulation via GSK-3 pathway in an  
in vitro model of streptozotocin-induced AD like condition

Article Type: Research Paper

Section/Category: Molecular and cellular pharmacology

Keywords: embelin, streptozotocin, Alzheimer's disease, neuroprotection,  
hippocampal neuronal culture

Corresponding Author: Dr. Mohd Farooq Shaikh,

Corresponding Author's Institution: Neuropharmacology Research  
Laboratory, Jeffrey Cheah School of Medicine and Health Sciences, Monash  
University Malaysia, Jalan Lagoon Selatan, Bandar Sunway 47500, Selangor  
Darul Ehsan, Malaysia.

First Author: Saatheeyavaane Bhuvanendran, BSc (Hons)

Order of Authors: Saatheeyavaane Bhuvanendran, BSc (Hons); Yatinesh  
Kumari, PhD; Iekhsan Othman, PhD; Mohd Farooq Shaikh

Abstract: Previous studies have shown that embelin has beneficial effects in scopolamine-induced amnesia rat model. However, the neuroprotective role of embelin and the mechanism of action in Alzheimer's disease (AD) are remain unclear. Therefore, in this study the neuroprotective effect of embelin against neuronal damage induced by streptozotocin (STZ) in rat hippocampal neuronal culture were evaluated. STZ had been proven to cause imbalance in glycogen synthase kinase 3 (GSK-3) along with amyloidogenesis, oxidative stress and neuroinflammation mimicking the condition of human AD. Present findings demonstrated that embelin (2.5-10 $\mu$ M) has efficiently protected hippocampal neurons against STZ-induced neurotoxicity. An increase in amyloid precursor protein (APP), microtubule-associated protein tau (Mpat), glycogen synthase kinase 3 alpha (GSK-3 $\alpha$ ) and glycogen synthase kinase 3 beta (GSK-3 $\beta$ ) expression levels was observed when STZ (8mM) stimulation was done for 24 hours in the hippocampal neurons. Meanwhile, a huge decrement was showed in APP, Mapt, GSK-3 $\alpha$  and GSK-3 $\beta$  mRNA expression levels suggesting that the pre-treatment with embelin has attenuated STZ-induced insulin signalling (IR) dysfunction. Furthermore, embelin demonstrated protective effect against STZ-induced oxidative stress and neuroinflammation, which was made apparent by the elevated expression of scavenger enzyme (SOD1) and reduction in neuroinflammatory markers (NF- $\kappa$ B) of mRNA levels. Moreover, the histological changes of amyloid beta protein expression induced by STZ were mitigated in a manner comparable to the control group by the pre-treatment of embelin. In conclusion, embelin can improve STZ induced sporadic AD-like condition and can be further developed into drug target to treat and prevent neuronal damages related to AD.

Cover Letter

To,

F. A. Redegeld

The Chief Editor,

European Journal of Pharmacology (Elsevier)

Subject: Submission of a manuscript entitled **“Embelin prevents amyloid beta accumulation via GSK3 pathway in an in-vitro model of streptozotocin-induced AD like condition”**

Dear Editor,

I would like to submit the article entitled above to your esteemed European Journal of Pharmacology (Elsevier). This original research article is regarding an investigation into the anti-AD potential of embelin, a plant derived bezoquinone for Alzheimer’s disease using an *in vitro* model of primary cultured rat hippocampal neurons. This research is important as embelin has been experimentally proven to possess many properties such as being anti-oxidant and anti-inflammatory, but no study so far has determined the mechanism of action of embelin against STZ induced neurotoxicity which mimics pathology of AD. Neither the manuscript nor any parts of its content are currently under consideration or published in another journal.

Kindly consider this article for publication in your esteemed journal.

Thanking you.

Regards,

Dr. Mohd. Farooq Shaikh, PhD

Senior Lecturer,

Jeffrey Cheah School of Medicine and Health Sciences,

Monash University Malaysia,

Petaling Jaya, Selangor 46150, Malaysia.

Tel: +601 42832410

Email: farooq.shaikh@monash.edu

1 **Embelin prevents amyloid beta accumulation via GSK-3 pathway in an *in vitro* model of**  
2 **streptozotocin-induced AD like condition**

3

4 Saatheeyavaane Bhuvanendran<sup>1</sup>, Yatinesh Kumari<sup>1</sup>, Iekhsan Othman<sup>1</sup>, Mohd Farooq Shaikh<sup>1\*</sup>

5 <sup>1</sup>Neuropharmacology Research Laboratory, Jeffrey Cheah School of Medicine and Health  
6 Sciences, Monash University Malaysia, Bandar Sunway 47500, Selangor, Malaysia

7

8

9

10

11

12

13

14 \*Corresponding author:

15 Mohd Farooq Shaikh

16 Neuropharmacology Research Laboratory, Jeffrey Cheah School of Medicine and Health  
17 Sciences, Monash University Malaysia, Bandar Sunway 47500, Selangor, Malaysia

18 Tel: +603-55144483; Fax: +603-55146323

19 E-mail: [farooq.shaikh@monash.edu](mailto:farooq.shaikh@monash.edu)

20

21

22

23

24

25

26 **Abstract**

27 Previous studies have shown that embelin has beneficial effects in scopolamine-induced amnesia  
28 rat model. Therefore, in this study the neuroprotective effect of embelin against neuronal damage  
29 induced by streptozotocin (STZ) in rat hippocampal neuronal culture were evaluated. STZ had  
30 been proven to cause imbalance in glycogen synthase kinase 3 (GSK-3) along with  
31 amyloidogenesis, oxidative stress and neuroinflammation mimicking the condition of human  
32 AD. Present findings demonstrated that embelin (2.5-10 $\mu$ M) has efficiently protected  
33 hippocampal neurons against STZ-induced neurotoxicity. An increase in amyloid precursor  
34 protein (APP), microtubule-associated protein tau (Mpat), glycogen synthase kinase 3 alpha  
35 (GSK-3 $\alpha$ ) and glycogen synthase kinase 3 beta (GSK-3 $\beta$ ) expression levels was observed when  
36 STZ (8mM) stimulation was done for 24 hours in the hippocampal neurons. A huge decrement  
37 was showed in APP, Mapt, GSK-3 $\alpha$  and GSK-3 $\beta$  mRNA expression levels suggesting that the  
38 pre-treatment with embelin has attenuated STZ-induced insulin signalling (IR) dysfunction.  
39 Furthermore, embelin has protective effect against STZ-induced oxidative stress and  
40 neuroinflammation, which was made apparent by the elevated expression of scavenger enzyme  
41 (SOD1) and reduction in neuroinflammatory markers (NF- $\kappa$ B) of mRNA levels. Moreover, the  
42 histological changes of amyloid beta protein expression induced by STZ were mitigated in a  
43 manner comparable to the control group by the pre-treatment of embelin. In conclusion, embelin  
44 can improve STZ induced sporadic AD-like condition and can be further developed into drug  
45 target to treat and prevent neuronal damages related to AD.

46

47 **Keywords: embelin, streptozotocin, Alzheimer's disease, neuroprotection, hippocampal**  
48 **neuronal culture**

49

50

51

52

53

## 54 **1. Introduction**

55 One of the most common brain neurodegenerative disorders is Alzheimer's Disease (AD), which  
56 is irreversible with clinical symptoms of severe cognitive and memory impairment (Bertram and  
57 Tanzi, 2005; Ferri et al., 2005; Qu et al., 2012; Zhao et al., 2016). The late onset sporadic AD  
58 condition can be represented by the range of people affected by this disease, which is ranging  
59 from 65 years old and above (Ghumatkar et al., 2015). On the contrary, very rare cases have  
60 been reported related to early onset familial AD on people age between 30 and 60 years  
61 (Sherrington et al., 1995) caused by missense mutation or inheritance (Šalković-Petrišić, 2008).  
62 Nonetheless, these two AD conditions have been reported to have a relationship with common  
63 pathological hallmarks with extracellular amyloid beta plaques and intracellular neurofibrillary  
64 tau tangles that disrupt synaptic connections, leading to neuronal death (Butterfield et al., 2006).

65 Recently, Plaschke and Kopitz (2015) showed a pathogenetic link between sporadic AD and type  
66 2 diabetes mellitus (T2DM). Both disorders share similar features associated with glucose  
67 metabolism dysfunction and insulin signalling impairment that results in neuronal damage and  
68 cognitive deficits (Candeias et al., 2012). High expression of insulin mRNA in the hippocampus  
69 and rise in insulin receptor (IR) at hippocampal synaptic membrane are functionally associated  
70 with improved learning and memory (Zhao et al., 2004; Zhao et al., 1999). Nonetheless, the  
71 exact mechanisms of insulin influencing learning and memory still remain vague. However,  
72 growing evidence from *in vitro* and *in vivo* studies revealed that impairment in IR signalling  
73 leads to the activation of glycogen synthase kinase 3 (GSK-3) through kinase/phosphatase  
74 imbalance state (Cross et al., 1995; Hooper et al., 2008; Šalković-Petrišić, 2008). Furthermore,  
75 studies have demonstrated that GSK-3 $\alpha$  and GSK-3 $\beta$  are the two key isoforms derived from  
76 GSK-3 regulating the production of amyloid beta (A $\beta$ ) (Hooper et al., 2008; Phiel et al., 2003)  
77 and tau phosphorylation (Ishiguro et al., 1993), respectively.

78 Streptozotocin (STZ) is chemically known as (2-deoxy-2-[3-methyl-3-nitrosourea] 1-D-  
79 glucopyranose), which is also popular as a diabetic inducing agent in animals (Goud et al.,  
80 2015). In AD related studies, STZ administration induced IR signalling impairment along with  
81 neuroinflammation and oxidative stress in *in vivo* and *in vitro* model, which mimic the human  
82 AD pathology (Calvo-Ochoa and Arias, 2015; Rajasekar et al., 2014; Salkovic-Petrisic et al.,  
83 2006).

84 Embelin, an active ingredient isolated from the fruits of *Embelia ribes* Burm has been  
85 traditionally used as brain tonic for the treatment of neurological related disorders (Kundap et al.,  
86 2017). Recently, this compound has been studied for its neuroprotective effects in AD like  
87 conditions in animals (Arora and Deshmukh, 2017; Bhuvanendran et al., 2018). In this study, the  
88 primary culture of hippocampal neurons was utilised as a model to study the neuroprotective  
89 effect of embelin in STZ-induced neurotoxicity. Using this model, it was aimed to develop a  
90 better understanding the factors that can reverse AD through IR signalling pathways mimicking  
91 the sporadic AD pathology by embelin derived benzoquinone.

92

## 93 **2. Materials and Methods**

### 94 **2.1 Primary neuronal culture**

95 Newborn P4 Sprague Dawley (SD) rat pups were acquired from the Animal Facility of Jeffery  
96 Cheah School of Medicine and Health Sciences, Monash University Malaysia. All animal  
97 experimentation was approved and performed in agreement with Monash Animal Research  
98 Platform (MARP) Animal Ethics Committee with reference number MARP/2017/032. Primary  
99 cultures of hippocampal neurons were prepared as described by Kaech and Banker (2006) and  
100 Qu et al. (2012) with a minor modification. After treatment with 5 mL of trypsin for 15 min at 37  
101 °C, the hippocampal neurons were washed in Krebs buffer for 5 min to stop the trypsin reaction.  
102 Then, the hippocampal neurons were suspended in fresh 2.5 mL Krebs buffer with 5 µL DNase  
103 and slowly triturated. Neurons were plated in poly-D-lysine-coated well, followed by 2 hours  
104 incubation at 37 °C in a humidified atmosphere comprising 95% air and 5% CO<sub>2</sub>. Once the  
105 neurons adhered to the coated well, the medium was substituted with serum-free Neurobasal A  
106 added with 2% B27, 1.34mM glutamine, 28mM Glucose and 1.34 mL antibiotic-antimycotic  
107 solution (100X) followed by 8– 10 days incubation period with half of the medium being  
108 changed each 2 days to guarantee neuronal growth (Chen et al., 2008; Chen et al., 2009).

### 109 **2.2 *In vitro* neuroprotection test**

110 To evaluate the protective role of embelin against STZ instigated neurotoxicity on primary  
111 hippocampal neuronal culture, three independent experiments were directed to examine (i) the  
112 effect of various concentrations of embelin on neuronal cultures, (ii) the impact of various

113 concentrations of STZ on neuronal cultures viability and lastly (iii) the defensive role of various  
114 embelin concentrations against the toxicity caused by STZ on the primary hippocampal neuronal  
115 cultures. Poly-D-lysine-coated 96 well flat-bottomed plates were used for seeding the neurons at  
116  $3 \times 10^4$  neurons per well. STZ and embelin were dissolved in DMEM at the chosen concentrations  
117 before adding them to neuronal culture.

### 118 **2.3 Effect of different concentrations of embelin on primary hippocampal neuronal culture**

119 Hippocampal neuronal cultures were treated with embelin at concentrations of 2.5, 5, 10, 20, 40,  
120 80, and 160 $\mu$ M for 24 hours. The untreated neuronal cultures served as control. Each sample  
121 were screened in triplicates and each test was repeated 3 times in order to confirm the accuracy  
122 of the results.

#### 123 **i. Effects of different concentrations of STZ on primary hippocampal neuronal** 124 **culture**

125 Hippocampal neuronal cultures were treated with STZ for 24 hours at varied concentrations of  
126 0.39, 0.78, 1.56, 3.125, 6.25 and 12.5mM. The control neurons did not receive any treatment.  
127 Each sample test was run in triplicate. The half maximal inhibitory concentration ( $IC_{50}$ ) of STZ  
128 was determined using following formula as described by Qu et al. (2012).

$$129 \text{Lg}(IC_{50}) = \text{Lg}(X_m) - \text{Lg}(I) \times [P_s - (3 - P_m - P_n)/4]$$

130 in which,  $X_m$  is the maximal concentration;  $I$  is the dilution factor;  $P_s$  is the sum of inhibition  
131 ratio;  $P_m$  is the maximal inhibition ratio;  $P_n$  is the minimal inhibition ratio and  $Lg$  is the common  
132 logarithm.

#### 133 **ii. Protective effects of different embelin concentrations against STZ induced toxicity**

134 Hippocampal neuronal cultures were pre-incubated with embelin for 2 hours at concentrations of  
135 0.04, 0.08, 0.16, 0.63, 1.25, 2.5, 5, 10 and 20 $\mu$ M before exposure to STZ for 24 hours at the  $IC_{50}$   
136 concentration of 8mM. Untreated neuronal cultures served as control.

### 137 **2.4 MTT Viability Assay**

138 The hippocampal neuronal culture viability was analysed using the standard MTT assay (Chen et  
139 al., 2009). After embelin and STZ treatment, 20  $\mu$ L MTT (5mg/mL) was added to the culture and

140 incubated for 4 hours. Then, 100  $\mu$ L DMSO was replaced with the culture medium and  
141 absorbance was read at 570 nm using a Microplate reader. The data was presented as a percent of  
142 control value. The percentage of hippocampal neuronal culture viability was ascertained using  
143 the formula as follows;

144 Percentage of neurons viability = (absorbance of treated neurons / absorbance of untreated  
145 neurons) x 100

## 146 **2.5 Grouping and Drug Treatment for Gene expression and Immunofluorescence Studies**

147 The hippocampal neurons were cultured on 24 well flat-bottomed plates coated with poly-D-  
148 lysine at  $5 \times 10^5$  neurons per well. The treatments were divided into 5 groups as below with  
149 triplicate well for each group.

- 150 (i) Group 1: Normal Control, Basic Culture Medium only;
- 151 (ii) Group 2: Negative control, Basic Culture Medium + STZ (8mM);
- 152 (iii) Group 3: Embelin (Low dose) 2.5 $\mu$ M + STZ (8mM);
- 153 (iv) Group 4: Embelin (Medium dose) 5 $\mu$ M + STZ (8mM);
- 154 (v) Group 5: Embelin (High dose) 10 $\mu$ M + STZ (8mM)

155 The treatment began once the neurons reached confluence. Further, the hippocampal neuronal  
156 cultures were pre-treated with embelin or basic media for 2 hours and then were induced with  
157 STZ 8mM for 24 hours. At the end of the experiment, the hippocampal neurons were extracted  
158 for gene expression and immunohistochemistry studies.

## 159 **2.6 Total RNA extraction and Real-Time PCR**

160 Total RNA was extracted from hippocampal neurons using Trizol reagent and phenol-  
161 chloroform extraction as described by Bhuvanendran et al. (2018) with a minor adjustments. The  
162 hippocampal neurons were homogenised in 200  $\mu$ L Trizol solution. Then by adding 40  $\mu$ L of  
163 chloroform, the homogenate was centrifuged at 13,500 rpm for 15 min at 4  $^{\circ}$ C. After that, the  
164 supernatant was deliberately removed and then precipitated with same volume of isopropanol in  
165 another Eppendorf tube. This was then pursued by centrifugation at 13,500 rpm for 10 min at 4  
166  $^{\circ}$ C. Then, the supernatant was delicately removed, and the pellet was washed twice with 70%

167 ethanol. Lastly, the pellet was suspended in 20  $\mu$ L of RNase free water. Nanodrop  
168 Spectrophotometer was utilised to obtain the total RNA concentration and purity. Later, 300 ng  
169 of total RNA from each sample was reverse transcribed to complementary DNA (cDNA) by  
170 using QuantiTect® Reverse Transcription Kit as described by the manufacturer's instruction.  
171 Next, the mRNA gene expression for encoding amyloid precursor protein (APP), microtubule-  
172 associated protein tau (Mpat), (GSK3 $\alpha$ ), (GSK3 $\beta$ ), nuclear factor kappa B (NF- $\kappa$ B), superoxide  
173 dismutase 1 (SOD1) and IMPDH2 in the treated neuronal cultures were estimated by using the  
174 below primer sets procured from Qiagen.

175

176 *APP: Rn\_App\_1\_SG QuantiTect Primer Assay (Cat no: QT00177408)*

177 *Mapt: Rn\_Mapt\_1\_SG QuantiTect Primer Assay (Cat no: QT00174797)*

178 *GSK3 $\alpha$ : Rn\_RGD:620351\_1\_SG QuantiTect Primer Assay (Cat no: QT00187453)*

179 *GSK3 $\beta$ : Rn\_Gsk3b\_1\_SG QuantiTect Primer Assay (Cat no: QT00182406)*

180 *Sod1: Rn\_Sod1\_1\_SG QuantiTect Primer Assay (Cat no: QT00174888)*

181 *NF- $\kappa$ B: Rn\_Nkapl\_va.1\_SG QuantiTect Primer Assay (Cat no: QT02476803)*

182 *IMPDH2: Rn\_Impdh2\_1\_SG QuantiTect Primer Assay (Cat no: QT01576036)*

183 The resulting cDNA and the primer sets for gene of interest were subjected to StepOne Real-  
184 Time PCR using QuantiNova™ SYBR® Green PCR kit. The thermal cycling conditions were  
185 set using a similar protocol that was used by Choo et al. (2018). Lastly, the level of expression  
186 for six genes of interest according to fold change was measured by normalising the comparative  
187 threshold (CT) cycle of target gene against reference gene, IMPDH2 utilising the equation:  $2^{-(CT_{IMPDH2} - CT_{target\ gene})}$   
188 (CT IMPDH2 - CT target gene).

## 189 **2.7 Amyloid beta immunofluorescent staining**

190 The hippocampal neurons that underwent treatment were fixed with 4% paraformaldehyde  
191 (PFA) for 1 hour and rinsed in TBS (50 mM Tris, 150 mM NaCl) followed by 30 mins  
192 incubation in 1% BSA (Sigma) to block non-specific binding sites. Following this, incubation  
193 with the anti-beta amyloid primary antibody (1:500; Abcam; ab68896) was carried out at 4 °C

194 and left overnight. After being washed in TBST (50 mM Tris, 150 mM NaCl, 0.05% Tween 20),  
195 the neurons were incubated with secondary goat anti-rabbit conjugated with IgG-H&L Alexa  
196 Fluor<sup>®</sup> 488 (1:2000, Abcam; ab150077) at room temperature for 1 hour followed by 3 times  
197 washing in TBST. The neurons were then counterstained with mounting media ProLong Gold  
198 antifade Reagent with DAPI (Invitrogen). Image from the samples were photographed using  
199 fluorescence microscope (BX41, Olympus) and neurons were quantified using DigiAcquis 2.0  
200 software. Data were expressed as the percentage ratio of amyloid beta positive neurons (green  
201 fluorescence; excitation 495 nm, emission 519 nm) to total neurons (blue fluorescence;  
202 excitation 358 nm, emission 461 nm) (Zhao et al., 2016).

## 203 **2.8 Statistical Analysis**

204 All results were expressed as mean  $\pm$  standard errors of the mean (SEM). One-way analysis of  
205 variance (ANOVA) with Dunnett's post hoc test were used to calculate the significance  
206 difference between treatment groups. Meanwhile, \* $P < 0.05$  was set as threshold of significance.  
207 Analysis were done using GraphPad Prism version 7.02 software (La Jolla, CA, USA).

208

## 209 **3. Results**

### 210 **3.1 Embelin pre-treatment ameliorated STZ-induced neuronal damage in rat primary** 211 **hippocampal neuronal culture**

212 The primary hippocampal neuronal culture was treated with increasing embelin concentrations  
213 ranging from 2.5 to 160 $\mu$ M to examine the neurotoxicity effect of embelin to neuronal culture.  
214 Findings presented that cell viability significantly decreased when primary hippocampal neurons  
215 treated with embelin at concentrations from 20 $\mu$ M and above displayed (Fig. 1A). The IC<sub>50</sub> of  
216 embelin for primary hippocampal neurons was found to be 37.5  $\mu$ M. Thus, the concentration of  
217 embelin from 10 $\mu$ M and below was selected for the next neuroprotection assay. Further, the  
218 primary hippocampal neuronal cultures were treated with increasing concentrations of STZ for  
219 24 hours and assessed by MTT assay (Fig. 1B) to obtain an optimal dose of STZ as an inducer  
220 for neuroprotection assay. The neurotoxicity effect of STZ on the primary hippocampal neurons  
221 was dose dependent and the IC<sub>50</sub> of STZ was found to be 8mM. Therefore, 8mM of STZ  
222 concentration was selected for further neuroprotection assay.

223 In the neuroprotection assay, the primary hippocampal neurons were pre-incubated with embelin  
224 at concentration ranging from 0.04 to 10 $\mu$ M for 2 hours, which was then followed by the  
225 stimulation of STZ at 8mM for 24 hours. For the neuroprotection assay, neurons viability were  
226 seen decreasing to 38.92%  $\pm$  3.06% after exposure to 8mM STZ for 24 hours, while the pre-  
227 treatment with embelin (2.5, 5, and 10 $\mu$ M) for 24 hours significantly enhance neurons viability to  
228 68.74%  $\pm$  2.96% ( $P < 0.0001$ ), 60.09%  $\pm$  2.77% ( $***P < 0.001$ ) and 56.98%  $\pm$  3.76% ( $**P <$   
229 0.01) (Fig. 2), respectively. Therefore, embelin at concentration of 2.5, 5 and 10 $\mu$ M were chosen  
230 for further gene expression and immunohistochemistry studies.

### 231 **3.2 Changes in mRNA levels in the primary rat hippocampal neuronal culture**

232 To observe the impact of STZ-induced neurotoxicity on gene expression, the mRNA expression  
233 level was studied using the real time PCR analysis. The expression levels of APP and Mapt  
234 mRNA were up-regulated by STZ (8mM) exposure in primary rat hippocampal neuronal culture  
235 when contrasted with the control. This up-regulation was reduced by embelin pre-treatment in  
236 contrast with the STZ treated group alone (Fig. 3A and B). However, elevated expression of  
237 Mapt mRNA level was seen in hippocampal neuronal culture with high dose of embelin at  
238 10 $\mu$ M. On the other hand, upregulation of GSK3 $\alpha$  and GSK3 $\beta$  mRNA levels were noted in STZ  
239 treated group compared to that of the control group. Moreover, 2.5 $\mu$ M embelin was seen to  
240 significantly decrease the expression of both mRNA levels. Nevertheless, there was a significant  
241 increment in GSK3 $\beta$  mRNA level by embelin treatment at 10 $\mu$ M (Fig. 4A and B).

242 Besides, STZ was observed to deplete SOD1 antioxidant mRNA in the hippocampal neuronal  
243 culture (Fig 5A). Meanwhile, embelin pre-treatment at 2.5 $\mu$ M and 5 $\mu$ M concentrations  
244 significantly up-regulated the SOD1 expression level with a significant value of  $***P < 0.001$   
245 and  $**P < 0.01$ , respectively. Furthermore, an increment in inflammatory mRNA expression of  
246 NF $\kappa$ B was noted in the hippocampal neuronal culture of STZ treated group. However, pre-  
247 treatment of embelin suppressed the expression level of NF $\kappa$ B mRNA in a dose-dependent  
248 manner ( $*P < 0.05$ ) with 1-fold changes in correlation with STZ treated group (Fig 5B).

### 249 **3.3 Embelin pre-treatment inhibited amyloid beta protein expression level in the primary** 250 **rat hippocampal neuronal culture**

251 To examine whether embelin can inhibit amyloid beta via its neuroprotective effects, the protein  
252 expression of amyloid beta was studied using immunofluorescence staining method. As shown in  
253 Fig. 6A, the green amyloid beta-positive neurons were significantly increased in STZ-treated  
254 neuronal culture, whereas few amyloid beta-positive neurons were identified in the control  
255 group. Besides, the quantity of amyloid beta-positive neurons were seen to be reduced in a dose  
256 dependent manner when treated with embelin. Statistical result on the ratio of amyloid beta  
257 staining to total neuron number as in Fig. 6B suggested that STZ had significantly increase the  
258 expression of amyloid beta (25.63% + 2.73%, \*\*\*\* $P < 0.0001$ ). Contrary, embelin pre-treatment  
259 (2.5, 5 and 10 $\mu$ M) has reversed the A $\beta$ -induced neuronal death (4.94% + 0.58%, \*\*\*\* $P <$   
260 0.0001; 7.64% + 0.60%, \*\*\*\* $P < 0.0001$ ; 11.76% + 1.53%, \*\* $P < 0.01$ ).

261

#### 262 **4. Discussion**

263 An *in vitro* AD model was established by inducing the rat primary hippocampal neuronal  
264 cultures with streptozotocin (STZ). Streptozotocin (STZ) is a glucosamine-nitrosourea  
265 compound utilised as an experimental tool to induce AD-like condition. The key features of this  
266 model mimicking the clinical AD are A $\beta$  fragments, A $\beta$  deposits and total tau protein (Kamat,  
267 2015). The present results demonstrated that embelin possess neuroprotective potential against  
268 STZ induced neurotoxicity in rat primary hippocampal neurons. This study is first of its kind  
269 demonstrating the neuroprotective effect of embelin on STZ induced AD-like condition in rat  
270 primary hippocampal neuronal culture. As of late, it has been demonstrated by Arora and  
271 Deshmukh (2017) that embelin administration had improved intracerebroventricular (icv) STZ-  
272 induced memory deficit in rats. The mechanism of action supporting the potential benefit of  
273 embelin on STZ induced AD-like condition was not studied in great detail. Therefore, this study  
274 was aimed to investigate the anti-AD effect of embelin in rat primary hippocampal neuronal  
275 cultures exposed to STZ. The exposure of cultured neurons to STZ resulted in neuronal death.  
276 The present findings demonstrated that pre-treatment with embelin (2.5-10 $\mu$ M) has significantly  
277 thwarted STZ-induced neurotoxicity and subsequent neuronal death as shown by MTT assay.  
278 These results suggest the potential of embelin as a neuroprotective drug against STZ-induced  
279 neurotoxicity without side effects. However based on our results, we found that embelin does not  
280 fully restore the neuronal viability. This is because more than 50% of neuronal death occurred

281 due to exposure of 8mM STZ during the treatment assay. Thus, pre-treatment with embelin can  
282 only lead to 70% increase in neuronal viability and it was significant when compared to STZ  
283 treated group alone.

284 In icv-STZ treated rats, alterations of brain insulin system lead to the insulin receptor (IR) and  
285 phosphatidylinositol-3 kinase (PI3K) signalling cascade dysfunction that induces insulin-  
286 resistant brain state (Šalković-Petrišić, 2008). This further leads to the activation of glycogen  
287 synthase kinase-3 (GSK-3) in which isoforms alpha and beta subsequently induce A $\beta$   
288 accumulation and tau hyperphosphorylation (Kamat, 2015), which share many features with a  
289 sporadic AD in humans. *In vitro* experiments of this study revealed a significant increase in APP  
290 and GSK-3 $\alpha$  mRNA when neurons isolated from the primary rat hippocampus were exposed to  
291 STZ for 24 hours. This is supported by Phiel et al. (2003) clarifying the close relationship  
292 between the increase in GSK-3 $\alpha$  activity and the processing of APP as well as generation of A $\beta$ .  
293 Furthermore, according to Rajasekar et al. (2016) there was increased in APP expression level in  
294 STZ induced astrocytes for 24 hours. In this study, it was demonstrated that pre-treating the rat  
295 hippocampal neurons with embelin for 2 hours has significantly decreased the APP and GSK-3 $\alpha$   
296 mRNA expression level. However in the case of APP, our results are unusual with embelin  
297 treated groups demonstrated significant decreased below the baseline vehicle control level.  
298 Generally APP play an important role in synaptic maintenance as well as neuronal migration  
299 during early embryogenesis (van der Kant and Goldstein, 2015). Yet in this study, it is still  
300 unclear whether the reduction in APP mRNA level is due to embelin alone or some other factors  
301 that may involve in neuroprotection.

302 On the other hand, STZ treatment significantly increased the GSK-3 $\beta$  mRNA level. As the GSK-  
303 3 $\beta$  regulates the expression level of Mapt in AD condition, it can be said that the Mapt  
304 expression level should mimic that of GSK-3 $\beta$ . In this study, there was an increment of Mapt  
305 mRNA expression similar to GSK-3 $\beta$  (Mendes et al., 2009), but not significant compared to the  
306 control group. The present results showed that only rat hippocampal neurons pre-treated with  
307 embelin 2.5 $\mu$ M concentration had significantly reduced the mRNA expression level of Mapt and  
308 GSK-3 $\beta$ . In the case of higher concentration, the results were unusual with embelin at 10 $\mu$ M  
309 dose having a significant rise in GSK-3 $\beta$  mRNA level and a similar pattern that can be observed  
310 in Mapt mRNA expression even though it was not significant. One plausible reason for this is

311 that once a drug had reached the plateau of the dose-effect curve, there is a very little benefit but  
312 a significantly greater risk for toxicity at higher doses (Lowe and Lertora, 2013). This theory is  
313 supported by the neuroprotection results in this study showing a bell shape dose-response curve  
314 whereby the neuroprotective effect of embelin at 5 and 10 $\mu$ M dose started to reduce in  
315 comparison to 2.5 $\mu$ M dose (Fig. 3).

316 The present study investigated whether the STZ-induced AD-like pathology is associated with  
317 alteration in the level of SOD1 in primary rat hippocampal neuronal culture. It was found that the  
318 induction of STZ decreased the expression of SOD1 mRNA expression neuronal culture.  
319 Besides, Qu et al. (2012) stated that STZ can cause oxidative stress in the brain and cognitive  
320 disabilities in rats. In this study, embelin with 2.5 and 5 $\mu$ M dose significantly attenuated STZ-  
321 induced SOD1 downregulation in neuronal culture. According to literature, embelin possesses  
322 potent antioxidative activities (Mahendran et al., 2011a; Poojari, 2014). Hence embelin might  
323 exert its neuroprotective effects through antioxidant activity with upregulation of SOD1 mRNA  
324 expression.

325 In the case of NF $\kappa$ B, it was discovered that there was an increment in NF $\kappa$ B expression due to  
326 STZ-induced neurotoxicity, which is predictable with the outcomes found in literature (Choi et  
327 al., 2014; Rajasekar et al., 2016) with somewhat higher expression level than that of the baseline  
328 vehicle control. Pre-treatment of embelin has significantly ameliorated the NF $\kappa$ B mRNA  
329 expression caused by STZ in a dose-dependent manner. Besides, previous literature yielded the  
330 anti-inflammatory abilities of embelin (Mahendran et al., 2011a; Mahendran et al., 2011b;  
331 Poojari, 2014) whereby the bright orange hydroxybenzoquinone embelin-rich from *Embelia*  
332 *ribes* had been utilised traditionally to treat inflammation. However, all embelin pre-treatments  
333 had a slightly lower expression level of NF $\kappa$ B mRNA when compared to baseline vehicle  
334 control. This suggests that embelin may at least partially exert its anti-AD effect by acting as  
335 anti-inflammatory agent as NF $\kappa$ B involved in activating systemic inflammation.

336 The protective effect of embelin was further verified through immunofluorescence staining  
337 technique in which the effect of embelin on amyloid beta expression was explored. Laboratory  
338 evidences have revealed that accumulation of amyloid beta found in STZ-treated rats (El  
339 Halawany et al., 2017; Wang et al., 2017). The present study showed that pre-treatment of  
340 embelin significantly diminished the amyloid beta expression level induced by STZ treatment in

341 a dose dependent manner similar to APP mRNA level. Thus, this result further supported the  
342 gene expression study as APP gene generates amyloid beta through enzyme secretases known as  
343 alpha, beta and gamma (Zhang et al., 2011). It can be concluded that this study is the first to  
344 report the protective effects of embelin against amyloid beta induced by STZ treatment on  
345 primary rat hippocampal neuronal culture. Figure 7 summarize the potential mechanism of action  
346 of embelin in STZ-induced neurotoxicity in primary rat hippocampal neuronal culture.

347 In conclusion, embelin showed a promising anti-AD like effect in *in vitro* model. Embelin exerts  
348 its neuroprotective effect through GSK-3 pathway, reducing oxidative stress and  
349 neuroinflammation and preventing amyloidogenesis in STZ-induced neurotoxicity in rat  
350 hippocampal neuronal culture.

351

### 352 **Acknowledgements**

353 The authors are grateful to Monash University Malaysia for the experimental laboratory supports  
354 and facilities. SB was supported by Monash University Malaysia Merit Scholarship.

### 355 **Author contributions**

356 MS and SB were involved in designing, conceptualising of the research project and data analysis.  
357 SB performed all the experiments and manuscript writing in its entirety. YS helped in data  
358 analysis and figures in the manuscript. IO gave critical feedback for this study. All authors gave  
359 their final approval for the submission of this manuscript.

### 360 **References**

- 361 Arora, R., Deshmukh, R., 2017. Embelin Attenuates Intracerebroventricular Streptozotocin-Induced  
362 Behavioral, Biochemical, and Neurochemical Abnormalities in Rats. *Molecular neurobiology* 54, 6670-  
363 6680.
- 364 Bertram, L., Tanzi, R.E., 2005. The genetic epidemiology of neurodegenerative disease. *The Journal of*  
365 *clinical investigation* 115, 1449-1457.
- 366 Bhuvanendran, S., Kumari, Y., Othman, I., Shaikh, M.F., 2018. Amelioration of cognitive deficit by  
367 embelin in a scopolamine-induced Alzheimer's disease-like condition in a rat model. *Frontiers in*  
368 *pharmacology* 9.
- 369 Butterfield, D.A., Perluigi, M., Sultana, R., 2006. Oxidative stress in Alzheimer's disease brain: new  
370 insights from redox proteomics. *European journal of pharmacology* 545, 39-50.

371 Calvo-Ochoa, E., Arias, C., 2015. Cellular and metabolic alterations in the hippocampus caused by insulin  
372 signalling dysfunction and its association with cognitive impairment during aging and Alzheimer's  
373 disease: studies in animal models. *Diabetes/metabolism research and reviews* 31, 1-13.

374 Candeias, E., Duarte, A.I., Carvalho, C., Correia, S.C., Cardoso, S., Santos, R.X., Plácido, A.I., Perry, G.,  
375 Moreira, P.I., 2012. The impairment of insulin signaling in Alzheimer's disease. *IUBMB life* 64, 951-957.

376 Chen, X., Liu, J., Gu, X., Ding, F., 2008. Salidroside attenuates glutamate-induced apoptotic cell death in  
377 primary cultured hippocampal neurons of rats. *Brain research* 1238, 189-198.

378 Chen, X., Zhang, Q., Cheng, Q., Ding, F., 2009. Protective effect of salidroside against H<sub>2</sub>O<sub>2</sub>-induced cell  
379 apoptosis in primary culture of rat hippocampal neurons. *Molecular and cellular biochemistry* 332, 85-  
380 93.

381 Choi, D.H., Kwon, I.S., Koo, J.H., Jang, Y.C., Kang, E.B., Byun, J.E., Um, H.S., Park, H.S., Yeom, D.C., Cho,  
382 I.H., 2014. The effect of treadmill exercise on inflammatory responses in rat model of streptozotocin-  
383 induced experimental dementia of Alzheimer's type. *Journal of exercise nutrition & biochemistry* 18,  
384 225.

385 Choo, B.K.M., Kundap, U.P., Kumari, Y., Hue, S.-M., Othman, I., Shaikh, M.F., 2018. Orthosiphon  
386 stamineus Leaf Extract Affects TNF- $\alpha$  and Seizures in a Zebrafish Model. *Frontiers in pharmacology* 9,  
387 139.

388 Cross, D.A., Alessi, D.R., Cohen, P., Andjelkovich, M., Hemmings, B.A., 1995. Inhibition of glycogen  
389 synthase kinase-3 by insulin mediated by protein kinase B. *Nature* 378, 785.

390 El Halawany, A.M., Sayed, N.S.E., Abdallah, H.M., El Dine, R.S., 2017. Protective effects of gingerol on  
391 streptozotocin-induced sporadic Alzheimer's disease: emphasis on inhibition of  $\beta$ -amyloid, COX-2, alpha-  
392 , beta-secretases and A $\beta$ 1a. *Scientific Reports* 7, 2902.

393 Ferri, C.P., Prince, M., Brayne, C., Brodaty, H., Fratiglioni, L., Ganguli, M., Hall, K., Hasegawa, K., Hendrie,  
394 H., Huang, Y., 2005. Global prevalence of dementia: a Delphi consensus study. *The lancet* 366, 2112-  
395 2117.

396 Ghumatkar, P.J., Patil, S.P., Jain, P.D., Tambe, R.M., Sathaye, S., 2015. Nootropic, neuroprotective and  
397 neurotrophic effects of phloretin in scopolamine induced amnesia in mice. *Pharmacology Biochemistry*  
398 *and Behavior* 135, 182-191.

399 Goud, B.J., Dwarakanath, V., Chikka, B., 2015. Streptozotocin-a diabetogenic agent in animal models.  
400 *International Journal of Pharmacy & Pharmaceutical Research* 3, 253-269.

401 Hooper, C., Killick, R., Lovestone, S., 2008. The GSK3 hypothesis of Alzheimer's disease. *Journal of*  
402 *neurochemistry* 104, 1433-1439.

403 Ishiguro, K., Shiratsuchi, A., Sato, S., Omori, A., Arioka, M., Kobayashi, S., Uchida, T., Imahori, K., 1993.  
404 Glycogen synthase kinase 3 $\beta$  is identical to tau protein kinase I generating several epitopes of paired  
405 helical filaments. *FEBS letters* 325, 167-172.

406 Kaeck, S., Banker, G., 2006. Culturing hippocampal neurons. *Nature protocols* 1, 2406.

407 Kamat, P.K., 2015. Streptozotocin induced Alzheimer's disease like changes and the underlying neural  
408 degeneration and regeneration mechanism. *Neural regeneration research* 10, 1050.

409 Kundap, U.P., Bhuvanendran, S., Kumari, Y., Othman, I., Shaikh, M., 2017. Plant derived  
410 phytochemical, embelin in CNS disorders: a systematic review. *Frontiers in pharmacology* 8, 76.

411 Lowe, E.S., Lertora, J.J., 2013. Dose–Effect and Concentration–Effect Analysis, *Principles of Clinical*  
412 *Pharmacology* (Third Edition). Elsevier, pp. 343-356.

413 Mahendran, S., Badami, S., Ravi, S., Thippeswamy, B., Veerapur, V., 2011a. Antioxidant, analgesic and  
414 anti-inflammatory properties of new ninhydrin adduct of embelin. *Pharmaceutical Chemistry Journal* 45,  
415 547-551.

416 Mahendran, S., Badami, S., Ravi, S., Thippeswamy, B.S., Veerapur, V.P., 2011b. Synthesis and evaluation  
417 of analgesic and anti-inflammatory activities of most active free radical scavenging derivatives of  
418 embelin—a structure–activity relationship. *Chemical and Pharmaceutical Bulletin* 59, 913-919.

419 Mendes, C.T., Mury, F.B., de Sá Moreira, E., Alberto, F.L., Forlenza, O.V., Dias-Neto, E., Gattaz, W.F.,  
420 2009. Lithium reduces Gsk3b mRNA levels: implications for Alzheimer disease. *European archives of*  
421 *psychiatry and clinical neuroscience* 259, 16-22.

422 Phiel, C.J., Wilson, C.A., Lee, V.M.-Y., Klein, P.S., 2003. GSK-3 $\alpha$  regulates production of Alzheimer's  
423 disease amyloid- $\beta$  peptides. *Nature* 423, 435.

424 Plaschke, K., Kopitz, J., 2015. In vitro streptozotocin model for modeling Alzheimer-like changes: effect  
425 on amyloid precursor protein secretases and glycogen synthase kinase-3. *Journal of Neural Transmission*  
426 122, 551-557.

427 Poojari, R., 2014. Embelin—a drug of antiquity: shifting the paradigm towards modern medicine. *Expert*  
428 *opinion on investigational drugs* 23, 427-444.

429 Qu, Z.-q., Zhou, Y., Zeng, Y.-s., Lin, Y.-k., Li, Y., Zhong, Z.-q., Chan, W.Y., 2012. Protective effects of a  
430 *Rhodiola crenulata* extract and salidroside on hippocampal neurogenesis against streptozotocin-induced  
431 neural injury in the rat. *PLoS one* 7, e29641.

432 Rajasekar, N., Dwivedi, S., Nath, C., Hanif, K., Shukla, R., 2014. Protection of streptozotocin induced  
433 insulin receptor dysfunction, neuroinflammation and amyloidogenesis in astrocytes by insulin.  
434 *Neuropharmacology* 86, 337-352.

435 Rajasekar, N., Nath, C., Hanif, K., Shukla, R., 2016. Inhibitory effect of memantine on streptozotocin-  
436 induced insulin receptor dysfunction, neuroinflammation, amyloidogenesis, and neurotrophic factor  
437 decline in astrocytes. *Molecular neurobiology* 53, 6730-6744.

438 Šalković-Petrišić, M., 2008. Amyloid cascade hypothesis: is it true for sporadic Alzheimer's disease.  
439 *Periodicum biologorum* 110, 17-25.

440 Salkovic-Petrisic, M., Tribl, F., Schmidt, M., Hoyer, S., Riederer, P., 2006. Alzheimer-like changes in  
441 protein kinase B and glycogen synthase kinase-3 in rat frontal cortex and hippocampus after damage to  
442 the insulin signalling pathway. *Journal of neurochemistry* 96, 1005-1015.

443 Sherrington, R., Rogaeve, E., Liang, Y.a., Rogaeve, E., Levesque, G., Ikeda, M., Chi, H., Lin, C., Li, G.,  
444 Holman, K., 1995. Cloning of a gene bearing missense mutations in early-onset familial Alzheimer's  
445 disease. *Nature* 375, 754.

446 van der Kant, R., Goldstein, L.S., 2015. Cellular functions of the amyloid precursor protein from  
447 development to dementia. *Developmental cell* 32, 502-515.

448 Wang, L., Liu, W., Fan, Y., Liu, T., Yu, C., 2017. Effect of rosiglitazone on amyloid precursor protein  
449 processing and A $\beta$  clearance in streptozotocin-induced rat model of Alzheimer's disease. *Iranian journal*  
450 *of basic medical sciences* 20, 474.

451 Zhang, Y.-w., Thompson, R., Zhang, H., Xu, H., 2011. APP processing in Alzheimer's disease. *Molecular*  
452 *brain* 4, 3.

453 Zhao, C., Lv, C., Li, H., Du, S., Liu, X., Li, Z., Xin, W., Zhang, W., 2016. Geniposide protects primary cortical  
454 neurons against oligomeric A $\beta$ 1-42-induced neurotoxicity through a mitochondrial pathway. *PLoS one*  
455 11, e0152551.

456 Zhao, W.-Q., Chen, H., Quon, M.J., Alkon, D.L., 2004. Insulin and the insulin receptor in experimental  
457 models of learning and memory. *European journal of pharmacology* 490, 71-81.

458 Zhao, W., Chen, H., Xu, H., Moore, E., Meiri, N., Quon, M.J., Alkon, D.L., 1999. Brain insulin receptors and  
459 spatial memory correlated changes in gene expression, tyrosine phosphorylation, and signaling  
460 molecules in the hippocampus of water maze trained rats. *Journal of Biological Chemistry* 274, 34893-  
461 34902.

462

463

464 **Legends**

465 **Fig 1.** Viability of primary rat hippocampal neurons at various doses of embelin and STZ  
466 assessed by MTT assay. (A) Neuronal cultures were incubated with various concentrations of  
467 embelin for 24 hours. Embelin at a concentration up to 10 $\mu$ M for 24 hours shows no toxic effect  
468 on primary rat hippocampal neurons. (B) Neuronal cultures were incubated with various  
469 concentrations of STZ for 24 hours. STZ shows a dose-dependent cytotoxic effect on primary rat  
470 hippocampal neurons. Data are expressed as Mean  $\pm$  SEM of three independent experiment ( $n =$   
471 3)

472 **Fig 2.** Neuroprotection effect of embelin on STZ-induced neurotoxicity in primary rat  
473 hippocampal neuronal culture. Hippocampal neuronal cultures were pre-incubated with various  
474 concentrations of embelin for 2 hours before stimulation of 8mM STZ. embelin at 2.5, 5 and  
475 10 $\mu$ M exhibits significant protection against STZ-induced neurotoxicity. Data are expressed as  
476 Mean  $\pm$  SEM of three independent experiment ( $n = 3$ ) and statistical analysis by one-way  
477 ANOVA followed by Dunnett test \* $P < 0.05$ , \*\* $P < 0.01$ , and \*\*\* $P < 0.001$ .

478 **Fig 3.** The effect of embelin treatment on STZ-induced neurotoxicity in primary rat hippocampal  
479 neuronal culture. A) APP and B) Mapt mRNA expression by real time-PCR. The expressions  
480 were normalized with IMPDH2. Data are expressed as Mean  $\pm$  SEM of three independent  
481 experiment ( $n = 3$ ) and statistical analysis by one-way ANOVA followed by Dunnett test \* $P <$   
482 0.05, \*\* $P < 0.01$ , and \*\*\* $P < 0.001$ .

483 **Fig 4.** The effect of embelin treatment on STZ-induced neurotoxicity in primary rat hippocampal  
484 neuronal culture. A) GSK-3 $\alpha$  and B) GSK-3 $\beta$  mRNA expression by real time-PCR. The  
485 expressions were normalized with IMPDH2. Data are expressed as Mean  $\pm$  SEM of three  
486 independent experiment ( $n = 3$ ) and statistical analysis by one-way ANOVA followed by  
487 Dunnett test \* $P < 0.05$ , \*\* $P < 0.01$ , and \*\*\* $P < 0.001$ .

488 **Fig 5.** The effect of embelin treatment on STZ-induced neurotoxicity in primary rat hippocampal  
489 neuronal culture. A) SOD1 and B) NF $\kappa$ B mRNA expression by real time-PCR. The expressions  
490 were normalized with IMPDH2. Data are expressed as Mean  $\pm$  SEM of three independent  
491 experiment ( $n = 3$ ) and statistical analysis by one-way ANOVA followed by Dunnett test \* $P <$   
492 0.05, \*\* $P < 0.01$ , and \*\*\* $P < 0.001$ .

493 **Fig 6.** The effect of embelin on STZ-induced amyloidogenesis. A) Photomicrographs of  
494 hippocampal neuronal culture treatment for 24 hr was (A1-3) Control (B1-3) STZ 8mM (C1-3)  
495 EMB 2.5 $\mu$ M + STZ 8mM (D1-3) EMB 5 $\mu$ M + STZ 8mM (E1-3) EMB 10 $\mu$ M + STZ 8mM  
496 Amyloid beta-positive neurons were stained in green while cells' nuclei were stained in blue.  
497 Representative photomicrographs were taken at magnifications of 40X B) Quantitative analysis  
498 for the ratio of Amyloid-beta neurons to total neuron number. Data are expressed as Mean  $\pm$   
499 SEM of three independent experiment ( $n = 3$ ) and statistical analysis by one-way ANOVA  
500 followed by Dunnett test \* $P < 0.05$ , \*\* $P < 0.01$ , and \*\*\* $P < 0.001$ .

501 **Fig 7.** Schematic diagram represents the mechanism of action of embelin in STZ-induced AD-  
502 like condition. The figure depicts STZ induced GSK-3 activation of both alpha and beta isoforms  
503 which further lead to amyloid pathology and tau pathology that mimics sporadic Alzheimer's  
504 disease. In addition, STZ increased NF $\kappa$ B mRNA and reduced SOD1 mRNA expression levels  
505 which contributed to neuroinflammation and oxidative stress respectively. Embelin treatment act  
506 as neuroprotectant agent by deactivating GSK-3 that cause reduction in APP and Mapt mRNA  
507 expression levels. Besides, embelin act as antioxidant by increasing SOD1 levels and in the case  
508 of inflammatory pathway, embelin reduces NF $\kappa$ B mRNA. With these effects, embelin treatment  
509 inhibit neurodegeneration and maintain healthy hippocampal neurons in the brain

Figure 1

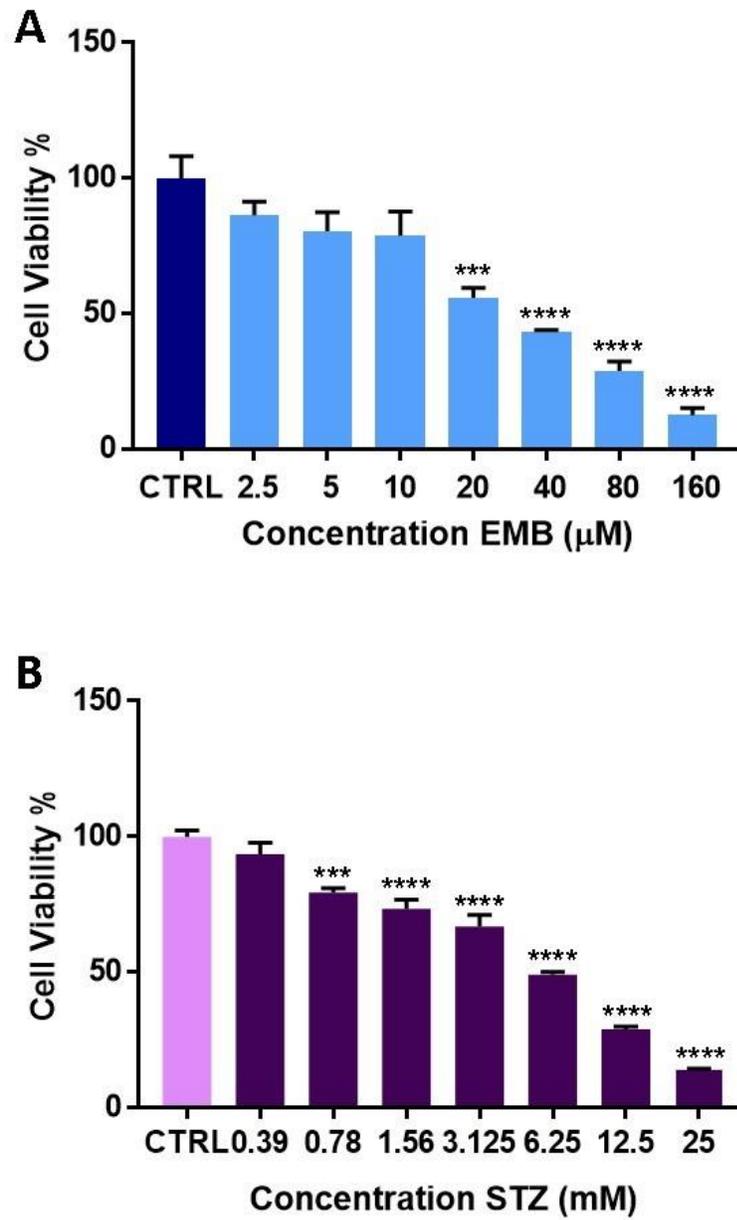


Figure 2

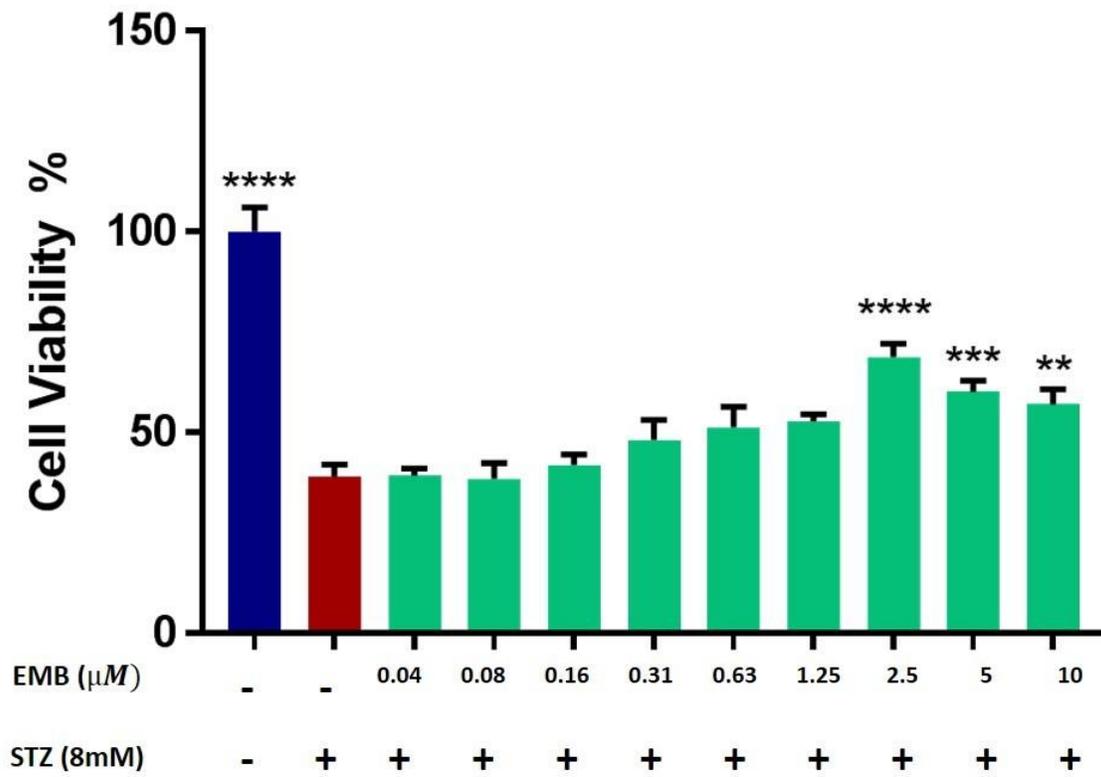


Figure 3

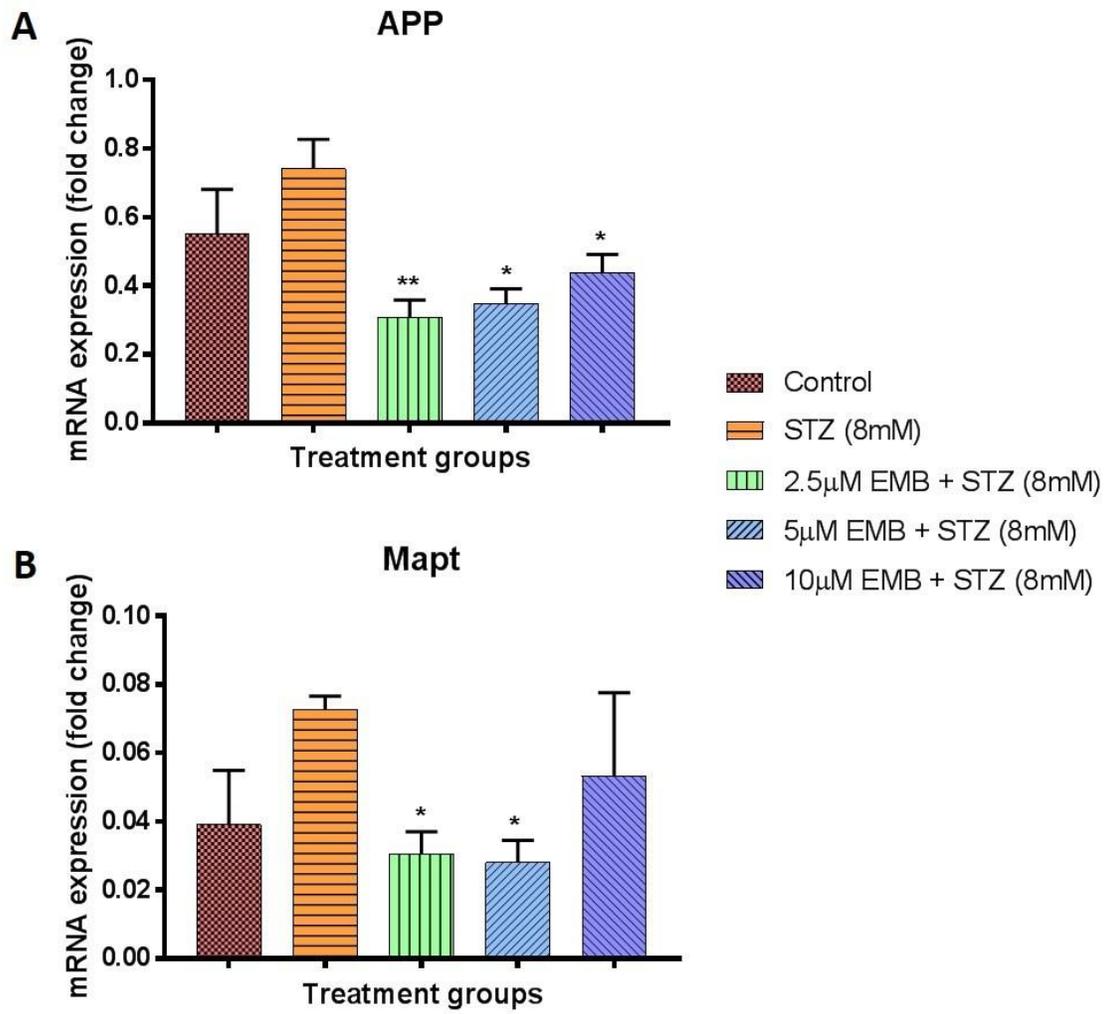


Figure 4

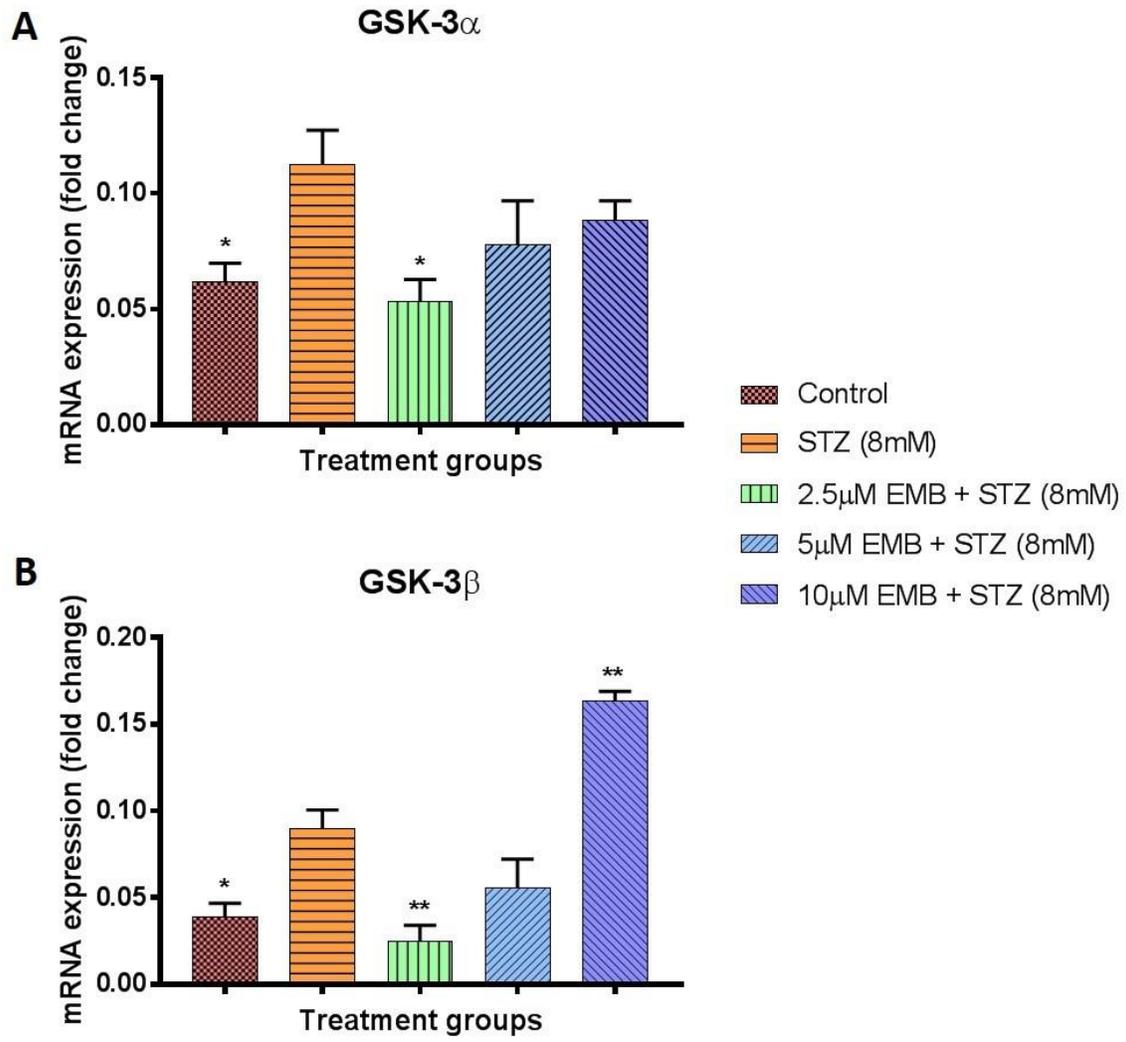


Figure 5

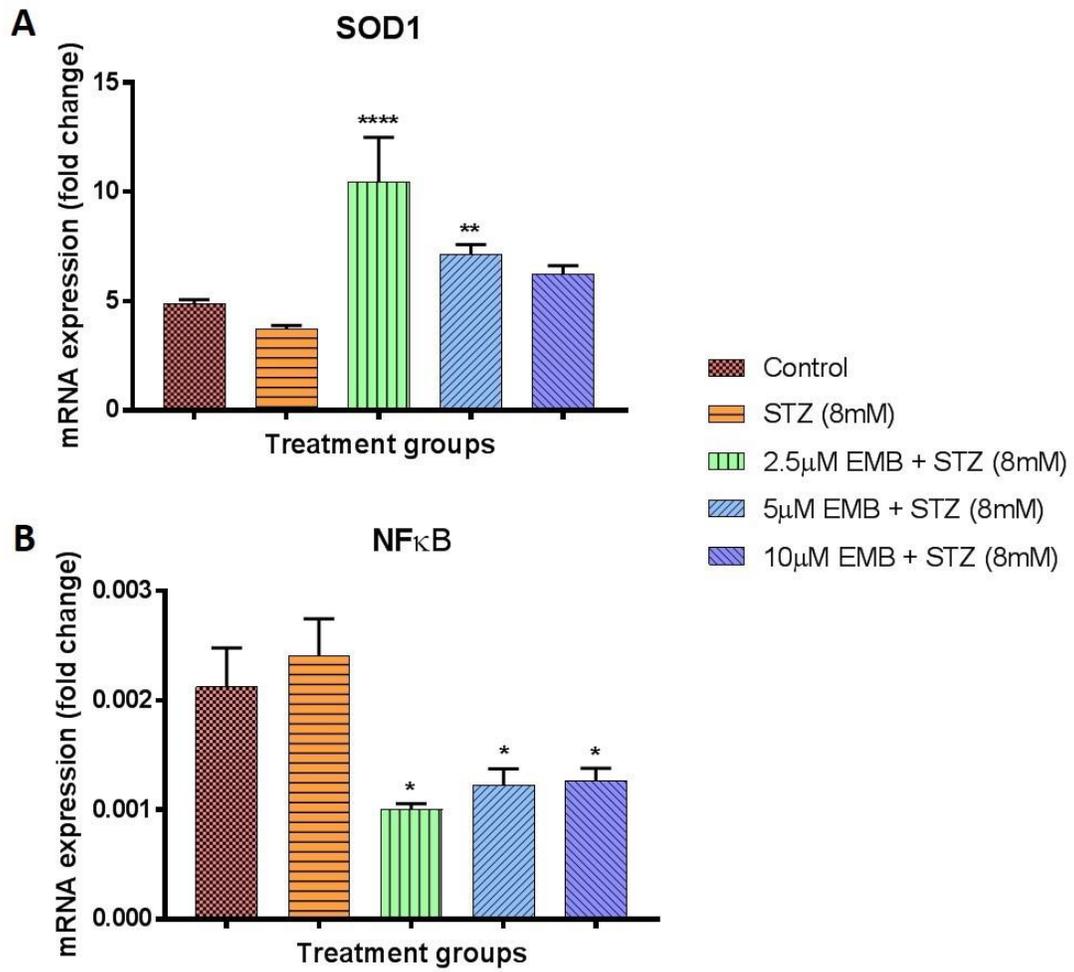


Figure 6

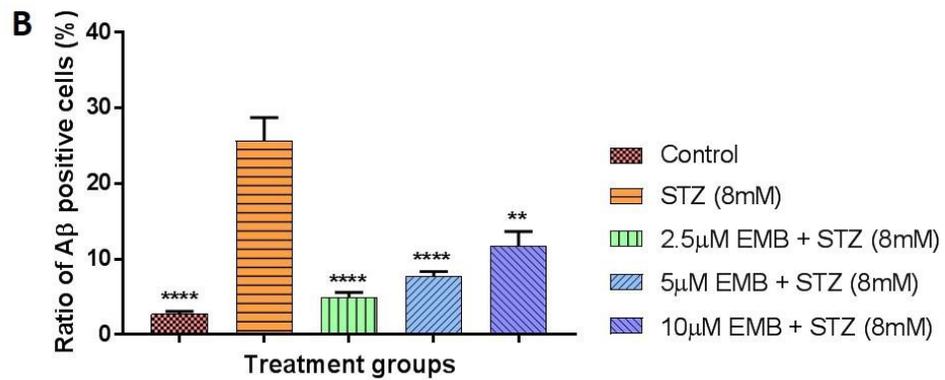
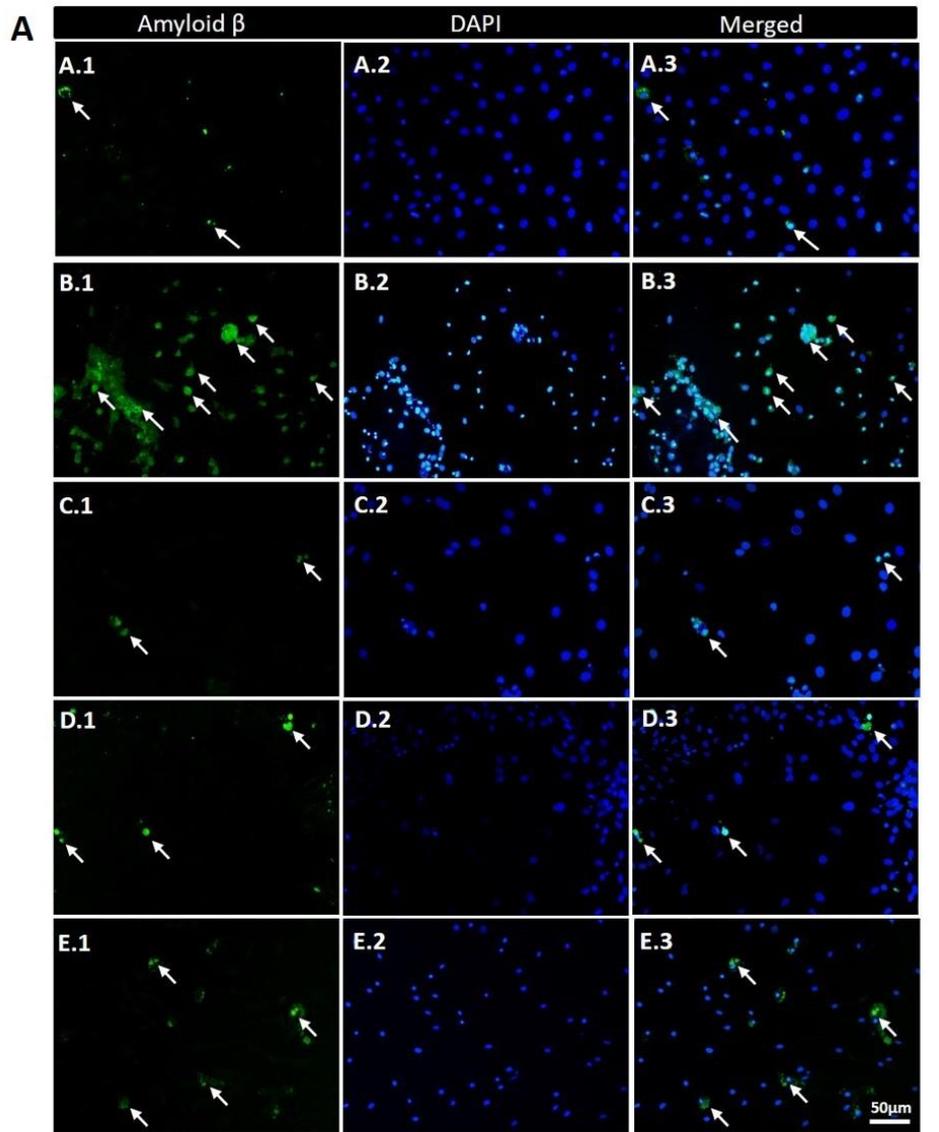
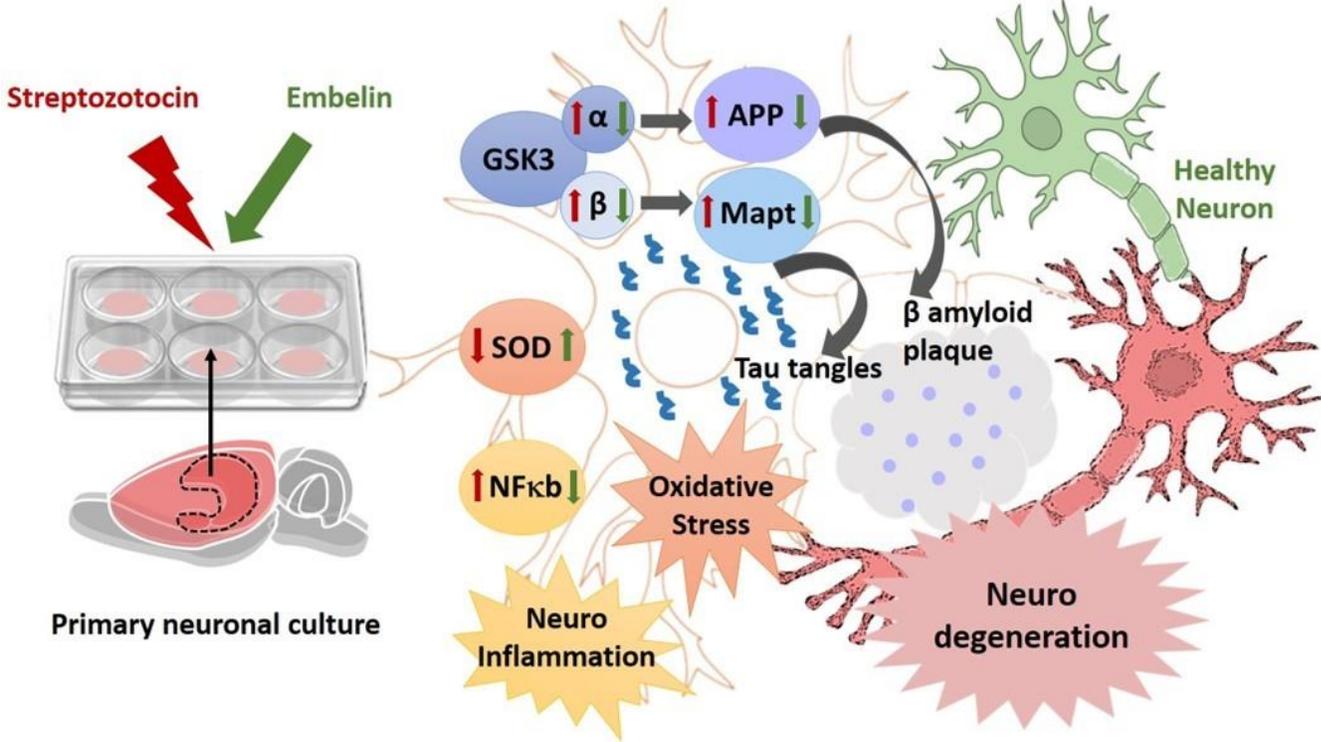


Figure 7



# Chapter 7

## 7.1 Introduction

The blood–brain barrier (BBB) permeability is an important factor to be considered during drug designing targeting central nervous system (73). This is due to the complexity nature of the BBB that restrict the entry of most therapeutics agents into the brain causing the compound to be pharmacologically ineffective (74). Thus, an *in vitro* BBB model become a useful tool as a permeability assay to measure the ability of compounds of interest to cross the BBB (75). Porcine brain endothelial cells (PBECs) model tend to retain most of their *in vivo* BBB characteristics when compared to *in vitro* BBB model (76). In this last part of our study, we reported BBB permeability of the embelin using the PBEC models. In addition to that, drug-like properties of embelin were evaluated using molecular docking studies in order to identify binding sites for AChE and A $\beta$  with embelin to further develop this compound into anti-AD drug.



1 Article

# 2 **Embelin, a potent molecule for Alzheimer's disease:** 3 **A proof of concept from BBB permeability, AChE** 4 **inhibition and molecular docking studies**

5 **Saatheeyavaane Bhuvanendran**<sup>1</sup>, **Nur Aziah Hanapi**<sup>2</sup>, **Nafees Ahmed**<sup>3,4</sup>, **Iekhsan Othman**<sup>1</sup>, **Siti**  
6 **Rafidah Yusof**<sup>2\*</sup> and **Mohd Farooq Shaikh**<sup>1\*</sup>

7 <sup>1</sup> Neuropharmacology Research laboratory, Jeffrey Cheah School of Medicine and Health Sciences, Monash  
8 University Malaysia, Bandar Sunway, Selangor, Malaysia

9 <sup>2</sup> Centre for Drug Research, Universiti Sains Malaysia, Penang, Malaysia

10 <sup>3</sup> School of Pharmacy, Monash University Malaysia, Bandar Sunway, Selangor, Malaysia

11 <sup>4</sup> Tropical Medicine and Biology Multidisciplinary Platform, Monash University Malaysia, Bandar Sunway,  
12 Selangor, Malaysia

13

14 \* Correspondence: farooq.shaikh@monash.edu; Tel.: +603-55144483 (M.F.S)

15 sryusof@usm.my; Tel.: +604-653 2141 (S.R. Y)

16 **Abstract:** Embelin is well known in ethnomedicine and reported to have central nervous system  
17 activities. However, there is no report on blood-brain barrier (BBB) permeability of embelin. Here  
18 the BBB permeability of embelin was evaluated using *in vitro* primary porcine brain endothelial cell  
19 (PBEC) model of the BBB. Embelin was also evaluated for acetylcholinesterase (AChE) inhibitory  
20 activity and docking prediction for interaction with AChE and amyloid beta (A $\beta$ ) binding sites.  
21 Embelin was found to be non-toxic to the PBECs and did not disturb the PBEC tight junction  
22 function. The PBECs showed restrictive tight junctions with average transendothelial electrical  
23 resistance of 365.37 $\pm$ 37.67  $\Omega$ .cm<sup>2</sup>. Permeability assays were conducted from apical-to-basolateral  
24 direction (blood-to-brain side). Embelin showed apparent permeability ( $P_{app}$ ) value of 35.46 $\pm$ 9.09  $\times$   
25  $10^{-6}$  cm/s with 85.53% recovery. *In vitro* AChE inhibitory assay demonstrated that embelin could  
26 inhibit the enzyme. Molecular docking study showed that embelin binds well to active site of AChE  
27 with CDOCKER interaction energy of -65.75 kcal/mol which correlates with the *in vitro* results.  
28 Docking of embelin with A $\beta$  peptides also revealed the promising binding with low CDOCKER  
29 interaction energy. Thus, findings from this study indicate that embelin could be a suitable molecule  
30 to be further developed as therapeutic molecule to treat neurological disorders particularly  
31 Alzheimer's disease.

32

33 **Keywords:** embelin, blood-brain barrier, permeability, acetylcholinesterase inhibitor, molecular  
34 docking, amyloid beta peptides

35

## 36 1. Introduction

37 The blood brain barrier (BBB) is highly selective interface that separates the brain and the central  
38 nervous system (CNS) from the bloodstream [1, 2]. The BBB is composed of brain endothelial cells  
39 that formed the cerebral microvasculature which are interconnected by tight junctions [2, 3]. The  
40 endothelium facilitates and regulates substance entry between the blood and the CNS, as well as  
41 protecting the brain from harmful toxins and pathogens. Unfortunately, the protective nature of the  
42 BBB becomes a disadvantage as it also restricts the entry of many potential therapeutic agents [4].  
43 Newly developed drugs targeting CNS disorders have the poorest success rate and often failed in  
44 clinical trial [5]. Around 98% of the potential drugs do not cross the BBB [6]. Due to their inability or

45 poor ability to cross BBB, they cannot be utilized for CNS related disorders [7] and this imposed major  
46 hurdles in pharmacological treatment of CNS disorders [8]. Therefore, it is very crucial to know  
47 whether a compound can cross the BBB and utilize this information during drug development before  
48 proceeding to clinical trial.

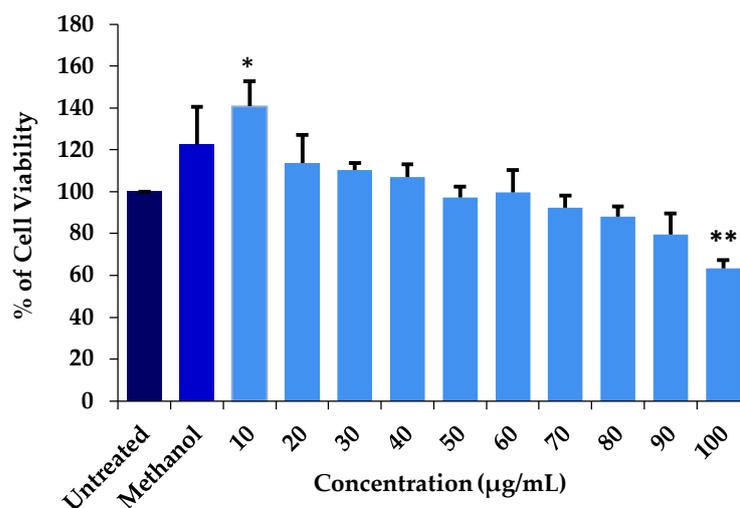
49 *In vivo* BBB methods provide the most reliable measurement for drug permeation due to the  
50 complex nature of the BBB, but with limitations of a low throughput and being labor intensive [9, 10].  
51 Thus, good *in vitro* BBB model which demonstrates restrictive tight junctions reflected by high  
52 transendothelial electrical resistance (TEER) [11] and resembles the *in vivo* conditions is very  
53 important for effective screening for BBB permeability in drug discoveries [10, 12]. Several studies  
54 have reported on *in vitro* BBB models from variety of species including from mice, rats, cows, pigs,  
55 and human [12-15]. However, some of the reported BBB models suffered from low TEER indicating  
56 leaky tight junctions [12]. For instance, the human cerebral microvascular endothelial cell line  
57 (hCMEC/D3) which showed TEER values of less than 50  $\Omega\cdot\text{cm}^2$  is probably not suitable for BBB  
58 permeability studies of small molecules even though it is of human origin [16-18]. *In vitro* BBB model  
59 from the PBECs has been reported to show well-developed tight junctions, polarized expression of  
60 functional transporters [19], which features comparable to that of human BBB. Additionally, the  
61 larger size of porcine brain compared to rodent brain enables higher cell yield, and it is relatively  
62 cheaper and more convenient to set up as porcine brains are by-product of the meat industry, and  
63 therefore do not require ethical approval [15, 20].

64 On the other hand, *in silico* modelling also allows for prediction of BBB permeation of  
65 compounds particularly for passive diffusion [9]. Modelling based on absorption, distribution,  
66 metabolism, excretion, and toxicity (ADMET)-related descriptors predicts the effectiveness and  
67 bioavailability of compounds based on pharmacokinetic properties [21]. Docking studies predict  
68 interaction between the compounds to their protein targets [22] which is also very crucial in drug  
69 designing. Recent reports indicated that embelin alleviates scopolamine-induced amnesia in rats and  
70 reversed memory impairment caused by streptozotocin (STZ) [23, 24]. However the BBB permeability  
71 of embelin and its mechanism of action are unknown. Here, assessment of embelin cytotoxicity, its  
72 effect on the BBB tight junction function and BBB permeability were performed using *in vitro* PBEC  
73 BBB model; its mechanism of action was determined using AChE inhibitory assay and docking  
74 studies, to investigate its potential as a new candidate for CNS therapeutic molecule particularly for  
75 the treatment of Alzheimer's disease (AD).

## 76 2. Results

### 77 2.1. Cytotoxicity of embelin towards the PBECs

78 Prior to the BBB permeability assay, viability of the PBECs in presence of embelin was  
79 determined. One-way ANOVA analysis shows a significant difference between the treatment groups  
80 and the cell viability ( $F=6.134$ ;  $P < 0.001$ ). As shown in **Figure 1**, the PBECs treated with embelin from  
81 10 to 90  $\mu\text{g/mL}$  did not show reduction in viability when compared to the untreated cells. However,  
82 embelin at 100  $\mu\text{g/mL}$  caused reduction in PBEC viability ( $P < 0.01$ ) compared to the untreated cells.

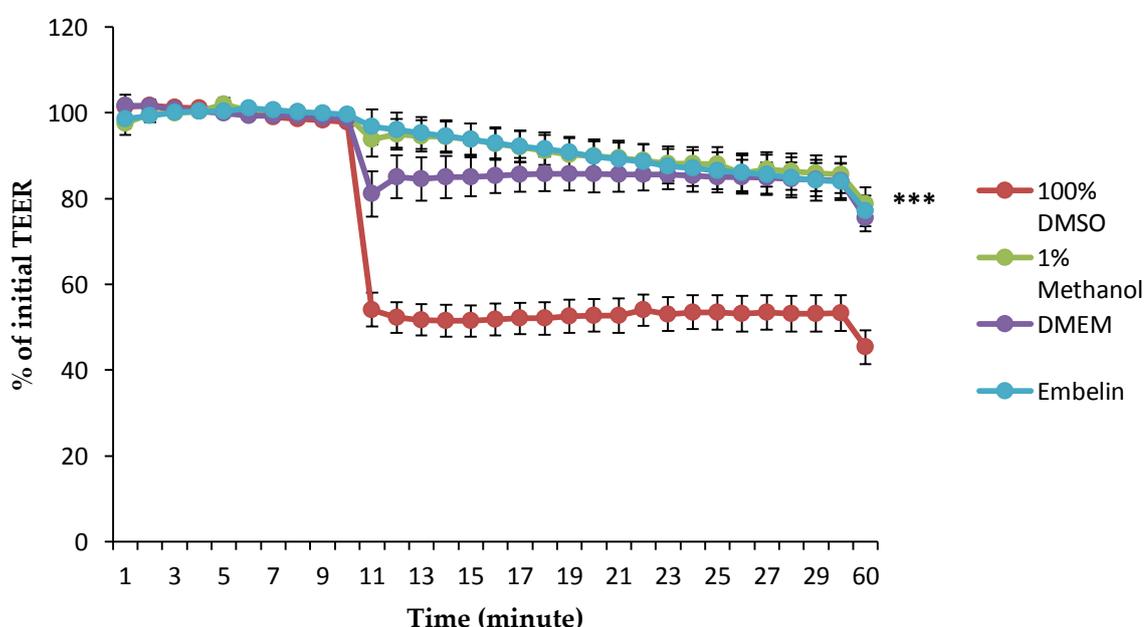


83

84 **Figure 1:** Effect of embelin at concentrations ranging from 10 - 100 µg/mL on PBECs viability, tested using (3-(4,  
85 5-dimethylthiazol-2-yl)-2, 5-diphenyltetrazolium bromide) MTT assay. Data are mean ± SEM, n = 3  
86 independent experiments. \*  $P < 0.05$  \*\*  $P < 0.01$ , as tested using One-way ANOVA followed by Dunnett test.

## 87 2.2 Real-time TEER assay

88 Tight junction integrity of the PBEC monolayer was determined by measuring the TEER at 1  
89 minute interval up to 60 minutes. Embelin was tested at 30 µg/mL, Dulbecco's modified Eagle's  
90 medium (DMEM) and dimethyl sulfoxide (DMSO) were used as negative and positive control  
91 respectively. As shown in **Figure 2**, embelin at 30 µg/mL did not disrupt the tight junction integrity  
92 and it was significant at  $P < 0.001$  when compared to 100% DMSO.



93

94 **Figure 2:** TEER across the PBEC monolayer was measured for 60 minutes at 1-minute interval using WPI STX-  
95 100C chopstick electrode pair connected to EVOM meter. Embelin (30 µg/mL), DMEM (negative control) and  
96 DMSO (positive control) were added separately to the inserts after minute 10 TEER was recorded. Data are mean  
97 ± SEM, n = 3 independent experiments.

98

### 99 2.3 *In vitro* BBB permeability assay

100 Permeability assay is conducted to measure the rate of BBB crossing for compounds. In this  
 101 study, the rate at which embelin transverse across the PBEC monolayer from apical to basolateral  
 102 direction (blood to brain side) was measured and reported as apparent permeability ( $P_{app}$ , cm/s). As  
 103 shown in **Table 1**, embelin demonstrated  $P_{app}$  value of  $35.46 \pm 9.09 \times 10^{-6}$  cm/s with 83.53% recovery.  
 104 Sodium fluorescein (NaF) as paracellular permeability marker compound showed low  $P_{app}$  of  
 105  $2.47 \pm 0.82 \times 10^{-6}$  cm/s, indicating that the tight junctional integrity was preserved during the assay.

106 **Table 1:**  $P_{app}$  values and % recovery of embelin and NaF

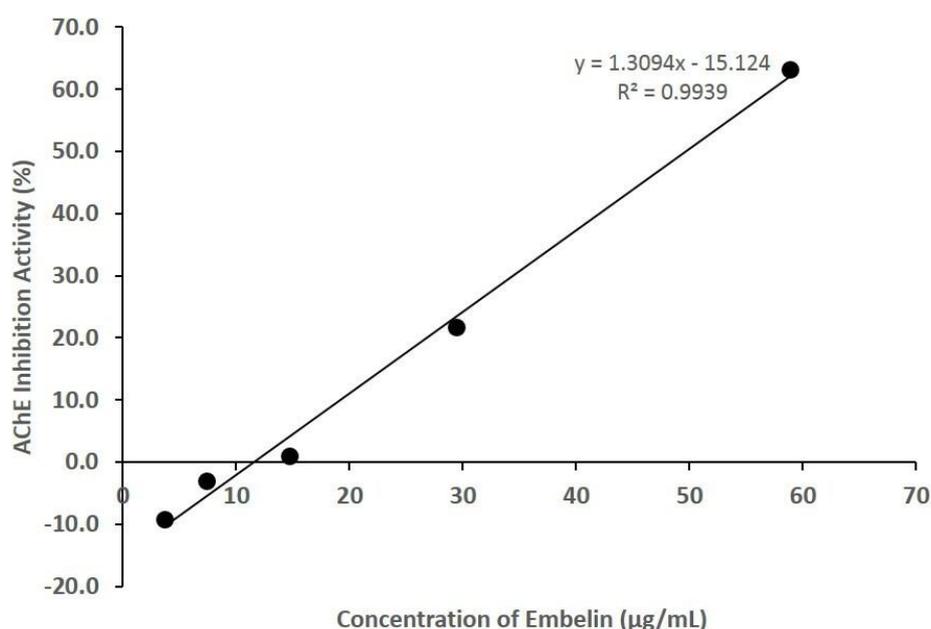
Insert	$P_{app}$ ( $10^{-6}$ cm/s)	% recovery
Embelin	$35.46 \pm 9.09$	$83.53 \pm 6.58$
NaF	$2.47 \pm 0.82$	$78.16 \pm 1.81$

107

### 108 2.4 AChE inhibitory assay

109 Embelin was evaluated for its inhibitory activity of AChE from electric eel (*Electrophorus*  
 110 *electricus*). Donepezil was used as a positive control and to validate the assay by comparing  $IC_{50}$  value  
 111 obtained in this study with reported values [25]. Embelin was tested at a series of concentration from  
 112 3.68 to 58.9  $\mu$ g/mL in order to determine the  $IC_{50}$  value using standard curve generated using  
 113 Microsoft Excel. As shown in Figure 3,  $IC_{50}$  value obtained for embelin against AChE is 4974  $\mu$ g/mL.

114



115

116 **Figure 3:** The anti-cholinesterase activity of embelin (3.68 - 58.9  $\mu$ g/mL) using an *in-vitro* AChE inhibitory  
 117 assay. The graph was plotted by keepin embelin concentration on X-axis against AChE inhibition activity (%)  
 118 on Y-axis.  $IC_{50}$  values was calculated using standard curve generated using Microsoft Excel.

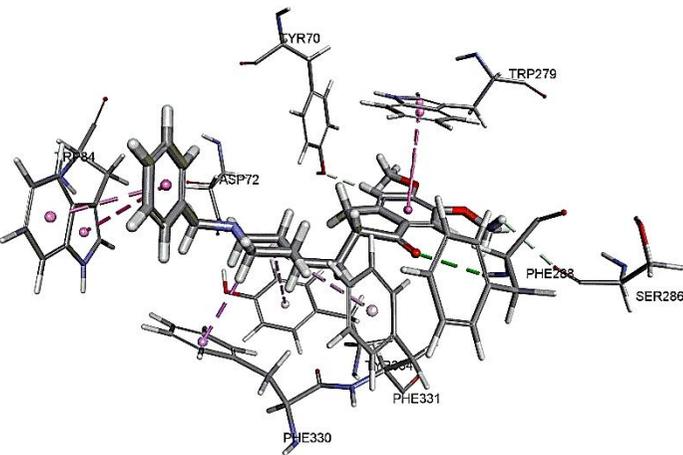
119

120 2.5 Molecular docking

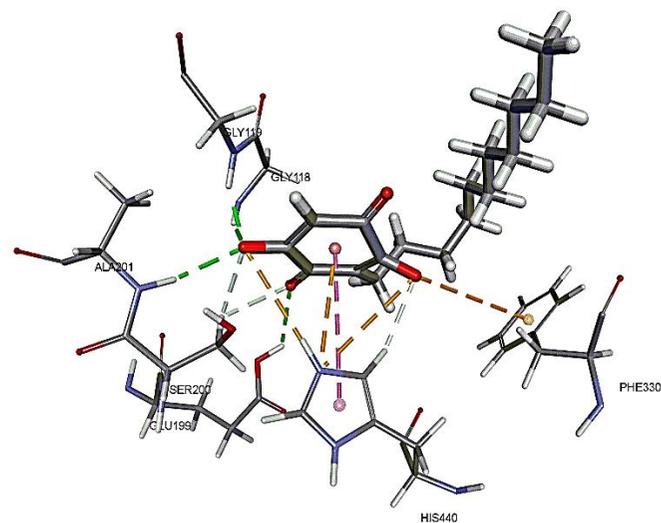
121 The results for docking studies are expressed as interaction energy in -kcal/mol. The docked conformations of donepezil and embelin and key interactions are  
122 summarized in **Table 2**. Based on the results, embelin has better binding to the AChE active site with the interaction energy of -65.75 kcal/mol compared to E2020.  
123 Likewise, the docked conformations of embelin and A $\beta$  and key interactions are summarized in **Table 3**. Binding to fibril 6A $\beta$ <sub>9–40</sub> and 5A $\beta$ <sub>17–42</sub> display high  
124 interaction energy of -54.01 and -38.77 respectively when compared with A $\beta$  monomers.

125

**Table 2:** Binding modes of embelin docked to AChE active sites

Compounds	Docked pose	CDocker Interaction Energy (-kcal/mol)	Non-bond Interactions
E2020 (reference)		48.5319	Hydrogen Bonds E2020 to PHE288 E2020 to ASP72 E2020 to SER286 E2020 to TYR70  Hydrophobic interactions -Pi-Sigma E2020 to PHE330 E2020 to TRP84 E2020 to TRP279 E2020 to PHE331 E2020 to TYR334

**Embelin**

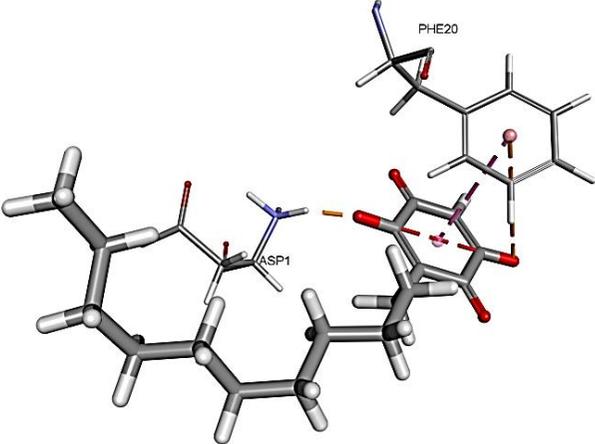
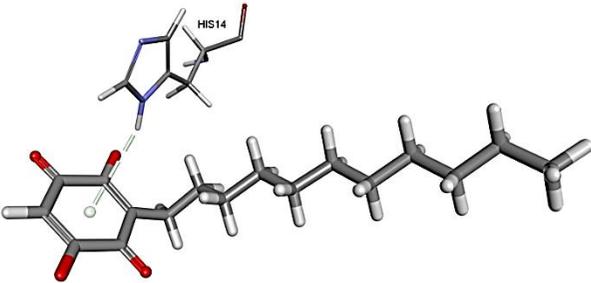


65.7525

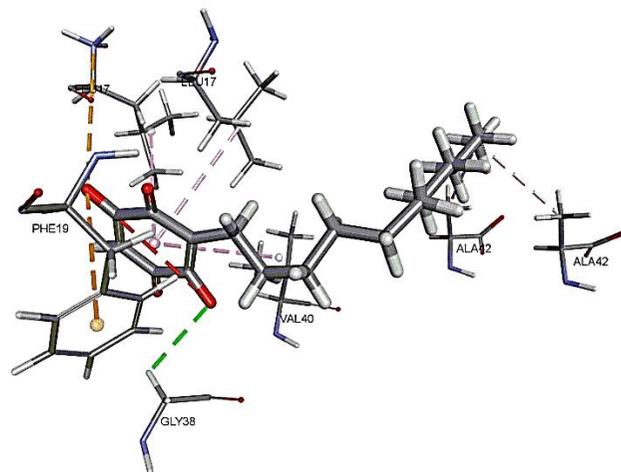
- Hydrogen Bonds
- Embelin to GLY118
- Embelin to GLY119
- Embelin to GLY199
- Embelin to ALA201
- Embelin to SER200
- Embelin to HIS440
  
- Hydrophobic interactions
- Pi-Pi
- Embelin to HIS440
  
- Electrostatic interactions
- HIS440 to Embelin
- Embelin to PHE330

126  
127  
128  
129  
130  
131

**Table 3:** Binding modes of embelin docked to A $\beta$  active sites

PDB ID	Docked pose	CDocker Interaction Energy (-kcal/mol)	Non-bond interactions
1BA4 (A $\beta$ 1-40)		34.1594	<p>Hydrogen Bonds Embelin to ASP1</p> <p>Hydrophobic interactions Embelin to PHE20</p> <p>Electrostatic interactions Embelin to PHE20</p>
1Z0Q (A $\beta$ 1-42)		24.2574	<p>Hydrogen Bonds Embelin to HIS14</p>

**2BEG**  
**(5A $\beta$  17-42)**



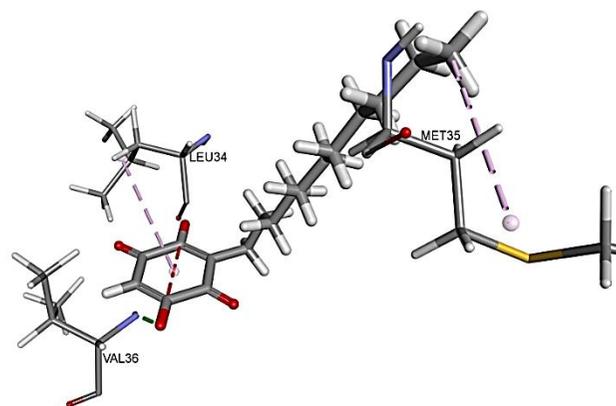
38.7666

Hydrogen Bonds  
Embelin to GLY38

Hydrophobic  
interactions  
Embelin to ALA42  
Embelin to ALA42  
Embelin to LEU17  
Embelin to LEU17  
Embelin to VAL40

Electrostatic  
interactions  
Embelin to PHE19

**2LMN**  
**(6A $\beta$  9-40)**



54.0122

Hydrogen Bond  
Embelin to VAL36

Hydrophobic  
interactions  
Embelin to MET35  
Embelin to LEU34



### 134 3. Discussion

135 To date, not a single study reported on BBB permeability of embelin [26]. According to Pathan  
136 et al. in order to cross the BBB, a compound should be in unionized form, molecular weight of less  
137 than 400 Da, log P value near to 2 and around 8-10 hydrogen bonds [8]. Embelin has all these  
138 characteristics, hence high possibility to permeate the BBB. Cell culture models are the most favored  
139 tools for assessing BBB permeation of compounds, giving information on passive permeability across  
140 cell membranes and also on carrier-mediated transport [27]. Therefore, we conducted *in vitro* BBB  
141 permeability assay of embelin using PBEC BBB model. Prior to the permeability assay, we established  
142 that embelin does not cause any toxicity to the PBECs up to 90 µg/mL, and embelin at 30 µg/mL does  
143 not affect BBB tight junctional integrity compared to the 100% DMSO. Based on our results, cells  
144 treated with methanol and embelin at 10 µg/mL have higher cell viability when compared to other  
145 groups. The increase in cell viability is due to stochastic effect which occurs when small stress (low  
146 concentration) is applied to the cells. However, stronger stress will disrupt the tight junctions and  
147 eventually will lead to cell death. Thus, this supported our results that cell viability decreased on cells  
148 treated with higher concentration of embelin. For the permeability assay, cell monolayer with TEER  
149 values exceeding 200 Ω.cm<sup>2</sup> was used as the cells were considered to have minimal tight junction  
150 leakiness [28]. From the results, embelin demonstrated high  $P_{app}$  value of  $35.46 \pm 9.09 \times 10^{-6}$  cm/s. The  
151  $P_{app}$  value is comparable to that of donepezil ( $30.6 \pm 9.09 \times 10^{-6}$  cm/s), reported by Liew et al. [11].

152 The high  $P_{app}$  value of embelin could consists of one or a combination of routes used by the  
153 compound to cross the BBB. Embelin could permeate via passive transcellular route across the cell  
154 membrane only or at the same time facilitated by membrane transporter expressed on cell  
155 membranes. To further dissect the mechanisms involved, bidirectional permeability assay could be  
156 conducted [11]. Additionally,  $P_{app}$  value of embelin is much higher than  $P_{app}$  of the paracellular  
157 marker compound used in this study i.e. NaF with  $P_{app}$  of  $2.47 \pm 0.82 \times 10^{-6}$  cm/s. This could indicate  
158 that embelin largely cross the BBB via transcellular route and not paracellular route (via the tight  
159 junction) *in vitro*. This is further supported by the outcome of ADMET for BBB penetration for  
160 embelin which is level 1. According to Ponnan et al. [21], ADMET BBB penetration level 1 indicates  
161 high penetration of a compound across the BBB after an oral administration.

162 The CDocker was used for docking of all compounds. The CDocker is CHARMM-based  
163 docking algorithm that uses the CHARMM family of force fields and offers all the advantages of full  
164 ligand flexibility (including bonds, angles, and dihedrals) and reasonable computation times [29].  
165 The CDocker algorithm adopts a strategy involving the generation of several initial ligand  
166 orientations in the active site of target protein followed by molecular dynamics based simulated  
167 annealing and final refinement by energy [30].

168 The molecular docking study was carried out to understand the binding mode of embelin within  
169 the active site of AChE using Discovery Studio suit 4.5 software. The x-ray crystal structure of AChE  
170 complexed with donepezil (or E2020) was retrieved from Protein Data Bank (PDB code: 1EVE). To  
171 validate the docking protocol, donepezil was first docked into AChE active site. As revealed by  
172 Kryger, et al. [31] phenyl ring of E2020 form  $\pi$ -stacking with Trp 84 and Phe 330 while another  
173 aromatic ring stacked with Trp279. Further, hydrogen bond was observed between Phe288 and  
174 ketone oxygen. The root mean square deviation (RMSD) and CDocker interaction energy (CDIE)  
175 were found to be 1.28Å and -48.53 kcal/mol respectively [31]. Embelin showed a promising favorable  
176 interaction with AChE binding site with CDocker interaction energy of -65.75 kcal/mol. This  
177 finding is consistent with AChE inhibitory activity for of embelin. Higher binding interaction energy  
178 indicating embelin may bound to the AChE active site which likely to trigger the catalytic site for its  
179 inhibitory activity for AChE [25].

180 Accumulation of research evidence over the last 20 years revealed that A $\beta$  oligomers is  
181 associated with AD pathogenesis [32]. Therefore, there is a pressing need to find compounds that are  
182 able to promote anti-A $\beta$  aggregation and clearance [33]. There are studies reported the potential of  
183 small molecules in converting toxic oligomers into non-toxic amorphous aggregates [34, 35].  
184 Furthermore, small molecules could also contribute in morphological changes of amyloid fibrils to  
185 inert form [36, 37]. Since A $\beta$  peptides are located in the brain, an efficient drug should be able to cross  
186 the BBB to interfere with their activities [33]. Similar to AChE docking study, embelin also interacted  
187 favourably with A $\beta$  peptides as evident from CDOCKER interaction energy as shown in **Table 3**.  
188 These results revealed that embelin has potential to bind with A $\beta$  peptides which may then slow  
189 down or degrade mature fibrils of A $\beta$  peptides.

## 190 4. Materials and Methods

### 191 4.1 Materials

192 Iscove's modified Dulbecco's medium (IMDM 1X), Dulbecco's modified Eagle's medium  
193 (DMEM) without Phenol Red, Hank's Balanced Salt Solution (HBSS) without calcium (Ca<sup>2+</sup>) and  
194 magnesium (Mg<sup>2+</sup>) and heat-inactivated fetal bovine serum (FBS) were purchased from Gibco Life  
195 Technologies (Grand Island, USA). Phosphodiesterase inhibitor (RO-20-1724) was obtained from  
196 Merck Chemicals Ltd. (Nottingham, UK). Corning Transwell® translucent polycarbonate filter inserts  
197 (product no. 3401, 12 mm diameter, 0.4  $\mu$ m pore size, 1  $\times$  10<sup>8</sup> pores/cm<sup>2</sup>, 1.12 cm<sup>2</sup> growth area) were  
198 obtained from Corning (New York, USA). All other chemicals were obtained from Sigma-Aldrich  
199 (Dorset, UK) unless otherwise stated.

### 200 4.2 Isolation of porcine brain microvessels and culture of the PBECs

201 The porcine brain microvessels were isolated using published method [10, 20] with slight  
202 modifications. Porcine brains from Department of Veterinary Services Penang abattoir (Sungai  
203 Pinang, Penang, Malaysia) were transported to the lab in ice-cold IMDM supplemented with FBS  
204 (10% v/v), penicillin (100 U/mL) and streptomycin (100  $\mu$ g/mL) on ice. The brains were stored at 4°C  
205 overnight prior to the isolation of microvessels due to schedule of animal slaughter at the abattoir.  
206 Microvessels obtained were stored in liquid nitrogen until further use. Here, the cryopreserved  
207 microvessels were thawed and cultured in flasks according to previous studies [10, 19] to obtain the  
208 PBECs. The PBECs were then passaged onto plates or Transwell® after 4 days in culture. For  
209 cytotoxicity assay, the PBECs were cultured in 96-well plates at a seeding density of 3.2 $\times$ 10<sup>4</sup> cells/well,  
210 while for TEER measurement and permeability assay, the PBECs were cultured on the Transwell®  
211 inserts at a density of 1 $\times$ 10<sup>5</sup> cells/insert.

212 When culturing in wells and inserts, culture medium used was DMEM (with Phenol Red; Sigma  
213 D5546) supplemented with 10% (v/v) FBS, penicillin (100 U/mL), streptomycin (100  $\mu$ g/mL), L-  
214 glutamine (2 mM) and heparin (125  $\mu$ g/mL). To further induce BBB differentiation of the PBECs  
215 cultured on the inserts, at confluency, the culture medium was replaced by serum-free medium with  
216 added hydrocortisone (550 nM). The PBECs were also treated with 8-(4-chlorophenylthio-cAMP) (250  
217  $\mu$ M) and phosphodiesterase inhibitor (RO-20-7024) (17.5  $\mu$ M) to increase tight junction tightness. Cell  
218 culture was conducted at 37°C in a humidified atmosphere with 5% CO<sub>2</sub> in air.

### 219 4.3 Cytotoxicity of embelin towards the PBECs

220 MTT assay was conducted as described by Mosmann [38] with slight modifications. Confluent  
221 PBECs in 96-well plate were incubated with embelin prepared in the culture medium at  
222 concentrations ranging from 10 to 100  $\mu$ g/mL, for 1 hour at 37°C. After 1 hour, embelin solution was  
223 discarded and the PBECs were incubated with 100  $\mu$ L MTT solution (1 mg/mL) prepared in DMEM  
224 without Phenol Red for 4 hours at 37°C. Untreated cells were used as control to represent total viable

225 cells. After 4 hours, the MTT solution was removed and replaced with 100  $\mu$ L of propan-2-ol to  
 226 dissolve formazan crystals formed. Absorbance was measured at 560 nm and 690 nm using Multiskan  
 227 Go Microplate Reader (Thermo Fisher Scientific, MA, USA). The experiment was conducted in  
 228 triplicate, in three independent experiments. Percentage of cell viability was calculated using  
 229 following equation:

$$230 \quad \% \text{ of cell viability} = \frac{(\text{Absorbance}_{560} - \text{Absorbance}_{690}) \text{ of treated cells}}{(\text{Absorbance}_{560} - \text{Absorbance}_{690}) \text{ of untreated cells}} \times 100 \quad (1)$$

#### 231 4.4 Real-time TEER assay

232 The assay was conducted to assess effect of embelin on the PBEC tight junction function.  
 233 Approximately 24 hours after the serum-free medium change and treatment with 8-CPT-cAMP and  
 234 RO-20-1724, TEER of the PBEC monolayer was measured using WPI STX-100C chopstick electrode  
 235 pair connected to EVOM meter (World Precision Instruments Inc., Sarasota, FL, USA) for 1 hour at 1  
 236 minute interval. After minute 10 TEER was recorded, embelin (30  $\mu$ g/mL), DMEM (negative control)  
 237 and DMSO (positive control) were added to inserts separately and the TEER measurement was  
 238 resumed until minute 60. TEER values of the cell monolayer were subtracted from value recorded for  
 239 blank insert (without cells) and multiplied by growth surface area as shown by the following  
 240 equation:

$$241 \quad \text{TEER } (\Omega \cdot \text{cm}^2) = (R_{\text{cell monolayer}} - R_{\text{blank}}) \times A \quad (2)$$

242 in which,  $R_{\text{cell monolayer}}$  is the resistance ( $\Omega$ ) of insert with cells,  $R_{\text{blank}}$  is the resistance ( $\Omega$ ) of blank insert  
 243 without cells and A is the surface area of insert (1.12  $\text{cm}^2$ ). For each insert, the TEER values obtained  
 244 at the different time points were then normalized to initial measurement at t = 0 minute, and results  
 245 are reported as percentage of initial TEER.  
 246

#### 247 4.5 *In vitro* BBB permeability assay

248 Cell monolayers with TEER values more than 200  $\Omega \cdot \text{cm}^2$  were selected for permeability assay.  
 249 Briefly, DMEM without Phenol Red supplemented with HEPES (25 mM) at pH 7.4 was used as buffer.  
 250 Embelin was dissolved in DMSO at 2 mg/mL and diluted in the buffer to obtain a concentration of 30  
 251  $\mu$ g/mL. NaF, a paracellular permeability marker compound was added to the embelin solution at  
 252 concentration of 5  $\mu$ M. To start the assay, the culture medium in the apical (filter insert) and the  
 253 basolateral (well) compartments was aspirated and the filter inserts containing the PBECs were  
 254 transferred to a 12-well plate containing the pre-warmed buffer on a shaker-incubator (THERMOstar,  
 255 BMG Labtech, Germany). To start the experiment, 500  $\mu$ L of the embelin solution was added to the  
 256 apical compartment. The assay was carried out at 37°C for 60 minutes under stirring condition at 150  
 257 rpm. At the end of the assay, samples were taken from each compartment (400  $\mu$ L from the apical  
 258 and 1200  $\mu$ L from the basolateral) for analysis.

259 The samples were processed using liquid-liquid extraction method using chloroform (organic  
 260 phase) to extract embelin from the buffer (aqueous phase), followed by drying using nitrogen gas.  
 261 When dried, methanol was added to tubes to re-dissolve embelin and the samples were analyzed  
 262 using liquid chromatography tandem mass spectrophotometry (LC-MS/MS). Fluorescence of NaF  
 263 was measured at 485 nm excitation and 535 nm emission using a fluorescence intensity plate reader  
 264 (CHAMELEON™ V, Hidex, Finland). Apparent permeability ( $P_{\text{app}}$ , cm/s) of embelin was calculated  
 265 using the following equation:

$$266 \quad P_{\text{app}} (x 10^{-6} \text{ cm/s}) = \frac{C_R \cdot V_R}{C_D \cdot V_D \cdot t \cdot A} V_D \quad (3)$$

267 in which  $C_R$  and  $C_D$  are embelin concentrations ( $\text{mol}/\text{cm}^3$ ) in the receiver and donor compartments  
 268 i.e. basolateral and apical compartment respectively,  $V_R$  and  $V_D$  are the volumes in the receiver  
 269 compartment (1500  $\mu\text{L}$ ) and the donor compartment (500  $\mu\text{L}$ ),  $t$  is the incubation time (60 minutes),  
 270 and  $A$  is the surface area of the filter insert (1.12  $\text{cm}^2$ ). Values obtained were divided by 60 to express  
 271 results in  $\text{cm}/\text{s}$ .

#### 272 4.6 LC-MS/MS for quantification of embelin

273 The concentrations of embelin in the apical and basolateral compartments from the BBB  
 274 permeability assay were quantified using LC-MS/MS. Standard solutions of embelin were prepared  
 275 in methanol with concentrations of 1, 2, 5, 7.5, and 10  $\mu\text{g}/\text{mL}$ . The standard solutions and samples  
 276 from the assay were injected at 10  $\mu\text{L}$  into Agilent 6410 Triple Quad LC/MS comprising ZORBAX  
 277 Eclipse plus C18 RRHD 2.1  $\times$  150 mm and 1.8  $\mu\text{m}$  column at a flow rate of 0.5  $\text{mL}/\text{min}$ . The mobile  
 278 phase was consisted of 0.1% formic acid in water (solvent A) and 0.1% formic acid in acetonitrile  
 279 (solvent B) with a total run time of 4 minutes. The gradient elution was set as (i) 0-1 minute, 75% B;  
 280 (ii) 1-2 minutes, 90% B; (iii) 2-3 minutes, 95% B; (iv) 3-4 minutes, 100% B. Electrospray ionization mass  
 281 spectrometry condition was programmed with gas temperature of 300 $^\circ\text{C}$ , nebulizer pressure of 40  
 282 psi, capillary voltage of 4000 V and drying gas flow at 10.0  $\text{L}/\text{minute}$ . The MS scan parameters had a  
 283 dwell time of 250 s with two products of 122.9 and 96 Da, performed in negative polarity mode.

#### 284 4.7 *In vitro* AChE inhibitory assay

285 AChE inhibition of embelin was evaluated using the Ellman's method [25, 39] with slight  
 286 modifications. A serial dilution of embelin which highest concentration lesser than 200  $\mu\text{M}$  was  
 287 prepared using DMSO and 0.1 M sodium phosphate buffer (pH 7.8), with DMSO final concentration  
 288 of less than 1% (v/v). Sodium phosphate buffer (140  $\mu\text{L}$ ) was added to 96-well plate followed by  
 289 sample solution (20  $\mu\text{L}$ ) and absorbance was measured at 412 nm. This reading served as blank. Then,  
 290 AChE enzyme from electric eel (0.2  $\text{U}/\text{mL}$ , 20  $\mu\text{L}$ ) was added to the wells and incubated for 15 minutes  
 291 at room temperature. Finally, 5,5'-dithiobis (2-nitrobenzoic acid) (DTNB) (3mM, 10  $\mu\text{L}$ ) was added,  
 292 followed by addition of acetylthiocholine iodide (ATCI) (15 mM, 10  $\mu\text{L}$ ). The rate of absorbance  
 293 change was measured at 412 nm for 30 minutes with a Multiskan Go Microplate Reader (Thermo  
 294 Fisher Scientific, MA, USA). Each assay was carried out with donepezil as positive control (0.015  $\mu\text{M}$ ).  
 295 The reactions were performed in three independent runs. Each run of a sample was performed in  
 296 triplicates and the  $\text{IC}_{50}$  values were determined from inhibition versus concentration plot. Below is  
 297 the equation to calculate AChE inhibition.

$$298 \text{ **Percentage inhibition** } (\%) = \left[ 1 - \left( \frac{\text{Sample}}{\text{Control}} \right) \right] \times 100 \quad (4)$$

#### 299 4.8 Molecular docking

300 All molecular docking studies were performed on Biovia Discovery Studio (BDS) 4.5  
 301 (www.3dsbiovia.com). For AChE, the x-ray crystal structure of AChE complexed with anti-  
 302 Alzheimer drug (donepezil or E2020) was retrieved from the Protein Data Bank (PDB code: 1EVE)  
 303 [31]. The water molecules were deleted, and hydrogen atoms were added. Finally, protein was  
 304 refined with CHARMM at physiological pH. To validate the docking reliability, co-crystallized ligand  
 305 donepezil was first re-docked to the binding site of AChE. Consequently, embelin was docked into  
 306 same active site; 30 conformations of the compound were obtained through CDOCKER. The  
 307 conformations with lowest energy were selected as the most probable binding conformation for each  
 308 ligand. Docking studies was further carried out with A $\beta$  peptide. Four receptors were chosen for A $\beta$   
 309 peptide docking including monomers A $\beta$ <sub>1-40</sub>, A $\beta$ <sub>1-42</sub> and fibril fragments 6A $\beta$ <sub>9-40</sub> and 5A $\beta$ <sub>17-42</sub> [33, 40-  
 310 42]. The structures of A $\beta$  were retrieved from Protein Data Bank and respective PDB ID are shown in  
 311 **Table 3**. Embelin was docked by using CDOCKER program. The BBB prediction for embelin was also  
 312 calculated using BDS 4.5.

313

## 314 4.9 Statistical analysis

315 Statistical analyses were performed using GraphPad Prism 5.0 software (La Jolla, CA). All data  
 316 are presented as mean  $\pm$  SEM and the samples were analyzed using One-way ANOVA followed by  
 317 Dunnett test. Statistical significance was reported as follows: \* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.001$

## 318 5. Conclusions

319 This study for the first time has demonstrated the use of *in vitro* PBEC BBB model in the evaluation  
 320 of embelin BBB permeability. This cell based model showed that embelin is able to cross the BBB  
 321 which further supported by *in silico* results. Besides that, this study has found embelin as a promising  
 322 AChE inhibitor as evidence from the AChE inhibition assay. Using molecular docking, we could  
 323 predict that embelin has favourable binding mode within the AChE and A $\beta$  peptide active sites.  
 324 Hence, based from this study we discovered that embelin is a favorable compound which can be  
 325 further developed into a potential therapeutic multipotent agent for AD.

326

327 **Author Contributions:** S.B conceived, performed experiments and wrote the manuscript. N.A.H helped in PBEC  
 328 *in vitro* studies and gave valuable input in writing the paper. N.A. performed molecular docking and helped in  
 329 writing of manuscript. I.O. was involved in LC-MS/MS and gave critical feedback for this study. S.R.Y and M.F.S.  
 330 were involved in conceptualization, designing the study, interpreted, supervised the study and contributed in  
 331 writing of the manuscript. All the authors read and approved the final manuscript.

332 **Funding:** This research received no external funding

333 **Acknowledgments:** The authors are grateful to Monash University Malaysia and Universiti Sains Malaysia for  
 334 the experimental laboratory support and facilities. SB was supported by Monash University Malaysia Merit  
 335 Scholarship.

336 **Conflicts of Interest:** The authors declare no conflict of interest.

## 337 Abbreviations

ADMET	Absorption, distribution, metabolism, excretion, and toxicity
AChE	Acetylcholinesterase
A $\beta$	Amyloid beta
ATCI	Acetylthiocholine iodide
BBB	Blood brain barrier
CNS	Central nervous system
CDIE	CDOCKER interaction energy
DMEM	Dulbecco's modified Eagle's medium
DMSO	Dimethyl sulfoxide
DTNB	5,5'-dithiobis (2-nitrobenzoic acid)
FBS	Fetal bovine serum
HBSS	Hank's Balanced Salt Solution
hCMEC/D3	Human cerebral microvascular endothelial cell line
IMDM	Iscove's modified Dulbecco's medium
LC-MS/MS	Liquid chromatography tandem mass spectrophotometry
MTT	3-(4, 5-dimethylthiazol-2-yl)-2, 5-diphenyltetrazolium bromide
NaF	Sodium fluorescein
P <sub>app</sub>	Apparent permeability
PBECs	Primary porcine brain endothelial cells
PDB	Protein Data Bank
RMSD	Root mean square deviation
STZ	Streptozotocin

TEER Transendothelial electrical resistance

338

339 **References**

- 340 1. Clark, D. E., In silico prediction of blood–brain barrier permeation. *Drug discovery today* **2003**, *8*, (20),  
341 927-933.
- 342 2. Abbott, N. J.; Patabendige, A. A.; Dolman, D. E.; Yusof, S. R.; Begley, D. J., Structure and function of the  
343 blood–brain barrier. *Neurobiology of disease* **2010**, *37*, (1), 13-25.
- 344 3. Daneman, R.; Prat, A., The blood–brain barrier. *Cold Spring Harbor perspectives in biology* **2015**, *7*, (1),  
345 a020412.
- 346 4. Czupalla, C. J.; Liebner, S.; Devraj, K., In vitro models of the blood–brain barrier. In *Cerebral*  
347 *Angiogenesis*, Springer: 2014; pp 415-437.
- 348 5. Ohtsuki, S.; Terasaki, T., Contribution of carrier-mediated transport systems to the blood–brain barrier  
349 as a supporting and protecting interface for the brain; importance for CNS drug discovery and  
350 development. *Pharmaceutical research* **2007**, *24*, (9), 1745-1758.
- 351 6. Devraj, K.; Klinger, M. E.; Myers, R. L.; Mokashi, A.; Hawkins, R. A.; Simpson, I. A., GLUT-1 glucose  
352 transporters in the blood–brain barrier: Differential phosphorylation. *Journal of neuroscience research*  
353 **2011**, *89*, (12), 1913-1925.
- 354 7. Pardridge, W. M., Crossing the blood-brain barrier: are we getting it right? *Drug discovery today* **2001**, *6*,  
355 (1), 1-2.
- 356 8. Pathan, S. A.; Iqbal, Z.; Zaidi, S.; Talegaonkar, S.; Vohra, D.; Jain, G. K.; Azeem, A.; Jain, N.; Lalani, J.  
357 R.; Khar, R. K., CNS drug delivery systems: novel approaches. *Recent patents on drug delivery &*  
358 *formulation* **2009**, *3*, (1), 71-89.
- 359 9. Abbott, N. J., Prediction of blood–brain barrier permeation in drug discovery from in vivo, in vitro and  
360 in silico models. *Drug Discovery Today: Technologies* **2004**, *1*, (4), 407-416.
- 361 10. Patabendige, A.; Skinner, R. A.; Abbott, N. J., Establishment of a simplified in vitro porcine blood–brain  
362 barrier model with high transendothelial electrical resistance. *Brain research* **2013**, 1521, 1-15.
- 363 11. Liew, K.-F.; Hanapi, N. A.; Chan, K.-L.; Yusof, S. R.; Lee, C.-Y., Assessment of the Blood-Brain Barrier  
364 Permeability of Potential Neuroprotective Aurones in Parallel Artificial Membrane Permeability Assay  
365 and Porcine Brain Endothelial Cell Models. *Journal of pharmaceutical sciences* **2017**, *106*, (2), 502-510.
- 366 12. Yusof, S. R.; Avdeef, A.; Abbott, N. J., In vitro porcine blood–brain barrier model for permeability  
367 studies: pCEL-X software pKaFLUX method for aqueous boundary layer correction and detailed data  
368 analysis. *European Journal of Pharmaceutical Sciences* **2014**, *65*, 98-111.
- 369 13. Xue, Q.; Liu, Y.; Qi, H.; Ma, Q.; Xu, L.; Chen, W.; Chen, G.; Xu, X., A novel brain neurovascular unit  
370 model with neurons, astrocytes and microvascular endothelial cells of rat. *International journal of*  
371 *biological sciences* **2013**, *9*, (2), 174.
- 372 14. Franke, H.; Galla, H.-J.; Beuckmann, C. T., Primary cultures of brain microvessel endothelial cells: a  
373 valid and flexible model to study drug transport through the blood–brain barrier in vitro. *Brain Research*  
374 *Protocols* **2000**, *5*, (3), 248-256.
- 375 15. Thomsen, L. B.; Burkhart, A.; Moos, T., A triple culture model of the blood-brain barrier using porcine  
376 brain endothelial cells, astrocytes and pericytes. *PloS one* **2015**, *10*, (8), e0134765.
- 377 16. Weksler, B.; Romero, I. A.; Couraud, P.-O., The hCMEC/D3 cell line as a model of the human blood  
378 brain barrier. *Fluids and Barriers of the CNS* **2013**, *10*, (1), 16.

- 379 17. Eigenmann, D. E.; Xue, G.; Kim, K. S.; Moses, A. V.; Hamburger, M.; Oufir, M., Comparative study of  
380 four immortalized human brain capillary endothelial cell lines, hCMEC/D3, hBMEC, TY10, and BB19,  
381 and optimization of culture conditions, for an in vitro blood–brain barrier model for drug permeability  
382 studies. *Fluids and Barriers of the CNS* **2013**, *10*, (1), 33.
- 383 18. Behrens, M.; Hüwel, S.; Galla, H.-J.; Humpf, H.-U., Blood-brain barrier effects of the Fusarium  
384 mycotoxins deoxynivalenol, 3 acetyldeoxynivalenol, and moniliformin and their transfer to the brain.  
385 *PloS one* **2015**, *10*, (11), e0143640.
- 386 19. Patabendige, A.; Abbott, N. J., Primary porcine brain microvessel endothelial cell isolation and culture.  
387 *Current protocols in neuroscience* **2014**, *69*, (1), 3.27. 1-3.27. 17.
- 388 20. Patabendige, A.; Skinner, R. A.; Morgan, L.; Abbott, N. J., A detailed method for preparation of a  
389 functional and flexible blood–brain barrier model using porcine brain endothelial cells. *Brain research*  
390 **2013**, *1521*, 16-30.
- 391 21. Ponnann, P.; Gupta, S.; Chopra, M.; Tandon, R.; Baghel, A. S.; Gupta, G.; Prasad, A. K.; Rastogi, R. C.;  
392 Bose, M.; Raj, H. G., 2D-QSAR, docking studies, and in silico ADMET prediction of polyphenolic  
393 acetates as substrates for protein acetyltransferase function of glutamine synthetase of Mycobacterium  
394 tuberculosis. *ISRN Structural Biology* **2013**, 2013.
- 395 22. Kitchen, D. B.; Decornez, H.; Furr, J. R.; Bajorath, J., Docking and scoring in virtual screening for drug  
396 discovery: methods and applications. *Nature reviews Drug discovery* **2004**, *3*, (11), 935.
- 397 23. Arora, R.; Deshmukh, R., Embelin Attenuates Intracerebroventricular Streptozotocin-Induced  
398 Behavioral, Biochemical, and Neurochemical Abnormalities in Rats. *Molecular neurobiology* **2017**, *54*, (9),  
399 6670-6680.
- 400 24. Bhuvanendran, S.; Kumari, Y.; Othman, I.; Shaikh, M. F., Amelioration of cognitive deficit by embelin  
401 in a scopolamine-induced Alzheimer’s disease-like condition in a rat model. *Frontiers in pharmacology*  
402 **2018**, *9*.
- 403 25. Liew, K.-F.; Chan, K.-L.; Lee, C.-Y., Blood–brain barrier permeable anticholinesterase aurones:  
404 Synthesis, structure–activity relationship, and drug-like properties. *European journal of medicinal*  
405 *chemistry* **2015**, *94*, 195-210.
- 406 26. Kundap, U. P.; Bhuvanendran, S.; Kumari, Y.; Othman, I.; Shaikh, M., Plant derived phyto compound,  
407 embelin in CNS disorders: a systematic review. *Frontiers in pharmacology* **2017**, *8*, 76.
- 408 27. Hakkarainen, J. J.; Jalkanen, A. J.; Kääriäinen, T. M.; Keski-Rahkonen, P.; Venäläinen, T.; Hokkanen, J.;  
409 Mönkkönen, J.; Suhonen, M.; Forsberg, M. M., Comparison of in vitro cell models in predicting in vivo  
410 brain entry of drugs. *International journal of pharmaceutics* **2010**, *402*, (1-2), 27-36.
- 411 28. Gaillard, P. J.; de Boer, A. G., Relationship between permeability status of the blood–brain barrier and  
412 in vitro permeability coefficient of a drug. *European journal of pharmaceutical sciences* **2000**, *12*, (2), 95-102.
- 413 29. Brooks, B. R.; Bruccoleri, R. E.; Olafson, B. D.; States, D. J.; Swaminathan, S. a.; Karplus, M., CHARMM:  
414 a program for macromolecular energy, minimization, and dynamics calculations. *Journal of*  
415 *computational chemistry* **1983**, *4*, (2), 187-217.
- 416 30. Mo, S.-L.; Liu, W.-F.; Li, C.-G.; Zhou, Z.-W.; Luo, H.-B.; Chew, H.; Liang, J.; Zhou, S.-F., Pharmacophore,  
417 QSAR, and binding mode studies of substrates of human cytochrome P450 2D6 (CYP2D6) using  
418 molecular docking and virtual mutations and an application to chinese herbal medicine screening.  
419 *Current pharmaceutical biotechnology* **2012**, *13*, (9), 1640-1704.
- 420 31. Kryger, G.; Silman, I.; Sussman, J. L., Structure of acetylcholinesterase complexed with E2020  
421 (Aricept®): implications for the design of new anti-Alzheimer drugs. *Structure* **1999**, *7*, (3), 297-307.

- 422 32. Hayden, E. Y.; Teplow, D. B., Amyloid  $\beta$ -protein oligomers and Alzheimer's disease. *Alzheimer's*  
423 *research & therapy* **2013**, 5, (6), 60.
- 424 33. Ngo, S. T.; Li, M. S., Top-leads from natural products for treatment of Alzheimer's disease: docking and  
425 molecular dynamics study. *Molecular Simulation* **2013**, 39, (4), 279-291.
- 426 34. Ehrnhoefer, D. E.; Bieschke, J.; Boeddrich, A.; Herbst, M.; Masino, L.; Lurz, R.; Engemann, S.; Pastore,  
427 A.; Wanker, E. E., EGCG redirects amyloidogenic polypeptides into unstructured, off-pathway  
428 oligomers. *Nature structural & molecular biology* **2008**, 15, (6), 558.
- 429 35. Bieschke, J.; Russ, J.; Friedrich, R. P.; Ehrnhoefer, D. E.; Wobst, H.; Neugebauer, K.; Wanker, E. E., EGCG  
430 remodels mature  $\alpha$ -synuclein and amyloid- $\beta$  fibrils and reduces cellular toxicity. *Proceedings of the*  
431 *National Academy of Sciences* **2010**, 107, (17), 7710-7715.
- 432 36. Dzwolak, W.; Grudzielanek, S.; Smirnovas, V.; Ravindra, R.; Nicolini, C.; Jansen, R.; Lokszejn, A.;  
433 Porowski, S.; Winter, R., Ethanol-perturbed amyloidogenic self-assembly of insulin: looking for origins  
434 of amyloid strains. *Biochemistry* **2005**, 44, (25), 8948-8958.
- 435 37. Sibley, S. P.; Sosinsky, K.; Gulian, L. E.; Gibbs, E. J.; Pasternack, R. F., Probing the mechanism of insulin  
436 aggregation with added metalloporphyrins. *Biochemistry* **2008**, 47, (9), 2858-2865.
- 437 38. Mosmann, T., Rapid colorimetric assay for cellular growth and survival: application to proliferation  
438 and cytotoxicity assays. *Journal of immunological methods* **1983**, 65, (1-2), 55-63.
- 439 39. Ellman, G. L.; Courtney, K. D.; Andres Jr, V.; Featherstone, R. M., A new and rapid colorimetric  
440 determination of acetylcholinesterase activity. *Biochemical pharmacology* **1961**, 7, (2), 88-95.
- 441 40. Petkova, A. T.; Ishii, Y.; Balbach, J. J.; Antzutkin, O. N.; Leapman, R. D.; Delaglio, F.; Tycko, R., A  
442 structural model for Alzheimer's  $\beta$ -amyloid fibrils based on experimental constraints from solid state  
443 NMR. *Proceedings of the National Academy of Sciences* **2002**, 99, (26), 16742-16747.
- 444 41. Petkova, A. T.; Yau, W.-M.; Tycko, R., Experimental constraints on quaternary structure in Alzheimer's  
445  $\beta$ -amyloid fibrils. *Biochemistry* **2006**, 45, (2), 498-512.
- 446 42. Lührs, T.; Ritter, C.; Adrian, M.; Riek-Loher, D.; Bohrmann, B.; Döbeli, H.; Schubert, D.; Riek, R., 3D  
447 structure of Alzheimer's amyloid- $\beta$  (1-42) fibrils. *Proceedings of the National Academy of Sciences* **2005**,  
448 102, (48), 17342-17347.
- 449



© 2018 by the authors. Submitted for possible open access publication under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).

# Chapter 8

## 8.1 Strength and Limitations

Till date, there is no cure or effective treatment for AD. Currently available therapies do not completely eradicate all the symptoms of AD but show no to little improvements in cognitive functions which are greatly affected in AD (77). Majority of the potential anti-AD drug candidates failed at early phase of the clinical trials due to their ability to target a single mechanism of AD pathogenesis. This is one of the major problem in the drug development phase as existing preclinical approaches does not allow to study the complexity of the disease using one single disease model. Presently, researchers focus on the existing hypotheses which emphasize on a single factor at a time that contributes to AD (78). As mentioned in the previous chapters, AD is a multifactorial condition caused by disparity of a collective action of several factors (19). Thus, it is impossible to halt or restore AD back to normal state by just aiming at a single target rather than a multi-targeted approach.

Therefore, in the current project, the neuroprotective potential of embelin was explored by using varied experimental models of AD. Animal models representing different hypotheses were deployed to discover whether embelin has the ability to work through multiple mechanisms and affect positively to prevent the genesis of the key hallmarks of the AD. Moreover, this project also focused on the understanding the BBB permeability of embelin. BBB permeability is another key factor, which failed many potential drug candidates in the translational phase from basic screening to preclinical phase in the anti-AD drug development. To the best of our knowledge, this is the first study of embelin's BBB permeability using an *in-vitro* PBECs cell model.

The approach for the assessment of anti-AD effect of embelin was comprehensive ranging from biochemical, behavioral, pharmacological and molecular docking studies. The outcomes demonstrated excellent research findings as embelin has a remarkable improvement in learning and memory in both cholinergic and vascular dysfunction rat models. Likewise, this study also revealed that embelin has the potential to down-regulate the APP and MAPT mRNA expression as well as prevented over expression of amyloid beta protein induced by STZ in an *in-vitro* model of AD. As mentioned in the previous chapters, STZ induced sporadic AD-like condition, which reflects amyloid and tau hypothesis of AD. Furthermore, the molecular docking study predicted that embelin has a favorable binding at the active sites namely AChE and amyloid beta, which are among key hallmarks of AD. Besides that, based on docking study using ADMET, we found that

BBB penetration level for embelin was at level 1. This result predicted the ability of embelin to cross BBB, which are very much in-line with the *in vitro* results using PBECs cell model.

Thus, it is found that embelin could act through multiple mechanisms in AD pathogenesis. Relevant to this PhD thesis, we reported our findings and submitted to four different peer-reviewed journals. Out of that four, one has been successfully published in *Frontiers in Pharmacology* and the other three are currently under review.

The main limitation on the current work is that we designed the study based on the different experimental model to test each hypothesis instead of just using a single experimental model. As AD is a complex neurodegenerative disorder that is affected by the multifactorial mechanism in the pathogenesis of the disease. In the past few years, transgenic rodent became a promising model as many studies have been carried out to describe the disease progression as it can mimics various characteristic at a time which closely resemble human AD (79). However, most of the transgenic AD models created by genetic modifications rely on the early onset familial AD forms which only represent 5% of AD cases (26). Whereas the majority of AD cases are sporadic in nature for which no ideal animal model has been developed so far. Therefore, it would be insignificant to evaluate the effectiveness of potential drug candidates using a transgenic model as it can mimic only the early onset type familial AD form.

Another limitation in this study is that the vascular dementia and experiments using PBOCCA rats are not completely linked to AD but it is more general to dementia. Based on our results, embelin is neuroprotective on CCH-induced PBOCCA rats. Therefore, this compound may be promising for other types of dementia as well since it possesses several effects on the brain. On the other hand, we used STZ induced *in-vitro* rat primary hippocampal neuronal cultures as a model to address the amyloid hypothesis. However, it would have been good if we also use *in vivo* STZ model to link the *in vitro* results with the animal behavior to further support the effectiveness of embelin in AD.

Another limitation to the current study is uni-directional permeability assay using PBECs cell model, which is from apical to basolateral (blood to the brain). Although it is reported that embelin can cross the blood brain barrier, but the mechanism involved in BBB permeation of embelin was not explained. The bi-directional permeability assay which involved both apical to basolateral and

vice versa could clarify the potential involvement of any carrier-mediated or efflux protein in the BBB permeation of embelin (80).

## **8.2 Future work**

The following is recommended as important and promising research directions for developing embelin as a potential anti-AD therapeutic drug.

- a) Whilst embelin has been studied in various CNS related disorders, no study have been done on human due to lack of knowledge on safety and toxicity profiles. Therefore, establishing safety profiles for embelin is very crucial because it helps to minimize risk before moving forward to clinical trials.
- b) Evaluating embelin in transgenic model could be another option to confirm its effectiveness.
- c) Bidirectional permeability assay could be conducted in future to further explore the mechanisms involved in embelin BBB permeation.

## **8.3 Conclusion**

This study has demonstrated the use of various experimental models addressing AD-related hypothesis for the evaluation of embelin as a therapeutic compound. In conclusion, the outcomes of this study give credence to embelin as a potential multi-targeted drug candidate for AD.

# Chapter 9

## 9.0 References

1. Shah RS, Lee H-G, Xiongwei Z, Perry G, Smith MA, Castellani RJ. Current approaches in the treatment of Alzheimer's disease. *Biomedicine & Pharmacotherapy*. 2008;62(4):199-207.
2. Schachter AS, Davis KL. Alzheimer's disease. *Dialogues Clin Neurosci*. 2000;2(2):91-100.
3. Association As. 2018 Alzheimer's disease facts and figures. *Alzheimer's & Dementia*. 2018;14(3):367-429.
4. Alzheimer A. Uber eine eigenartige Erkrankung der Hirnrinde. *Zentralbl Nervenpsych*. 1907;18:177-9.
5. Braak H, Braak E, Bohl J, Reintjes R. Age, neurofibrillary changes, A $\beta$ -amyloid and the onset of Alzheimer's disease. *Neuroscience letters*. 1996;210(2):87-90.
6. Anonymous. World Alzheimer's Report (The state of the art of dementia research: New frontiers). *Alzheimer's Disease International*. 2018:1-48.
7. Bhardwaj D, Mitra C, Narasimhulu CA, Riad A, Doomra M, Parthasarathy S. Alzheimer's Disease—Current Status and Future Directions. *Journal of medicinal food*. 2017;20(12):1141-51.
8. Prince M, Wimo A, Guerchet M, Ali GC, Wu Y, AM P. World Alzheimer Report 2015: The Global Impact of Dementia. An Analysis of Prevalence, Incidence, Costs and Trends. *Alzheimer's Disease International*. 2015.
9. Nuri M, Huda T, Hong YH, Ming LC, Mohd Joffry S, Othman MF, et al. Knowledge on Alzheimer's disease among public hospitals and health clinics pharmacists in the State of Selangor, Malaysia. *Frontiers in pharmacology*. 2017;8:739.
10. PACIFIC A. DEMENTIA IN THE ASIA PACIFIC REGION: THE EPIDEMIC IS HERE. 2006.
11. Tey NP, Siraj SB, Kamaruzzaman SBB, Chin AV, Tan MP, Sinnappan GS, et al. Aging in multi-ethnic Malaysia. *The Gerontologist*. 2015;56(4):603-9.
12. Association As. Stages of Alzheimer's [cited 2018 22 November]. Available from: <https://www.alz.org/alzheimers-dementia/stages>.
13. Morris J, Storandt M, McKeel D, Rubin E, Price J, Grant E, et al. Cerebral amyloid deposition and diffuse plaques in "normal" aging Evidence for presymptomatic and very mild Alzheimer's disease. *Neurology*. 1996;46(3):707-19.

14. Petersen RC, Smith GE, Waring SC, Ivnik RJ, Tangalos EG, Kokmen E. Mild cognitive impairment: clinical characterization and outcome. *Archives of neurology*. 1999;56(3):303-8.
15. Selkoe DJ, Schenk D. Alzheimer's disease: molecular understanding predicts amyloid-based therapeutics. *Annual review of pharmacology and toxicology*. 2003;43(1):545-84.
16. Ward A, Tardiff S, Dye C, Arrighi HM. Rate of conversion from prodromal Alzheimer's disease to Alzheimer's dementia: a systematic review of the literature. *Dementia and geriatric cognitive disorders extra*. 2013;3(1):320-32.
17. Association As. Medications for Memory [cited 2018 22 November]. Available from: <https://www.alz.org/alzheimers-dementia/treatments/medications-for-memory>.
18. Cummings J, Lee G, Ritter A, Zhong K. Alzheimer's disease drug development pipeline: 2018. *Alzheimer's & Dementia: Translational Research & Clinical Interventions*. 2018;4:195-214.
19. Carmo Carreiras M, Mendes E, Jesus Perry M, Paula Francisco A, Marco-Contelles J. The multifactorial nature of Alzheimer's disease for developing potential therapeutics. *Current topics in medicinal chemistry*. 2013;13(15):1745-70.
20. Du X, Wang X, Geng M. Alzheimer's disease hypothesis and related therapies. *Translational neurodegeneration*. 2018;7(1):2.
21. Carrillo-Mora P, Luna R, Colín-Barenque L. Amyloid beta: multiple mechanisms of toxicity and only some protective effects? *Oxidative medicine and cellular longevity*. 2014;2014.
22. Hardy JA, Higgins GA. Alzheimer's disease: the amyloid cascade hypothesis. *Science*. 1992;256(5054):184.
23. Eckman CB, Eckman EA. An update on the amyloid hypothesis. *Neurologic clinics*. 2007;25(3):669-82.
24. Sanabria-Castro A, Alvarado-Echeverría I, Monge-Bonilla C. Molecular pathogenesis of Alzheimer's disease: an update. *Annals of neurosciences*. 2017;24(1):46-54.
25. Deane R, Bell R, Sagare A, Zlokovic B. Clearance of amyloid- $\beta$  peptide across the blood-brain barrier: implication for therapies in Alzheimer's disease. *CNS & Neurological Disorders-Drug Targets (Formerly Current Drug Targets-CNS & Neurological Disorders)*. 2009;8(1):16-30.
26. Šalković-Petrišić M. Amyloid cascade hypothesis: is it true for sporadic Alzheimer's disease. *Periodicum biologorum*. 2008;110(1):17-25.

27. Miyashita N, Straub JE, Thirumalai D, Sugita Y. Transmembrane structures of amyloid precursor protein dimer predicted by replica-exchange molecular dynamics simulations. *Journal of the American Chemical Society*. 2009;131(10):3438-9.
28. Kang J, Lemaire H-G, Unterbeck A, Salbaum JM, Masters CL, Grzeschik K-H, et al. The precursor of Alzheimer's disease amyloid A4 protein resembles a cell-surface receptor. *Nature*. 1987;325(6106):733.
29. Kametani F, Hasegawa M. Reconsideration of Amyloid Hypothesis and Tau Hypothesis in Alzheimer's Disease. *Frontiers in neuroscience*. 2018;12:25.
30. Kolata G. Down syndrome–Alzheimer's linked. *Science*. 1985.
31. Iqbal K, Liu F, Gong C-X. Tau and neurodegenerative disease: the story so far. *Nature Reviews Neurology*. 2016;12(1):15.
32. Kosik KS, Crandall JE, Mufson EJ, Neve RL. Tau in situ hybridization in normal and Alzheimer brain: localization in the somatodendritic compartment. *Annals of Neurology: Official Journal of the American Neurological Association and the Child Neurology Society*. 1989;26(3):352-61.
33. Rossi G, Dalprà L, Crosti F, Lissoni S, Sciacca FL, Catania M, et al. A new function of microtubule-associated protein tau: involvement in chromosome stability. *Cell cycle*. 2008;7(12):1788-94.
34. Johnson GV, Hartigan JA. Tau protein in normal and Alzheimer's disease brain: an update. *Journal of Alzheimer's disease*. 1999;1(4-5):329-51.
35. Lindwall G, Cole RD. Phosphorylation affects the ability of tau protein to promote microtubule assembly. *Journal of Biological Chemistry*. 1984;259(8):5301-5.
36. Alonso AdC, Zaidi T, Grundke-Iqbal I, Iqbal K. Role of abnormally phosphorylated tau in the breakdown of microtubules in Alzheimer disease. *Proceedings of the National Academy of Sciences*. 1994;91(12):5562-6.
37. Ren Y, Sahara N. Characteristics of tau oligomers. *Frontiers in neurology*. 2013;4:102.
38. Brion J-P. Neurofibrillary tangles and Alzheimer's disease. *European neurology*. 1998;40(3):130-40.
39. Khan UA, Liu L, Provenzano FA, Berman DE, Profaci CP, Sloan R, et al. Molecular drivers and cortical spread of lateral entorhinal cortex dysfunction in preclinical Alzheimer's disease. *Nature neuroscience*. 2014;17(2):304.

40. De Calignon A, Polydoro M, Suárez-Calvet M, William C, Adamowicz DH, Kopeikina KJ, et al. Propagation of tau pathology in a model of early Alzheimer's disease. *Neuron*. 2012;73(4):685-97.
41. Dickey CA, Dunmore J, Lu B, Wang J-W, Lee WC, Kamal A, et al. HSP induction mediates selective clearance of tau phosphorylated at proline-directed Ser/Thr sites but not KXGS (MARK) sites. *The FASEB journal*. 2006;20(6):753-5.
42. Khlistunova I, Biernat J, Wang Y, Pickhardt M, von Bergen M, Gazova Z, et al. Inducible expression of Tau repeat domain in cell models of tauopathy aggregation is toxic to cells but can be reversed by inhibitor drugs. *Journal of Biological Chemistry*. 2006;281(2):1205-14.
43. Davies P, Maloney A. Selective loss of central cholinergic neurons in Alzheimer's disease. *The Lancet*. 1976;308(8000):1403.
44. Perry E, Curtis M, Dick D, Candy J, Atack J, Bloxham C, et al. Cholinergic correlates of cognitive impairment in Parkinson's disease: comparisons with Alzheimer's disease. *Journal of Neurology, Neurosurgery & Psychiatry*. 1985;48(5):413-21.
45. Hasselmo ME, Anderson BP, Bower JM, editors. Cholinergic modulation may enhance cortical associative memory function. *Advances in neural information processing systems*; 1991.
46. Sarter M, Bruno JP. Cognitive functions of cortical acetylcholine: toward a unifying hypothesis. *Brain Research Reviews*. 1997;23(1-2):28-46.
47. Terry AV, Buccafusco J. The cholinergic hypothesis of age and Alzheimer's disease-related cognitive deficits: recent challenges and their implications for novel drug development. *Journal of Pharmacology and Experimental Therapeutics*. 2003;306(3):821-7.
48. Khan RA, Khan MR, Sahreen S. Brain antioxidant markers, cognitive performance and acetylcholinesterase activity of rats: efficiency of *Sonchus asper*. *Behavioral and Brain functions*. 2012;8(1):21.
49. Kamal Z, Ullah F, Ayaz M, Sadiq A, Ahmad S, Zeb A, et al. Anticholinesterase and antioxidant investigations of crude extracts, subsequent fractions, saponins and flavonoids of *atriplex laciniata* L.: potential effectiveness in Alzheimer's and other neurological disorders. *Biological research*. 2015;48(1):21.
50. De la Torre J, Mussivan T. Can disturbed brain microcirculation cause Alzheimer's disease? *Neurological research*. 1993;15(3):146-53.

51. Jack C. The vascular hypothesis of Alzheimer's disease: bench to bedside and beyond. *Neurodegenerative Diseases*. 2010;7(1-3):116-21.
52. Pristerà A, Sarauili D, Farioli-Vecchioli S, Strimpakos G, Costanzi M, di Certo MG, et al. Impact of N-tau on adult hippocampal neurogenesis, anxiety, and memory. *Neurobiology of aging*. 2013;34(11):2551-63.
53. Zhao Y, Gong C-X. From chronic cerebral hypoperfusion to Alzheimer-like brain pathology and neurodegeneration. *Cellular and molecular neurobiology*. 2015;35(1):101-10.
54. Ruitenbergh A, den Heijer T, Bakker SL, van Swieten JC, Koudstaal PJ, Hofman A, et al. Cerebral hypoperfusion and clinical onset of dementia: the Rotterdam Study. *Annals of Neurology: Official Journal of the American Neurological Association and the Child Neurology Society*. 2005;57(6):789-94.
55. Parsons CG, Danysz W, Dekundy A, Pulte I. Memantine and cholinesterase inhibitors: complementary mechanisms in the treatment of Alzheimer's disease. *Neurotoxicity research*. 2013;24(3):358-69.
56. Ballard C. Advances in the treatment of Alzheimer's disease: benefits of dual cholinesterase inhibition. *European neurology*. 2002;47(1):64-70.
57. Deng Y-H, Wang N-N, Zou Z-X, Zhang L, Xu K-P, Chen AF, et al. Multi-Target Screening and Experimental Validation of Natural Products from Selaginella Plants against Alzheimer's Disease. *Frontiers in pharmacology*. 2017;8:539.
58. Wollen KA. Alzheimer's disease: the pros and cons of pharmaceutical, nutritional, botanical, and stimulatory therapies, with a discussion of treatment strategies from the perspective of patients and practitioners. *Altern Med Rev*. 2010;15(3):223-44.
59. Thippeswamy B, Nagakannan P, Shivasharan B, Mahendran S, Veerapur V, Badami S. Protective effect of embelin from *Embelia ribes* Burm. against transient global ischemia-induced brain damage in rats. *Neurotoxicity research*. 2011;20(4):379.
60. Giacobini E. The cholinergic system in Alzheimer disease. *Progress in brain research*. 84: Elsevier; 1990. p. 321-32.
61. Kwon S-H, Lee H-K, Kim J-A, Hong S-I, Kim H-C, Jo T-H, et al. Neuroprotective effects of chlorogenic acid on scopolamine-induced amnesia via anti-acetylcholinesterase and anti-oxidative activities in mice. *European journal of pharmacology*. 2010;649(1-3):210-7.

62. Kim C-Y, Seo Y, Lee C, Park GH, Jang J-H. Neuroprotective Effect and Molecular Mechanism of [6]-Gingerol against Scopolamine-Induced Amnesia in C57BL/6 Mice. Evidence-Based Complementary and Alternative Medicine. 2018;2018.
63. Terry Jr A. Muscarinic receptor antagonists in rats animal models of cognitive impairment. Taylor & Francis Group, LLC, Boca Raton; 2006.
64. Kopelman MD. The cholinergic neurotransmitter system in human memory and dementia: a review. The Quarterly Journal of Experimental Psychology. 1986;38(4):535-73.
65. Malik J, Kaur J, Choudhary S. Standardized extract of *Lactuca sativa* Linn. and its fractions abrogates scopolamine-induced amnesia in mice: a possible cholinergic and antioxidant mechanism. Nutritional neuroscience. 2018;21(5):361-72.
66. Zlokovic BV. Neurovascular mechanisms of Alzheimer's neurodegeneration. Trends in neurosciences. 2005;28(4):202-8.
67. Iadecola C. The pathobiology of vascular dementia. Neuron. 2013;80(4):844-66.
68. Damodaran T, Hassan Z, Navaratnam V, Muzaimi M, Ng G, Müller CP, et al. Time course of motor and cognitive functions after chronic cerebral ischemia in rats. Behavioural brain research. 2014;275:252-8.
69. Farkas E, Luiten PG, Bari F. Permanent, bilateral common carotid artery occlusion in the rat: a model for chronic cerebral hypoperfusion-related neurodegenerative diseases. Brain research reviews. 2007;54(1):162-80.
70. Eleazu CO, Eleazu KC, Chukwuma S, Essien UN. Review of the mechanism of cell death resulting from streptozotocin challenge in experimental animals, its practical use and potential risk to humans. Journal of diabetes and metabolic disorders. 2013;12(1):60-.
71. Kamat PK. Streptozotocin induced Alzheimer's disease like changes and the underlying neural degeneration and regeneration mechanism. Neural regeneration research. 2015;10(7):1050-2.
72. Salkovic-Petrisic M, Knezovic A, Hoyer S, Riederer P. What have we learned from the streptozotocin-induced animal model of sporadic Alzheimer's disease, about the therapeutic strategies in Alzheimer's research. Journal of neural transmission. 2013;120(1):233-52.
73. Abbott NJ. Prediction of blood-brain barrier permeation in drug discovery from in vivo, in vitro and in silico models. Drug discovery today Technologies. 2004;1(4):407-16.

74. Upadhyay RK. Drug delivery systems, CNS protection, and the blood brain barrier. *BioMed research international*. 2014;2014:869269-.
75. Yusof SR, Avdeef A, Abbott NJ. In vitro porcine blood-brain barrier model for permeability studies: pCEL-X software pKa(FLUX) method for aqueous boundary layer correction and detailed data analysis. *European journal of pharmaceutical sciences : official journal of the European Federation for Pharmaceutical Sciences*. 2014;65:98-111.
76. Patabendige A, Abbott NJ. Primary porcine brain microvessel endothelial cell isolation and culture. *Current protocols in neuroscience*. 2014;69:3.27.1-17.
77. Iqbal K, Liu F, Gong C-X. Alzheimer disease therapeutics: focus on the disease and not just plaques and tangles. *Biochemical pharmacology*. 2014;88(4):631-9.
78. Gong C-X, Liu F, Iqbal K. Multifactorial Hypothesis and Multi-Targets for Alzheimer's Disease. *Journal of Alzheimer's Disease*. 2016(Preprint):1-11.
79. Balmus IM, Ciobica A, Negura A, Negura L, Anton E. BASIC ASPECTS IN SELECTING A SUITABLE TRANSGENIC RODENT MODEL FOR ALZHEIMER'S DISEASE. *Psychiatria Danubina*. 2015;27(4):0-345.
80. Liew K-F, Hanapi NA, Chan K-L, Yusof SR, Lee C-Y. Assessment of the Blood-Brain Barrier Permeability of Potential Neuroprotective Aurones in Parallel Artificial Membrane Permeability Assay and Porcine Brain Endothelial Cell Models. *Journal of pharmaceutical sciences*. 2017;106(2):502-10.